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Data Processing Methodolgies

U.S. International Biological Program

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DESERT BIOME US/IBP ANALYSIS OF ECOSYSTEMS

MODELS

RM 73-51

Data Processing

Data Processing Methodologies

US/IBP Analysis of Ecosystems

Desert Biome

Data Processing Methodologies

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September 1973

 $\chi(\sigma)$, $\gamma(\sigma) = \delta^{-1/2}$

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Table 1. Data Set Used to Illustrate Cluster Analysis. 3.2.3.-5

Preface

The intention of this report is to meaningfully relate computer data processing to the research of the ecologist working within the Desert Biome. To the "computer shy" ecologist it points out the advantages of automated data processing in his research. To the computer-using ecologist it suggests alternative modes of analysis and increased efficiency.

The computer analysis programs used by the Desert Biome Data Processing Group (DBDPG) represent a substantial resource both in program logic and personnel skilled in interpretation. As a package, they provide maximum information by means of analysis from many directions. These programs were acquired or written by the DBDPG as the need or anticipated need arose.

Programmers Alma Olsen and Kim Marshall deserve credit for programs written by the DBDPG. Brien Norton, Assistant to the Directorate, and programmer Verne King guided the formulation of the Data Set Abstract Search System. James MacMahon suggested that the species diversity program (SPECDIV) be written and provided the formulation of diversity indices.

Requests for analyses within the Desert Biome should be directed to:

> Charles Romesburg, Coordinator Data Processing Desert Biome Utah State University Logan, Utah 84322 (752-4100, Ext.: 7619)

For the interested reader outside the Desert Biome, requests will be honored for additional information pertaining to these programs as well as for acquisition of particular programs.

> Charles Romesburg, Coordinator Data Processing

September, 1973

1.0 Introduction

This report maintains that better ecological research results when the investigator records data in computer readable form (punched cards or magnetic tape). Research projects usually generate large amounts of data and the only reasonable mode of analyses is via the computer. This process leads to greater efficiency and increases the possibility of insight since data sets once established can be subject to alternative forms of analysis using existing programs.

For example, a project measuring plant cover generates a "raw" data set containing the cover dimensions by species, plot, and date. A computer program can then calculate percent cover from the individual cover dimensions to create a "reduced" data set. Both the raw and reduced data sets, stored on magnetic tape, can be analyzed from a number of different directions using computer analysis programs.

Section 2.0 describes procedures in establishing computer readable data sets. Kinds of data analyses (programs) a researcher may request are covered in Section 3.0. The identification of data sets stored in the Desert Biome data bank relevant to an investigator's needs may be made through the Data Set Abstract Search System, described in Section 4.0.

The DBDPG is to be regarded as a resource useful for consulting, execution of analyses, interpretation of computer printouts, and locating specific data stored in the data bank. Since DBDPG personnel are trained primarily in computer programming and analytical methods, the process of defining data sets, deciding on appropriate analyses, validating hypotheses, and, in general, finding insights, are the responsibilities of the the ecologist.

1.0.-1

2.0 Establishment of Computer Readable Data Sets

Getting raw data into computer readable form usually seems to the uninitiated more effort than the expected rewards are worth and, indeed, the process is more structured and hence more time consuming than entering data into field notebooks. After going through the process one or more times, however, it becomes much less of a chore. The steps needed to establish computer readable data sets are described in detail in the Desert Biome Report: Data Bank and Data Processing. The main steps, necessary to understand the relation to data set analysis, follow.

Data is entered into the data bank by recording data on a Data Set Coding Form, an example of which appears in Figure 1. Each row represents a punched card while the column headings identify where different information is to be placed within the punched card. These forms are completed by the DBDPG from information supplied by the investigator on a Data Set Description Form (not shown).

From the Data Set Coding Form with data transcribed, cards are punched. These cards are then entered as a data set into the data bank with the card images stored on magnetic tape. The data set created is referenced by a unique seven character Data Set Code (DSCode), and all requests for data set analyses necessarily include references to the DSCode.

A computer printout of the stored magnetic tape card images processed for readability is called a Data Set Listing. The Data Set Coding Form appearing in Figure 1, with data entered, generates the example Data Set Listing shown in Figure 2 (only the first page of the listing *is* shown.

Additional information describing experimental procedures used to generate a DSCode is provided by a Data Set Abstract. In the DBDPG all abstracts are stored on magnetic tape apart from the DSCodes they describe. Figure 3 shows the Data Set Abstract corresponding to the data set listing appearing in Figure 2.

 $2.0. -1$

Example of a Data Set Coding Form (Reduced)

Example of a Data Set Listing

Example of a Data Set Abstract

ASCUDES AND COULTO FORM NUMBERS A_3 STU1 A340 PRILIECT TITLE FVALUATION OF CHITICAL SOIL PROPERTIES NEEDED TO PREDICT SUIL-WATER FLOW WHILE HESIFT CUNDITIONS ABSIRACT IIILE SUTL WATER TRANSFER PROPERTIES **IRVESITGATING** DR. L. H. SIULZY (PRINCIPAL INVESTIGATOR) DEPL. OF SOIL SCIENCE, UNTVERSITY OF CALIFURNIA» KIVERSIDE, CALIFURNIA 92502 TELEPHONE $710 - 707 - 5112$ DK. J. LETET (PRINCIPAL INVESTIGATOR = 714-787-5116) G. H. MENUYS CRESEARCH ASSISTANT - 714-787-51131 GEOGRAPHIC INFIRMATION PUIN VALLEY SANIA HIIA **PARAMITERS** SUTE SUCTION COM OF WATER), VULUMETRIC WATER CUNTENT (CM**3/CM**3), HYDRAULIC CUND'ICTIVITY (HRZCM), SUIL=WATER DIFFUSIVITY (HRZCM**2) IIME OF SAMPLING (SUIL SAMPLES TAKEN IN MARCH, MAY, AND NUVEMBER 1972) 01 MAR 72 **EXPERIMENIAL** METHUUS. TRANSIENT OUTFLOW METHOL FOR EVALUATION OF HYDRAULIC CONDUCTIVITY AND SUIL-WATER UIFFUSIVITY (WEEKS) ET AL., 1967). SOIL SUCTIUN MEASURED WITH TENSIOMETERS AND PSYCHRUMETERS. FXPERIMENTAL UFSIGN RUTH UNUISTURBED AND LUUSE SAMPLES WERE TAKEN AT THE SANTA RITA. TUCSUN SITE. UNDISTURBED CURES WERE 10 CM IN DIAMETER BY 30 CM LUNG. LUOSE SAMPLES WERE TAKEN TO THE 1.20 M DEPTH AT EVERY 30 CM INTERVAL. BECAILST OF THE EXTREME STUNINESS OF THE RUCH VALLEY, NEVADA SOIL, ONLY LOGSE SAMPLES WERE CULLECTED, TO A DEPTH OF 50 CM. LUCATIONS FUR SAMPLING WERE CHOSEN AT RAMDUM WITHIN EACH VALIDATION SITE. SUIL CULUMNS 10 CM 1606 BY 30 CM LONG, EITHER RECONSTITUTED IN THE LAB FROM LULISE SAMPLES UR TAKEN IN SITU, WERE USED THROUGHOUT. CITATIONS WEFRS, L. V., AND S. J. PICHARDS. 1967. SUIL WATER PROPERTIES CUMPUTED ERUM TRANSIENT FLOW DATA. SOIL SCE. SUC. AMER. PROC. $31 - 121 = 725.$ SUPPORTING POCUMENTS PROPUSAL 1971: 5.6.0.-1 RESEARCH MEMURANDUM 73-43

In general, data set processing requires that either a specific program be written or the use of a pre-written "canned" program. This report emphasizes the latter for the reason that there is much less to be said about specific programs. In fact, specific programs constitute a substantial portion of the programming within the DBDPG.

The questions of what data to collect and how the data should be analyzed are interrelated, a choice for one constraining the possible choices for the other. The intention in presenting computer analysis techniques is not to suggest that data should be collected in such a way that these techniques can be used but to suggest new directions of approach of which the investigator might be unaware.

Each technique is minimally described by what it will accomplish. Where it is felt understanding will benefit, an example is given accompanied by an illustration of the computer output. Each program is given a name followed by a reference to its origin.

Section 3.1 covers the process of submitting requests for analysis. Section 3.2 presents data display techniques where statistical inference to a larger parent population is infeasible. Programs for commonly occurring problems in making statistical inferences are given in Section 3.3. Other programs available for standard statistical analysis, numerical analysis, and optimization problems are listed in Section 3.4.

Because geographic location often prohibits direct contact between investigator and the DBDPG it is important that written instructions for analysis be complete. There are several general rules which lead to good communication. The request should include text explaining what should be calculated and the DSCodes from which the raw data are obtained. Within each DSCode the data fields applying to the calculation should be specified by column numbers. The request should procede in clear, step-wise fashion and if possible, the calculations should include a numerical example. Figure 4 is regarded as a clearly presented request for a specific program analysis.

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Example of Written Request for Program Analysis

TO: Charles Romesburg Desert Bi ome Utah State University Logan, Utah 84322

FROM: Russell P. Balda Northern Arizona University Flagstaff, Arizona

Please find enclosed a total of 14 coding forms, #17. The first 9 are for trapping period 9, the last 5 for period 10. Data will always be analyzed by trapping period so it is important to keep the periods separate.

Before we spend any more time preparing our raw data and transposing it to the coding forms, we would like to analyze this data to make certain we are doing everything correctly. Please note that we used two spaces from the TAXON CODE fields (24 & 25, 36 & 37, 48 & 49, 60 & 61) for a new entry, i.e. frequency. Also note that the plant data for some mammal capture-sites will
not fit on one card but will need to be placed on two cards. Where a second card is necessary, the first 18 fields are left blank, meaning the information in the remaining fields goes with the capture-site immediately above. This is necessary because the number of plant species per capture-site is highly variable.

I hope your staff will be able to put these data on cards, write the appropriate program, and then run the analysis for us. The biggest problem will be my written directions, but I will try to be as clear as possible. We would like the data to be analyzed in a number of different ways.

- Part A: For each capture-site, we would like a print-out showing relative density, relative frequency, relative dominance, the sum of the above three, percent dominance for the herbaceous plants, and percent dominance for the overstory. This analysis is to be done as follows for the capture-site of each mammal. On all plant species with a taxon code number from 0001 to 0299 do the following:
	- 1. Relative density = <u>Total density of each species</u> x 100

	Total density of all species x 100

For example, on sheet one the first capture-site has five plant species with densities of 7,1,1,3,l. The total density is 13. Thus, the relative density of each plant species is as follows:

2. Relative dominance = Total dominance of a species **Total dominance of a species** x 100

All measurements in the dominance categories for plants with taxon codes between 0001 and 0299 are in hundredths of feet so the decimal is always 0.00. For example, again using the first capture-site on sheet one, the five plant species had dominances of 3. 09 ft. , 0. 53 ft. , 0. 22 ft. , 0. 98 ft. , O. 12 ft. This totals to 4. 94 ft.

3. Relative Frequency = Frequency of a species Frequency of a species x 100
Frequency of all species

Using the data from the first capture-site on sheet one, the relative frequency is calculated as follows. The frequency values for the five plant species total to 12, therefore:

4. Importance Value is the sum of 1., 2., and 3., above; for example: For the five plant species at the capture-site used above.

> 0046 - 53.84 + 62.55 + 58.33 = 174.72
0021 - 7.69 + 10.72 + 8.33 = 26.74
0013 - 7.69 + 4.45 + 8.33 = 20.47
0047 - 23.07 + 19.83 + 16.66 = 59.56 0041 - 7.69 + 2.42 + 8.33 = 18.44 $0046 - 53.84 + 62.55 + 58.33 = 174.72$ $7.69 + 4.45 + 8.33 = 20.47$ $0047 - 23.07 + 19.83 + 16.66 = 59.56$

5. Percent herbaceous dominance is calculated as follows:

% Herbaceous Dominance= Total dominance for all species X 100 **11** .00

The total dominance figure is the same one used in AZ. above. In the example it is 4.94 ft. The 11 is a constant; it is the maximum number of feet that could be covered with vegetation. Thus the percent herbaceous dominance for our example is:

$$
\frac{4.94}{11.00} \times 100 = 44.90\%
$$

6. Percent overstory dominance is to be calculated only for plants with taxon codes from 0300 to 0599. This is the only calculation in which these plants are used. Also, the dominance field for these plants is the only one with entries. For an example, check the second capture-site as the first one has no overstory plants present.

The 21 is the maximum number of feet that could be covered by these plants.

09 - PERBAI - CF10 - 1

The printout for Part A., as calculated above, should read something like the example given below for the first capture-site on Page 1. Each capture-site should be listed in succession as on the coding forms.

percent herbaceous dominance = $44.90%$ percent overstory dominance = 00.00%

Part B. For each species of mammal we need the above information totaled by habitat (1 or 2; not 3 in this data set) and sampling period. For example, we would like these data for all PERBAI during sampling period 9 in habitat 1. The calculations will be similar to those done above.

1. Relative Density = $Total$ density of each plant species at all PERBAI \times 100 Total density of all plant species at all PERBAI

Sec.

 $\sim \infty$ $_{\rm{eq}}$

- 2. Relative Dominance = Total dominance of each plant species at all PERBAI x 100 Total dominance of all plant species at all PERBAI
- 3. Relative Frequency = Total frequency of each species at all PERBAI X 100

Total frequency of all species at all PERBAI
- 4. Importance Value is the same as done in Part A.; the sum of l., 2., and 3., above.
- 5. Percent herbaceous dominance is calculated differently since, rather than use 11, we must multiply 11 by the number of capture-sites before dividing by the total dominance figure as calculated in 82. For example, if there are 10 PERBAI, then percent herbaceous dominance is:

Total dominance of all plant species at all PERBAI $\begin{array}{cc} \chi & 100 \end{array}$

- 6. We also need to know additional information about the absolute dominance provided by each species per species of mammal. This is an additional calculation not performed in Part A. Simply use the total density of each plant species at all PERBAI as was done for Bl., and total dominance figures for each plant species as was done for 82., and simply print it out (see below).
- 7. Percent oversotry dominance is calculated as explained in A6, with the modifications discussed in 86. It is done only on plants with taxon codes from 0300 to 0599. However, rather than divide by 21, we must multiply 21 by the number of capture-site
then divide by the total diminance figure of all these plants.
- 8. We need to know how much overstory was contributed by each different species of plant with a taxon code between 0300 and 0599. This is done by adding up the dominance figures for each species of plant separately and then dividing by 21 times the number of capture-sites. The total of these percentages for each species will sum to the figure obtained in 87.

The print out for Part B should read as follows for each species of mammal:

09 - PERBAI -

The above analysis is to be done for each species of mammal by habitat type. For example, DIPMER occurs in both habitat 1., and 2. The analysis needs to be done separately for each habitat. Please note that habitat 3 was not represented in any of the data submitted this time. In the future, however, we will be submitting data with capture-sites in 3. When this occurs we will want analysis of all three habitats separately and then habitats 2 and 3 combined as if they were the same.

- Part C. For each habitat studied, (in this case 1 and 2), we need <u>exactly</u> the same data as was calculated in Part B. Everything is to be done exactly the same as for Part B, but this time the species of mammal can be ignored. In B5, B7, and B8, where number of capture-sites by species was multiplied by either 11 or 21, this time simply substitute number of capture-sites of all mammal species per habitat. Thus, the print out will be in two sections, one for habitat 1, and one for habitat 2. If the animals had been captured in habitat 3, then the print out for Part C would need to be in four sections: One section for habitat 1, one for habitat 2, one for habitat 3, and one for habitat 2 and 3 combined. The print out for each of these sections should follow the format shown for Part 8 except for the headings which should show sampling period and habitat. The four headings for the data included here would be:
	- 09 ALL species 1 09 ALL species - 2 10 - ALL species - 1 10 - ALL species - 2

This is all the analysis we can envision for these data and all others to be sent at a later date on coding form #17. However, it would be wise to put these data on magnetic tape because we will most likely want other information as we proceed with the project.

I realize the problem of trying to write a compact program for these data based on my written instructions. When problems arise please call and we can work them out verbally, as I realize my shortcomings in trying to convey my instructions by mail.

If possible, after the cards are punched we would appreciate having the coding forms back for our reference. Thank you for your help and cooperation in this matter.

3.2 Non-inferential Data Analyses

3.2.1 Data Sorts and Sunnnaries

Often it is desirable to rearrange and/or summarize raw data. By example, data are usually recorded on the Data Set Coding Form by sequential date of collection, site of observation, taxon, replicate, observer, etc. It may be more meaningful, however, to list the data by a different hierarchy, e.g., perhaps by taxon first followed by replicate, date, site, and observer. In addition, it may be useful to count the number of occurrences of each taxon, that is, summarize the raw data. Rearrangements and summaries of large data sets are all but impossible without using the computer.

The DBDPG has created a series of programs to accomplish this under the name SORT. The procedure is illustrated for a data set which is in part shown in Figure 5. These data were summarized by counting the occurrences of each taxon by size class, with the output appearing in Figure 6.

Portion of Data Set Used to Illustrate SORT Analysis

Example Output from SORT Analysis

3.2.2 Visual Data Display Analyses

Two of the most used modes of visual data display in science are histograms and "curve drawing". Two programs have been written by the DBDPG to automate this display process. Histograms are presented by program HIST which computes the frequency of occurrence for attributes of interest by class interval (Figure 7). Program GRAPH plots points for x-y variables and although automated point connection is not provided this is easily accomplished by hand (Figure 8). For both programs, scaling of axes is optionally input by the user or computed automatically.

Example Histogram Drawn by HIST Program

THIS IS THE INITIAL AGE DISTRIBUTION:

PERCENT OF TOTAL SEED IN EACH SEED COHORT

 $3.2.2. - 2$

 \vee

 $m \leq C \Gamma C$

 $\boldsymbol{\%}$ M

DISTSTS

Figure 8

 $3.2.2.2. - 3$

3.2.3 Classification and Ordination Analyses

Classification and ordination techniques are useful for revealing fundamental underlying structure of multivariate data. The data basic to each consists of measurements made for a set of attributes across a set of "individuals". An individual is the entity the measurement is made on while an attribute is what is measured. The problem is to understand the relation among individuals given the attribute measurements for each. Most frequently attention is directed to finding those individuals which are either alike or dissimilar.

The data are usually arranged in matrix form with individuals being the columns and attributes the rows. A given column-row entry is a datum giving information on the attribute for the individual. For example, individuals might be taxa and attributes measurements on taxa such as biomass, size, etc., or individuals could be sites with attributes specified by measurements on taxa found there. In the first case interest might be focused on distinguishing among taxon and in the second case among sites.

Classification is concerned with the ordering of individuals into groups on the basis of their attribute relationship. Ordination techniques are used to reduce the dimensionality of the Attribute space by replacing the attribute set with a new and smaller set; often the reduced space allows for relations among individuals to be identified. In general, the basic data matrix can be subject to both classification and ordination treatment, and this is the usual procedure. The results of multivariate data analysis are seldom black and white but subtle shades of gray. It is usually beneficial to treat the problem from several directions and integrate the results to arrive at a conclusion.

The multivariate programs used within the DBDPG are:

Ordination

Only the classification programs are discussed here and illustrated by example although a complete analysis would include running the ordination programs.

Brief descriptions of the classification programs follow:

MINF0 (Goldstein and Grigal, 1972)

This method considers each individual as a separate group at the start of the algorithm. During each clustering cycle, a pair of groups is joined which results in the minimum increase in mutual information, where information is in accordance with Shannon's (1949) definition. The clustered pair then becomes a new group (cluster) and the process continues until all individuals are contained within a single group.

MDISP (Godstein and Grigal, 1972)

This method is similar to MINFO except that clustering occurs for that pair of groups which provide the minimum increase in within-group dispersion.

MINT (Rohlf, 1971)

This program has a number of user options for "similarity coefficients" and "clustering methods". Similarity coefficients define alikeness among individuals or groups while the various clutering methods represent choices for forming clusters, i.e., discriminating for alikeness. The options follow Sokal and Sneath (1963, 1973):

Similarity Coefficient (interval data)

- 1. correlation coefficient
- 2. average Euclidean distance
- 3. average Manhattan distance
- 4. variance-convariance matrix

Similarity Coefficient (qualitative data)

- 1. simple matching coefficient
- 2. Jaccard coefficient
- 3. Dice coefficient
- 4. Yule coefficient

Clustering Methods

- 1. weighted pair-group method using arithmetic averages
- 2. unweighted pair-group method using arithmetic averages
- 3. single linkage
- 4. complete linkage
- 5. centroid linkage
- 6. flexible linkage

The provision for qualitative data is useful when it is desirable or of necessity to measure attributes on a present-absence basis.

An example problem using the MINT program with 10 individuals and 47 attributes illustrates cluster analysis. The individuals in this case are streams while the attributes are the presenceabsence (coded as 1 or 0, respectively) of aquatic invertebrate taxa. The data matrix is shown in Table 1. The simple matching coefficient of similarity and the single linkage method of clustering is employed. The output is shown in the form of a dendrogram, or tree diagram, in Figure 9. Horizontal lines in the tree represent nodes where diverse subgroups are joined. Heterogeneity is greatest

at the bottom of the tree where each group is separate, consisting of one individual. At the top of the tree there is a single group containing all individuals. As one procedes from the bottom to the top of the tree individuals will group to form clusters. A measure of similarity of clustering groups is the distance from the bottom of the tree to the node of the clustering group. The smaller this distance the more alike are the members clustering. Thus, in Figure 9 individuals 1 and 3 cluster first and are considered more alike than, say, the cluster of group 1,3,9 with individual 7.

 $\overline{\overline{E}}$

 ∞ Analys Cluster ble 1
lustrate Set Used to Data

Dendrograms Drawn by MINT Program

3.2.4 Synagraphic Mapping

The data for synagraphic mapping consists of pairs of observations in the order of: a) quantity measured; b) location of measurement. From these data the measured values can be plotted on a map of the site from which they were obtained. Synagraphic mapping is a technique for projecting the continuous spatial distribution of the measurements based upon the sample measurements. Typical measurements as a function of location are plant cover, numbers, biomass, and microclimatological data.

These maps are produced by the SYMAP* program with the output appearing as a shaded map by contrast printing. The darkest areas denote highest measurement values while the least darkest denote the lowest. There are several user options for generating the continuous spatial surface by interpolation between values at given data points and for drawing surface contour lines.

Figure 10 illustrates the contour map option of SYMAP generated by recording at random locations a measured value, in this case, soil surface temperature. Temperature is recorded as falling into one of four discrete classes numbered from 1 to 4, e.g., $51 - 60$ °F, $61 - 70$ °F, etc., and the locations and values sampled are superimposed on Figure 10 according to this numbering scheme.

The synagraphic mapping program SYMAP was originally written by Howard T. Fisher at Northwestern Technological University in 1963. The version used here was purchased from the Harvard University Laboratory for Computer Graphics and then made compatible with the Burroughs BL 6700 computer.

Conformal Map Drawn by SYMAP Program (Reduced)

j

 $3.2.4.-2$

3.2.5 Species Diversity Programs

The SPECDIV program written by the DBDPG, takes as input a list of the number of individuals obtained by sampling for each of two or more species. From this the frequently discussed measures of species diversity are calculated.

The notation used is:

 $\mathbf{n_i}$ = Number of individuals of the i'th specie

 $N =$ Number of species in collection

I = Number of individuals in collection (I = N \sum_{i} n₁ $i=1$ $\frac{1}{i}$

Several symbols used to represent diversity:

D (subscripted) H H' (H-prime)

For each diversity index D the originator's name is parenthetically enclosed. The references from which they were obtained is given at the end of this section.

Species Diversity Measures

Dl (Origin unknown)

Dl=N

That is, the number of species is a measure of diversity without regard to the number of individuals or their distribution.

It is possible to compute lower and upper bounds on Dll. Denoting these by LB and UB:

 $\mathcal{R}^{(n)}(t)$. We can be seen to be a second contribution of $\mathcal{R}^{(n)}$

$$
LB = I/\sqrt{N}
$$

$$
UB = \sqrt{[I - (N-1)]^{2} + (N-1)}
$$

Therefore, LB \le D11 \le UB

D12 (Margalef, 1957)

Margalef partitions the quantity of information into:

- 1) Count the number of species
- 2) Distribute the individuals according to species
- 3) Localize the species
- 4) Localize the species of equal frequency (hypothetical situation)
- 5) Localize the individuals (maximum information)

These are denoted by $D(1)$, $D(2)$, $D(3)$, $D(4)$, $D(5)$ and are given by the following formulas:

The constant 1.443 converts the information into "bits". The above express the total information. This is given on a per individual basis by dividing each index by the number of individuals I. These are denoted:

 $D(1)/I$, $D(2)/I$, etc.

 $D(5) = 1.443 log_e I!$

The interested user should consult Margalef's paper.

Dl3 (Fisher, 1943)

D13 is given by α in the following transcendental equation:

$$
N = \alpha \log_e \left(1 + \frac{1}{\alpha}\right)
$$

D15 (Hurlbert, 1971)

D15 =
$$
\begin{bmatrix} I \\ I-1 \end{bmatrix} \begin{bmatrix} \frac{N}{1} - \sum_{i=1}^{N} (\frac{n_i}{I})^2 \end{bmatrix}
$$

H, HMAX, J (Pielou, 1969)

His the uncertainty measure appropriate to a finite population.

$$
H = \frac{1}{I} \log_e \frac{I!}{n_1! \ n_2! \ \cdots \ n_N!}
$$

HMAX is the maximum diversity a collection of I individuals can have. It is calculated by assuming all individuals are equally distributed in number among all N species.

×.

$$
HMAX = \frac{1}{I} \log_{e} \frac{I!}{\sqrt{\left[\frac{I}{N}\right]! \sqrt{\left(\frac{I}{N}\right)! + 1}}}
$$
\n
$$
W = I = N \left[\frac{I}{N}\right] + r
$$

and [·] means the integer part of the argument

$$
J = \frac{H}{HMAX}
$$
 is a measure of eveness of distribution.

0 < J < 1.0 *l l* uneven even

HPRIME, HPRIMEMAX, JPRIME (Pielou, 1969)

The index HPRIME corresponds to the diversity of the population from which samples are obtained. It assumes knowledge of the true population proportion of the jth species, p_j . In practice p_j is often estimated by (n_j/I) . Use of the estimated value for p_j in the formula results, however, in a biased estimate of HPRIME (Pielou, 1969). HPRIME is included in SPECDIV because incorrect usage persists.

HPRIME =
$$
-\sum_{j=1}^{N} \frac{n_j}{1} \cdot \log_e \frac{n_j}{1}
$$

\nHPRIMENTAX = log_e (N)
\nJPRIME = $\frac{HPRIME}{HPRIMEMAX}$

The following references were used in the development of SPECDIV: Auclair and Goff, 1971 - Dl, D2, D3, D4, D5, D6, D7, D8, D9, D10, D11 Margolef, 1957 - D12 Fisher, et. al., 1943 - D13 $Hurlbert$, 1971 - D15

An example problem used with SPECDIV is shown in Figure 11.

Example Problem Using SPECDIV Program

Problem Input

 $N = 9$

Problem Output

```
01 = 9.00000E+00000D2(WILLIS, 1922)=
D3(GLEASON, 1922)= 1.39153L+00000
                                       D4(PRESTON, 1948) =
D5(MARGALEF, 1957)= 1.23692E+00000
                                       D7(MENHINICK,1964)= 3.39723E=00001
D8(MENHINICK,1964) = 3.54050E=00001
                                       D9(MINK, 1966) = 1.39752E = 00002013(FISHER, 1943)= 1.48091E=00000
                                       D10(SIMPSON*1949) = 1.29711E=0000106(MACARTHUR.1957)= 2.11004E+00000
015(HURLBERT, 19/1)= 8,71642E=00001D11(MCINTUSH/1907) = 2.31940L+00002LOWER BOUND 2.1466/E+00002 UPPER BOUND 6.36006E+00002
D12(MARGALEF.1957)
0(1) = 9.33289E*00000D(2) = 5.92950E*000010(3) = 1.92782E+00003D(4) = 2.00819E+000030(5) = 5.08708E+00003
0(1) / I = 1.44921E=00002
0(2) / I = 9.20/30E=00002
0(3) / I = 2.99351E*00000D(4) / I = 3.11031E+000000(5) / I = 7.89919E+00000
H = 2.07450E+00000HMAX = 2.16097E*00000J = 9.59988E = 00001HPRIME = 2.11004E+00000HPRIMENT = 2.19722E*00000JPRIME = 9.60323E-00001HPRIMELOG2 = 3.04415E+00000HPRIMEMAXLUG2 = 3.16993E+00000JPRIMEL0G2 = 9.60323E=00001
```
 $\tilde{V}^{(n)}$, the contract of $\tilde{S}^{(n)}$, where $\tilde{S}^{(n)}$ is a contraction of

3.2.6 Random Field Plot Locater

Program RANDPLØ, written by the DBDPG, locates points within a geographical site for random quadrat placement. The site is stratified into subregions with the number of locations by subregion specified by the investigator. Random point locations are subsequently determined by RANDPL \emptyset for sampling within each subregion. These are printed out in the form of a map superimposed upon an x-y coordinate system.

3.2.7 Special Programs

Often a standard data analysis is modified in some way by an investigator for his own needs. Some of the more important of these programs written by the DBDPG are:

Rodent Pouch and Stomach Content Summary Emlen Census Analysis Fish Life Table Analysis Rainfall Probability Analysis

With slight modification these programs can be used for data with similar analysis objectives.

3.3.1 Elementary Statistics

Elementary statistical measures are provided by the STATS program, written by the DBDPG. Given below are descriptive statistics and statistics useful for infering to a larger population on the basis of a sample.

Descriptive Statistics:

The following are printed or computed from the sample:

Number of observations Mean Variance Standard deviation Standard error Coefficient of variation

Population Statistics:

Based upon the t-statistic, both 90 and 95 percent confidence intervals on the true population means are given. For cases where the assumption of normality is suspect, 90 and 95 percent confidence intervals based upon the distribution-free Tchebycheff inequality are also computed.

STATS estimates requirements for future sampling based on the sample variance (normality is assumed). For α levels of 0.9 and 0.95 an estimated sample size is computed such that the sample mean differs from the population mean by not more than a given fraction R of the population standard deviation (the bample s.d. is used to estimate the population s.d.). Example STATS output is shown in Figure 12.

Example of STATS Program Output

START 12 LENGTH 5 DECIMAL 10 NUMBER OF CBS. 12_b \equiv MEAN 43.424 \overline{a} VARTANCE $\frac{d\mathbf{w}}{d\mathbf{w}}$ **U305.744** STANDARD DEVIALSON 19.786 $\frac{1}{1600}$ STANDARD ENRUR 7.052 $\frac{1}{2}$ $CUEF$. OF VARIALION = 103.710 THE 90 AND 95 & CUNFLOENCE INTERVALS RASED ON THE T STATISTIC ALPHA LUNTH SHOW UPPER DININI $90₉$ 31.93 55.020 95.1 24.607 = 51.251 THE QU AND 95 9 CUNFIDENCE INTERVALS DASED LIV TCHERYCHEFF INEWPALITY ALPHA LUWER BOULD UPPER BUUND $c1.1c1 =$ $90x$ 65.730 959 $11.891 74.901$ ESTIMATED SAMPLE SIZE SUCH THAT WITH A GIVEN PRUBABILITY CEXPRESSED BY THE ALMHA LEVELD THE SAMPLE NEAN DIFFERS. FROM THE POPULATION NEAN OY JOT MORE THAN A GIVEN FRACTION R.OF THE OF THE HOPOLATION STANDARD DEVIATION. NURMALITY IS ASSIGINU 904 MIN. UDSCH. H 3636 0.05 Q_A U .lü 005 $U_0 15$ $U \circ c$ $2c$ 146 0.25 1_U1 0.30 95 % MIN. IBSER. \mathbf{r} 5106 $U_0 U_2$ 1796 U_0 10 516 0.15 324 $U_0 Z U$ 201 0.25 144 0.30

3.3.2 Multi-way Contingency Table Analysis

Count data are often analyzed in contingency table form using the Chi-Square test statistic. Classification of the observed sample in this case is limited to two criteria. For example, rodents trapped can be classified by age and sex, sex and trap type, age and species, etc. When the data are classified by more than two criteria a multi-way contingency table is created. In the above example the data could be used in a four-way table listing age, sex, trap type, and species criteria.

The proper G test statistic for multi-way contingency analysis is based on information theory as developed by Kullback (1959). (Sokal and Rohlf [1969) give a short discussion and an example illustrating application.) Interpretation of a multi-way analysis requires detailed theoretical understanding, and the user is advised to consult a knowledgeable statistician to ensure proper use.

The MULTITAB program was written by the DBDPG for multi-way contingency analysis. Contingency tables up to five-way can be handled. A series of hypothesis tests are possible; for example, a three-way table with criteria A,B,C will allow the following tests:

It should be noted that lack of independence among n criteria does not imply independence among (n-1)-way, (n-2)-way, etc. classifications.

Example Program Output:

The following hypothetical data represents classification of

rodents trapped according to 3 species, 2 trap types, 2 ages, and 2 sexes.

A portion of the computer output is shown in Figure 13. Under "Source" is the criteria being tested, under "DF" is degrees of freedom, and under "G" is the value of the G statistic. The G value is to be compared with a critical chi-square value with appropriate degrees of freedcm. A, B, C, D represent species, trap type, age, sex respectively. From the analysis, the hypothesis of four-way independence is rejected. At the three-way level AXBXC independence and AXB and AXC two-way independence are rejected. The four-way interaction term corresponds roughly in meaning to that given by a four-way Analysis of Variance; it is not rejected.

Example of MULTITAB Program Output

ANALYSIS OF INFURMATION

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3.4 General Statistical Numerical Analysis and Optimization Programs

The DBDPG maintains a library of programs useful in statistical, numerical analysis, and optimization problems. At present, over 150 programs have been acquired, and their use and interpretation is understood by DBDPG personnel. This resource is drawn upon as needed in order to analyze given problems from diverse directions. Statistical routines are useful in analyzing data, numerical analysis is useful for both data and modelling activities, and optimization programs exist for the projected trend of Desert Biome research in desert resource management decision models. A partial list of these programs follows.

Statistical

ANOVA

Regression and Correlation Analysis Least Squares Fit by Orthogonal Polynomials Curve Fitting with Constraints Factor Analysis Principal Components Analysis Discriminant Function Analysis Probability Similarity Index Normal Random Deviates

Numerical Analysis

Numerical Integration Polynomial Equation Roots Complex and Real Exponential Integral Matrix Operations Interpolation by Aitken Numerical Differentiation Fibonacci Search

Optimization

Linear Programming Goal Programming Quadratic Programming General Non-Linear Programming Critical Path Scheduling Integer Programming Least Cost Network Flow Minimal Spanning Tree

4.0 Data Set Abstract Search System

At the end of 1972 the Desert Biome Data Bank contained over 450 data sets. As individual data sets become less manageable with increased size so, too, do collections of data sets. The DBDPG views the data set collection as one large data set under the management control of a computer information retrieval system. This system, named the Data Set Abstract Search System, provides information, in the form of data set abstracts (illustrative abstract shown in Figure 3), describing data covered.

Within the Desert Biome, data sets are available to investigators for use in modelling, ecosystem comparison and syntheses, etc. The retrieval program selects a set of DSCodes of interest to the user. This is accomplished by specifying as input one or more "profile elements," selected from a Data Set Search Profile List, which are relevant to the user's interest. Each DSCode is associated with a "relevant profile element" set. The retrieval program selects DSCodes by comparing the set of "profile elements" specified by the user with the "relevant profile set" of each DSCode (retrieval programs of this principle are commonly known as Keyword Systems).

To use the program the user selects appropriate numbers representing elements from the Profile sheet. For an abstract to qualify for retrieval the "relevant profile set" must contain all of the selected elements. For example, if the user specifies "shrubs" and "Curlew Valley" only abstracts for data sets applying to shrubs in Curlew Valley will be retrieved.

Instructions for abstract retrieval requests should be requested from:

> Steve Black Central Office Assistant Desert Biome Utah State University Logan, Utah 84322

When the user receives these instructions, the request form should be filled out and returned to the same address.

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