Adhesion of Staphylococci to Breast Prosthesis Biomaterials: An Electron Microscopic Evaluation

T. A. Fassel  
*University of Wisconsin-Madison*

J. E. Van Over  
*Medical College of Wisconsin*

C. C. Hauser  
*Medical College of Wisconsin*

C. E. Edmiston  
*Medical College of Wisconsin*

J. R. Sanger  
*Medical College of Wisconsin*

Follow this and additional works at: [https://digitalcommons.usu.edu/cellsandmaterials](https://digitalcommons.usu.edu/cellsandmaterials)

Part of the [Biological Engineering Commons](https://digitalcommons.usu.edu/cellsandmaterials)

Recommended Citation

Available at: [https://digitalcommons.usu.edu/cellsandmaterials/vol1/iss3/2](https://digitalcommons.usu.edu/cellsandmaterials/vol1/iss3/2)
ADHESION OF STAPHYLOCOCCI TO BREAST PROSTHESIS BIOMATERIALS: 
AN ELECTRON MICROSCOPIC EVALUATION

T.A. Fassel$^{1,2,*}$, J.E. Van Over$^2$, C.C. Hauser$^2$, C.E. Edmiston$^3$, J.R. Sanger$^2$

$^1$ Muscle Biology Laboratory, University of Wisconsin-Madison, Madison, Wisconsin 53706.
$^2$ Department of Plastic and Reconstructive Surgery and
$^3$ Surgical Microbiology Research Laboratory, Department of Surgery,
Medical College of Wisconsin, Milwaukee, WI 53226.

(Received for publication February 23, 1991, and in revised form August 9, 1991)

Abstract

Gram-positive coagulase-negative staphylococci have been implicated in breast prosthes infection. Staphylococcus aureus, Staphylococcus epidermidis, and Staphylococcus hominis strains were examined for adhesion and proliferation morphology to polyurethane foam and smooth silicone rubber prosthetic biomaterials. Ruthenium red, alcin blue and ruthenium red-lysine in en bloc procedures were applied to optimally visualize the polysaccharide slime. Ruthenium red processing resulted in observation of additional outer material close to the cell wall, more than seen in the absence of cationic reagent. Alcin blue preserved fibrous or netlike strands or meshworks of material. Ruthenium red-lysine preserved considerable amounts of slime covering the cocc as well as extending between them and the prosthetic surface. Elongate strands of slime appeared to facilitate attachment to the substrate and projection of cells away from the substrate. Where this material was lacking, cell mass buildup was required for projection from the substrate. The amount of fibrous material observed corresponded to levels of slime production known for each staphylococci strain. Some extracellular material was observed for even the classified "non-slime" S. hominis strain SP2. The ruthenium red-lysine approach was optimal for visualization of slime for all bacterial species in this study.

Key words: staphylococci, bacterial adhesion, bacterial proliferation, breast prosthesis, slime, ruthenium red, ruthenium red-lysine, alcin blue, silicone rubber, polyurethane foam.

*Address for correspondence:
T. A. Fassel,
Muscle Biology Laboratory,
University of Wisconsin-Madison,
1805 Linden Drive,
Madison, Wisconsin 53706.
Phone No.: (608)-262-1547 or (608)-262-1066
FAX: 608 262 5157

Introduction

The use of biomaterials in medicine has increased dramatically over the past 20 years. The increased use of bioprosthetics in surgery has been associated with a concomitant increase in device associated infections. The staphylococci are a ubiquitous group of microorganisms which colonize the skin and are often recovered from the surface of these infected biomaterials (Edmiston et al., 1989). The adherence of these organisms to the biomaterial surface is often influenced by the structure and composition of the prosthetic along with the specific surgical technique used to insert the device (Gristina, 1987; Schmitt et al., 1986).

Breast implants are of great clinical value in augmentation and reconstructive surgery. Typically, the prosthesis consists of a silicone rubber shell or envelope which may be filled with saline or silicone gel. The outer surface may be smooth or textured silicone rubber or coated with polyurethane foam. Infections, though rare, are generally caused by gram-positive staphylococci. Overt infections caused by coagulase-positive S. aureus may result in extrusion of the prosthesis (Courtiss et al., 1979; McGrath and Burkhardt, 1984). Severe capsular contracture may result. Capsular contracture is the formation of a constricting fibrous envelope around the prosthesis, which can cause firmness and distortion of the breast (Asplund, 1984). This is the most common complication of breast prostheses and occurs generally without overt infections. Recently an association with subclinical infection by coagulase-negative staphylococci and fibrous capsular contracture has been suggested as causal (Shah et al., 1981; Burkhardt et al., 1981; Burkhardt et al., 1986). Therefore, adhesion and proliferation of staphylococci to breast prostheses has great clinical relevance.

Following a rapid adhesion of bacteria to prosthetic biomaterials is the production of polysaccharide slime (Costerton et al., 1981; Edmiston et al., 1989). It covers the bacteria and substrate, becoming a protective layer for bacterial proliferation and microcolony formation (Costerton et al., 1981; Peters et al., 1982; Franson et
al., 1984). The present mechanisms of initial microbial adherence are poorly understood. But extracellular polysaccharide which we collectively call slime is likely responsible for persistence of the organisms to the biomaterial surface. Production of slime is a known virulence factor (Edmiston et al., 1989) and is enhanced in the presence of a foreign body (Christensen et al., 1983). Secretion and proliferation of polysaccharide slime is an important part of the complex phenomenon of bacteria polymer associated infection. Staphylococcus aureus strain RP62 are known producers of slime. Staphylococcus hominis strain SP2 is classified as a non-slime producer. The aim of this approach was two-fold. First, to optimize visualization of the polysaccharide slime layers in prosthetic infection culture. Second, to observe the morphology of staphylococci adhesion and proliferation to the biomaterial surface.

**Materials and Methods**

**Bacterial Adherence Studies**

Three staphylococci strains were tested for adherence characteristics against polyurethane foam and silicone rubber; *S. aureus* ATCC25923, *S. epidermidis* RP62 (slime producing strains) and *S. hominis* SP2 (characterized as a non-slime producer). The organisms were recovered from frozen storage (-70°C) and plated to blood agar plates to check viability. After 24 hours the organisms were inoculated to trypticase soy broth and incubated for an additional 18 hours at 35°C. The test substrates were polyurethane foam and smooth silicone rubber (Surgitek, Inc., Racine, WI) which were cut into 1 cm² segments for standardized studies. The prosthesis segments were not exposed to additional reagents or precleaned.

Adherence studies were performed by incubating 1 cm² polyurethane or smooth silicone rubber segments in a standardized staphylococcal inoculum (7.0 log₁₀ colony forming units/ml). This was determined using optical densitometry and comparison with known standards. Prosthetic samples were removed at 4 and 24 hours post inoculation. Non-adherent cocci were removed by washing the prosthetic segments in Phosphate Buffered Saline pH 7.0 (3x) followed by ultrasonic oscillation at 20 KHz for 10 minutes which dislodged adherent cocci. Scanning electron microscopy of sonicated specimens was performed to confirm adequacy of dislodgement. The sonicate was serially diluted, plated on trypticase soy agar and the plates inspected at 4 and 24 hours. Microbial adherence was expressed as the log₁₀ counts per cm² of prosthetic surface.

**Scanning Electron Microscopy**

For the glutaraldehyde/Os₄ procedure, samples were fixed in 2.5% glutaraldehyde in buffer for 2 hours, washed three times for 10 minutes duration each wash, postfixed in 1% (w/v) Os₄ for 2.5 hours, and washed three times for 10 minutes each. The buffer, 0.1 M cacodylate pH 7.3, was used for all solutions.

For the RR procedure, pre-fixation was in 0.2% glutaraldehyde and 0.15% RR for 30 minutes. This was followed by fixation in 1% glutaraldehyde and 0.05% RR for 2 hours. The 0.05% RR was included with 0.1 M cacodylate buffer for three washes of 10 minute duration each. Postfixation in 2% Os₄ and 0.05% RR for 2.5 hours preceded another wash cycle (Fassel et al., 1990). For the alcian blue procedure, AB was substituted for RR in all solutions at identical percentages as in the RR procedure (Fassel et al., 1991). For the RR-lysine procedure, samples were prefixed in 75 mM lysine, 0.075% RR and 2.5% glutaraldehyde for 20 minutes. Fixation in 0.075% RR and 2.5%...
Staphylococci adhesion to breast prosthesis

Figure 1A: Polyurethane foam biomaterial has an interstitial nature and honeycomb appearance with a considerable surface for bacterial adhesion and proliferation. Bar = 500 µm.

Figure 1B: Clusters of *S. epidermidis* RP62 (arrows) preserved by the RR-lysine procedure adhere to the curving foam surface. Bar = 100 µm.

Figure 2A: The silicone rubber biomaterial, characterized by a smooth single surface plane, has several *S. epidermidis* RP62 cells (several enclosed in box) attached, glutaraldehyde/OsO₄ fixation. Bar = 10 µm.

Figure 2B: Clusters of *S. epidermidis* RP62 project outward from the silicone rubber surface. Note also, the absence of extracellular slime by the glutaraldehyde/OsO₄ fixation. Bar = 5 µm.

Results

The polyurethane foam biomaterial has a honeycomb appearance (Figure 1A) with many interstices and surfaces that provide numerous opportunities for adhesion and proliferation by staphylococci. Figure 1B (RR-lysine procedure) shows clusters of *S. epidermidis* RP62 (arrows) on the curved surface.

In contrast, the silicone rubber biomaterial provides a single surface plane with infrequent irregularities. Irregularities in sterile control specimens have been
Figure 3A: By the RR procedure, fibrous cellular surface material (arrowheads) and strand-like extensions between cells are preserved for *S. epidermidis* RP62 cocci on polyurethane foam. Bar = 2.5 µm.

Figure 3B: By the RR-lysine procedure, a more extensive elaboration of slime is preserved with fibrous material (arrowheads) densely covering cells. Strands also extend between *S. epidermidis* RP62 cells and the polyurethane foam surface (arrow). Bar = 5 µm.

Figure 3C: By the AB procedure, fine fibrous strands (arrows) cover *S. epidermidis* RP62 on silicone rubber. Bar = 5 µm.

Figure 3D: Also by the AB procedure, extensive net-like meshwork (arrows) are seen holding *S. epidermidis* RP62 in stable projections from polyurethane foam substrate. Bar = 10 µm.

observed previously and are also associated with colony formation in *in vitro* studies (Sanger et al., 1989). Individual *S. epidermidis* RP62 cells adhered to the smooth surface are shown in Figure 2A, glutaraldehyde/OsO₄ fixation. The buildup of clusters project outward (Figure 2B). Additional irregular material is frequently found on the silicone rubber surface beneath or in the vicinity of cells. The absence of extracellular bacterial slime on the smooth round cocci is also illustrated here, by the glutaraldehyde/OsO₄ fixation. Some fibrous cellular surface material (arrowheads) is preserved, as well as, strand-like extensions between some *S. epidermidis* RP62 cocci on polyurethane foam (Figure 3A) by the RR procedure. A more extensive elaboration of slime is seen for the RR-lysine procedure (Figure 3B). Here, fibrous material (arrowheads) densely covers cocci and clusters. Strands also extend between cells and the foam surface (arrow). By the AB procedure, a fine material of fibrous strands (arrows, Figure 3C) is also preserved, shown here for *S. epidermidis* RP62 on silicone rubber.
Staphylococci adhesion to breast prosthesis

Figure 4A: For *S. hominis* SP2 cocci project into the interstitial space of the polyurethane foam. Few fibrous strands (arrows) extend between the smooth cells by the glutaraldehyde/OsO₄ procedure. Bar = 5 µm.

Figure 4B: By the RR procedure, more fibrous strands (arrows) are preserved between smooth *S. hominis* SP2 cocci on silicone rubber. Bar = 5 µm.

Figure 4C: By the AB procedure, some surface material (arrowheads) is seen on some *S. hominis* SP2 cells. Fine strands also extend between cells (arrows). Bar = 5 µm.

Figure 4D: By the RR-lysine procedure, most extensive extracellular material upon and between cells (arrowheads) is seen. Bar = 10 µm.

Occasionally, an extensive net-like meshwork (arrows) is formed between cells, holding them in apparently stable configurations that can project well away from the polyurethane foam substrate (Figure 3D).

For the "non-slime" former, *S. hominis* SP2, a smooth surface characterizes the cocci (Figure 4A) by the glutaraldehyde/OsO₄ procedure. The ability to build stable projections away from the polyurethane foam and extending into the interstitial space is still present, although made up of many more cells (Figure 4A versus Figure 3D). A few fibrous strands can be observed between cells (arrows, Figure 4A). The strands are more extensively preserved by RR (Figure 4B, arrows) with many extensions seen between smooth cells, networking across the silicone rubber surface. By the AB procedure (Figure 4C), the observations are similar, with some additional surface material (arrowheads) on some cells and fine strands extending between cells (arrows). Deposits of material are also found. The most extensive buildup of extracellular material upon and between cells (arrowheads) is seen by the RR-lysine procedure (Figure 4D).

203
S. aureus shows a smooth cell surface and absence of slime by the glutaraldehyde/OsO₄ fixation (Figure 5A, silicone rubber). Ruthenium red (Figure 5B) reveals a cluster of cells building up from the polyurethane foam substrate. Extracellular material is infrequent. Many more fibrous strands (arrow) cover some cells and extend between cells and foam substrate (arrowhead, Figure 5C) by AB. Figures 5D, E and F compare the increased preservation of slime by the RR-lysine procedure between silicone rubber and polyurethane foam substrates for S. aureus. For both substrates, the slime material is more elaborate over cells than by the other
Staphylococci adhesion to breast prosthesis

procedures. On the silicone rubber (Figures 5D and 5E) extracellular slime covers cells forming thickened connections between cells (long arrow). It is occasionally observed to increase, forming extensive overlapping masses of strands (Figure 5E left). Cells from this area (Figure 5E right) show some surface material. An extracellular meshwork of this density has not been observed on polyurethane foam to date for *S. aureus*, although cell surface material and more extended strands between cells are seen (Figure 5F).

Interestingly, adhesion kinetic data (Sanger et al., submitted) suggests more *S. aureus* cocci adhere to silicone rubber (14,800 ± 6,800 CFU/sq.cm.) than to polyurethane foam (8,000 ± 1,673 CFU/sq.cm.). The noted slime producer *S. epidermidis* RP62 prefers the polyurethane foam substrate (276,800 ± 28,869 CFU/sq.cm.) to the silicone rubber (20,000 ± 1,789 CFU/sq.cm.). To polyurethane foam, the "non-slime" producer *S. hominis* SP2, shows less tendency to adhere to either substrate (polyurethane foam: 10,400 ± 5,075 CFU/sq.cm., silicone rubber: 2,400 ± 1,166 CFU/sq.cm.).

Legend for Figure 5 on the facing page

Figure 5A: *S. aureus* shows a smooth cell surface and absence of slime by the glutaraldehyde/OsO₄ fixation on silicone rubber. Bar = 2.5 µm.

Figure 5B: By the RR procedure, extracellular material is infrequently seen between a cluster of *S. aureus* cells from the polyurethane foam substrate. Bar = 2.5 µm.

Figure 5C: By the AB procedure, fibrous strands (arrow) cover some cells. Extensions of slime (arrowhead) are seen between cells and the foam substrate. Bar = 5 µm.

Figure 5D, E and F: By the RR-lysine procedure, silicone rubber (D and E) and polyurethane foam (F) substrates for *S. aureus* are shown. Extracellular slime covering cells (arrow) and forming thickened connections between cells (long arrow) appears on silicone (D). This extracellular slime is occasionally observed to increase to form extensive overlapping masses of strands (E left). Cells from this area (enclosed in box, E left) are shown at increased magnification in E right. Some surface material is apparent (arrow). This extensive formation has not been observed to date on polyurethane foam (F), although cell surface material and more extended strands between cells are seen (arrows). Bars = 5, 50, 5 and 5 µm respectively.

Discussion

Visualization of polysaccharide slime layers important in bacterial adhesion and proliferation to breast prosthetic material was improved by use of cationic reagents, RR-lysine and AB, in *en bloc* procedures. Lysine, a diamine, is positive at physiological pH. It is thought to form large cross-linked polymers with glutaraldehyde. Biological structure fixed with these complexes have improved stability through later stages of electron microscopy processing (Boyles, 1984; Boyles et al., 1985). Ruthenium red reacts with highly negatively charged polyanions, such as acidic mucopolysaccharides (Luft, 1971a; 1971b). Ruthenium red-lysine procedures have been highly effective in improving visualization of bacterial outer layers (Akin and Rigsby, 1990; Davies and Borriello, 1990; Jacques and Graham, 1989; Jacques et al., 1990). Alcian blue has also been used previously in improving visualization of extensive outer bacterial layers (Progulske and Holt, 1980; Herald and Zottala, 1988; Fassl et al., 1991). Its reaction with acidic mucopolysaccharide through either electrostatic or ionic interactions (Luft, 1971a; Scott et al., 1964) improves visualization of polysaccharide slime material.

The RR-lysine procedure was most effective in preserving extensive elaborations of slime. It was similar for all species and substrates. Additionally, thickened masses of fibrous strands were seen for *S. aureus* on smooth silicone rubber. It is unknown whether this observation may concur with the observed tendency for *S. aureus* cells to preferentially adhere to smooth silicone rubber versus polyurethane foam. Such an extensive build up of slime could provide an additional protection and favorable environment for bacterial proliferation. This buildup would favor infection by *S. aureus* of silicone rubber prosthesis. Infection of a breast prosthesis by coagulase-positive *S. aureus* results in a purulent discharge often controlled only by removal of the prostheses (Courtiss et al., 1979). Coagulase-negative staphylococci also adhere well to smooth silicone rubber. Clinically these organisms tend not to produce a purulent response and have been found in association with fibrous capsules (Shah et al., 1981; Burkhardt et al., 1981). Perioperative antibiotics have reduced the incidence of capsular contracture (Burkhardt et al., 1986) around smooth silicone implants. As demonstrated in this study, polyurethane foam provides an excellent surface for adherence and proliferation of coagulase-negative staphylococci. However, early clinical results show a lower incidence of contractures with polyurethane foam covered prostheses than smooth silicone rubber (Capozzi and Pennesi, 1981; Melmed, 1988; Shapiro, 1988). Longer follow up will be needed to determine if coagulase-negative infections emerge as a clinical problem.
The AB procedure mainly preserved extracellular strands of slime between *S. aureus* and *S. hominis* SP2 cells. More extensive elaborations were seen for the noted slime producer, *S. epidermidis* RP62. For *S. epidermidis* RP62, extensive material covered cells in a manner similar to RR-lysine. Strands of slime also provided sufficient support for projection of cells out into the interstitial space of polyurethane foam. *S. epidermidis* RP62 preference for proliferation on polyurethane foam over the one surface plane of silicone rubber, appears to agree with this scanning electron microscopy observation.

Ruthenium red provided the least improvement in slime preservation of the procedures compared here. It did increase strands for *S. hominis* SP2 and *S. epidermidis* RP62, as well as some fibrous cell surface material for *S. epidermidis* RP62. It was least effective for *S. aureus*.

The least effective of all the procedures tested was the glutaraldehyde/OsO₄ fixation where no cationic reagents were employed. For this procedure, only a few extracellular strands were seen for *S. epidermidis* RP62 or *S. hominis* SP2. None were observed for *S. aureus*.

Thus, RR-lysine was optimal for all species tested. The effectiveness of AB varied with the species, with it the most effective for *S. epidermidis* RP62. This variation in species effectiveness of slime preservation may be due to some difference in polysaccharide composition of the slime between species.

Morphological features of elongate strands appear to facilitate attachment to the biomaterial substrate. Strands between cells aided the buildup and projection of cocci away from the substrate surface, this was most notable for *S. epidermidis* RP62. This could agree with the strong tendency for *S. epidermidis* RP62 cells to adhere preferentially to polyurethane foam versus silicone rubber (Sanger *et al.*, submitted). However, buildup of a cluster away from the substrate was also noted with proliferation of individual cells, as for *S. hominis* SP2.

A classified "non-slime" producer, *S. hominis* SP2, was found to form extracellular features similar to that of other slime producers, although apparently less in quantity. This agrees with other work (Goheen *et al.*, 1990), where use of cationic reagents preserved extracellular slime for this strain. This further illustrates the advantage of this approach of using cationic reagents to improve slime preservation for scanning electron microscopy.

References


Davies HA, Borriello, SP (1990) Detection of capsule in strains of *Clostridium difficile* of varying virulence and toxigenicity. Microbial Pathogenesis. 9, 141-146.


Fassel TA, Schaller MJ, and Remsen CC (1991) Comparison of alcian blue and ruthenium red effects on
Staphylococci adhesion to breast prosthesis


Discussion with Reviewers

H. Freeman: Your study indicates greater attachment of S. aureus on silicone rubber substrates than on polyurethane foam. Is this observation a consequence of the highly convoluted nature of the foam or an inherent difference in the biomaterial surface itself? What is a plausible reason for such a difference?

Authors: We feel an inherent difference of the biomaterial surface is more likely. The convolutions of the foam do not appear to interfere with dislodgement by sonication. SEM of sonicated specimens indicates efficient bacterial removal from either substrate. Also, S. epidermidis RP62 has a greater attachment to the polyurethane foam than silicone rubber (Sanger et al., submitted). The three dimensional curving foam surface versus the one dimensional surface plane of the silicone rubber presents the same problem for both species. A preference, in either case, for the chemical nature or other inherent property of the substrate may be more likely. Clarification of this issue awaits further study.

H. Freeman: Did you use a smooth or textured silicone rubber prosthesis envelope in your studies? Would you expect there to be a difference in the adhesion of staphylococci strains?

Authors: Only smooth silicone rubber was used in this study. It is currently unknown how smooth versus textured silicone rubber affects adhesion. This is a subject of future study.

A. Molinari: In previous studies, the use of lectins to avoid the loss of capsular material of gram-positive bacteria during the sample preparation procedures for electron microscopy, was introduced. Data obtained indicated that lectins are useful agents in preserving...
highly water-soluble capsular components for both unembedded and embedded samples (G. Orefici et al., 1986 FEMS Microbial. Lett. 34: 111-115; A. Molinari et al., 1988 Histochemical Journal 20, 526-530). Lectins show specific affinity to different glucidic residues, then they can give more information about the difference in polysaccharide composition of the surface structures of the microorganisms. This methodological approach might also be applied in the study of the production of polysaccharide slime. Could the authors discuss this point?

Authors: The use of lectins in the visualization of microbial capsular material has been documented by other investigators. It is plausible that specific lectin-glucidic acid residue interactions could be used to study the production of microbial slime under different physiologic and environmental conditions.