Abstract
Microgravity adversely affects several physiological systems. As space exploration moves to extended flights, understanding which mechanisms microgravity influences may contribute to the health and safety of astronauts aboard. Maximum oxygen consumption (VO$_2$ max) testing is used to monitor changes to cardiorespiratory function. Currently the equipment used to such testing aboard the International Space Station (ISS) is large and requires a significant amount of power. We propose the use of a modified metabolic management system, currently approved for clinical use, for exercise stress testing.

I. INTRODUCTION
The absence of gravity in space adversely affects several physiological systems of the body including calcium absorption resulting in bone density loss, cardiovascular deconditioning, muscular atrophy, and neurovestibular function. Upon returning to the Earth’s gravitation field some of these conditions are only partially reversible. Previous missions have demonstrated that the hazards associated with such an unforgiving environment often require maximum cognitive performance and physical fitness for survival. As space exploration progresses from short-duration flights to long term residency, minimizing the effects of weightlessness on the body will be of increasing importance for the safety of crew members for both inflight well-being and postflight recovery.  

A combination of aerobic and resistance exercise regimen protocols have been suggested as a possible countermeasure against detrimental physical deconditioning due to microgravity. Since the Skylab missions, periodic stress tests, which have the advantage of being noninvasive, were administered to crew members as a way of assessing the health status of multiple major body systems. The most commonly measured parameter during exercise stress tests is maximum oxygen uptake (VO$_2$ max). During exercise the cardiovascular system must function optimally to provide systemic oxygen transport and utilization. At some point during maximal exercise the linear relationship between O$_2$ consumption and mechanical power plateaus. This point, known as VO$_2$ max, is considered the best indicator of cardiorespiratory endurance and aerobic fitness. Consequently it can be used to quantify and track cardiovascular deconditioning.

To determine the VO$_2$ max, respiratory gas flow and the concentration of inspired and expired oxygen and carbon dioxide must be recorded simultaneously while the subject is performing a specific exercise protocol. Aboard the space station, the standard protocol, which consisted of regular increases in the speed and intensity of exercise, is performed via bicycle ergometer (Figure 1). Since the rate of oxygen uptake directly correlates with the rate of metabolic energy expenditure, this metabolic gas analysis system can also be used to determine nutritional requirements for crew members. This process is known as indirect calorimetry and is the primary clinical method for assessing metabolic rate.

![Figure 1](image_url) Officer Jeff Williams assists Flight Engineer Thomas Reiter perform a VO$_2$ max test during ISS Expedition 13
The metabolic gas analyzer, GASMAP, integrated into the human research facility (HRF) aboard the International Space Station (ISS) system is large (one 1 x 8 panel unit drawer and two 1x4 panel unit drawers, 51 Kg total) and requires a significant amount of power (205 watts), Figure 2. The prototype metabolic measurement system was developed as research project at the University of Utah. It was refined and delivered to Johnson Space Center in 1988.\textsuperscript{15} The system was self-calibrating and offered many advantages over earlier metabolic measurement systems.

During the past 3 years, our group at the University of Utah has worked in collaboration with a division of Philips Electronics and others to develop a metabolic measurement system for use in anesthesia and intensive care. This system, shown in Figure 3, is much smaller than the GASMAP device (less than 5 in\textsuperscript{3}) weights less than 0.5 Kg and requires less than 2 watts. We have demonstrated the accuracy and robustness of this device in a patient simulator, in human volunteers and in the intensive care unit (ICU).

The main limitation of the present system is that it is not capable of measuring oxygen uptake during exercise conditions with larger gas flows. We propose that by modifying the breathing apparatus and increasing the lumen diameter we could build a prototype version of the critical care metabolic gas analyzer that would be suitable for exercise testing.

\textbf{II. SENSOR DESIGN CONSIDERATIONS}

Because our system uses a differential pressure type flow sensor, it requires a compromise between the resistance to flow experienced by the user, and the magnitude of the differential pressure signal from which flow is measured. On one hand, a large differential pressure in response to gas flow gives a stronger signal relative to noise; on the other hand, a resistive sensor restricts gas flow inhibiting the subjects breathing leading to a possible falsely low VO\textsubscript{2} max readings. It is critical that the flow sensor design offers the minimal amount of flow restriction needed while still producing enough signal to meet the measurement accuracy goals.

The breathing apparatus must not restrict intake airflow. In an ICU setting the flow is relatively small (± 180 liters/min maximum) and only requires a small lumen (<15 mm diameter) to support gas flow. During
exercise gas flows are in the range of ±400 liters/min. We calculate a lumen between 23-27 mm will be required to support high flow volumes and minimize resistance felt by the subject.

To build the prototype breathing apparatus we customized a flow sensor utilized in a commercial system that is currently used to determine VO₂ max during exercise. Pressure ports, already included on the housing, determine differential pressure and could be connected via tubing to a pressure transducer. As the atmosphere on the space station is a controlled environment it is only necessary to measure the concentration of expired carbon dioxide. An air-way non-dispersive infrared (NDIR) CO₂ gas analysis sensor was placed at a 45° angle into the lumen of the tube. A portion of the sensor extruded into the lumen to divert airflow to the sensor. To prevent ambient air diffusion across the sensor that may produce inaccurate readings, resistance was added to the end of the sensor in the form of additional length, Figure 4.

Figure 4 Breathing apparatus with the sensor at a 45° angle with added length to prevent diffusion back across the sensor.

III. CHARACTERIZATION OF SENSOR

Changes in sensor geometry have been shown to cause significant changes in the reported measured flow; therefore, it was necessary to verify that flow rates were not being compromised. To do this the mixed end-tidal CO₂ (ETCO₂) measurements of a reference sensor and the test sensor were compared.

A test lung was powered by an ESPRIT ventilator with the settings shown in Table 1. Carbon dioxide levels were set so that the partial pressure of ETCO₂ was between 34-36 mmHg, within an acceptable range for human ETCO₂. The mechanical lung provided a tidal volume for each breath of 1 liter.

<table>
<thead>
<tr>
<th>Table 1. ESPRIT Ventilator Parameters</th>
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Flow Host software was used to record the flow parameters. After a one minute stabilization period the flow was recorded for 5 breaths on the reference sensor. The capnostat was moved to the test sensor and flow was again allowed to stabilize for 1 minute. After collecting the flow over a period of 5 breaths on the test sensor, the reference flow was recorded once more. The average partial pressure of ETCO₂ for 5 breaths at the reference site, test site, and reference site was 35.8±0.08, 35.7±0.08, and 35.8±0.08mmHg, respectively. These findings were repeated 3 times and the ETCO₂ was consistently found to be within an acceptable range, Figure 5.

Figure 5 The average partial pressure of ETCO₂ for 5 breaths at the reference site, test site, and reference site was 35.8±0.08, 35.7±0.08, and 35.8±0.08mmHg, respectively.
IV. DISCHARGE COEFFICIENT TABLE

The relationship between flow and differential pressure is derived using a modified version Bernouilli’s equation, so that:

\[ Flow = \frac{P_m T_m}{P_{std} T_{std}} K \sqrt{\frac{\Delta P T_m}{M_w P_m}} \]  

(1)

where \( P_m, P_{std}, T_m, \) and \( T_{std} \) are the measured and standard pressures (mmHg) and temperatures (K), respectively; \( \Delta P \) is the differential pressure (mmHg), and \( K \) is a correction factor that is flow dependent and includes gas composition, flow-discharge coefficients, and other factors.\(^{16} \)

The correction factor, \( K \), is used to adjust for errors resulting from assumptions made in the derivation process.

\[ K = \frac{Actual\ Flow\ Rate}{Theoretical\ Flow\ Rate} \]  

(2)

It is a dimensionless unit that is commonly referred to as the discharge coefficient. For our purposes, \( K \) is stored as an index dependent on barometric pressure, temperature, and humidity and will be stored in a lookup table.

In order to prove that our prototype sensor and algorithms could be used to predict flow, we tested the linearity of the flow versus differential pressure relationship. An ESPRIT ventilator was used to blow pure oxygen at a known flow. The flow of the ventilator was verified using a VT Plus GasFlow Analyzer. The VT Plus was calibrated using a 3 liter syringe and found to have a percent error of ±0.83%. The differential pressure was measured using a NICO Cardiopulmonary Management System.

Figure 6 shows the relationship between the average flow and the square root of the differential pressure. At low flows the differential pressure was much more sensitive to noise, so it was necessary to gather data at 2 LPM increments. At higher flows the pressure was more stable and the relationship was very linear with an R-squared value of 0.996. The next step in building the coefficient table will be to incorporate the barometric pressure, temperature, and molecular weights of the substance to develop equations that can be used to translate the data to different conditions, i.e. in space using air instead of pure oxygen.

Figure 7 shows the discharge coefficient correction factors. For the lookup table it will be necessary to interpolate between the known values. At higher levels of flow the differential pressure is more stable, and the \( K \) correction factor becomes almost constant.

![Figure 6](image-url)

Figure 6 The linear relationship between the average flow and differential pressure. This relationship does not take into account the temperature, barometric pressure, or molecular weight of the substance.

![Figure 7](image-url)

Figure 7 The discharge coefficient, \( K \), over the differential flow.

V. FUTURE WORK

The discharge coefficients will be incorporated into the oxygen uptake system software and the flow measurement will be tested for accuracy over the range of interest.
±400 L/min. We expect that the flow measurement will be accurate to within ±3%

We will incorporate the modified flow sensor and flow measurement algorithms with the existing oxygen uptake measurement system. We will use our propane combustion patient lung simulator system to simulate an exercising subject. We will test the accuracy of the oxygen consumption measurement at 500, 1000, and 2000 ml/minute of oxygen consumption. Exercise ventilation will be simulated using a respiration rate of 20 breaths per minute and tidal volumes of 0.5, 1.0 and 2.0 liters/breath of air. We will compare the measured oxygen consumption against the known simulated oxygen consumption as calculated by the directly measured rate of propane flow into the flame. Algorithm adjustments will be made as needed until the desired accuracy of less than 5% error has been achieved.

VI. REFERENCES

2. Hawkey A. The physical price of a ticket into space. JBIS-J Br Interplanet Soc 2003;56.