**ABSTRACT**

Spider silks have long been a focus of research due to their remarkable mechanical properties including strength, toughness and elasticity. Moreover, biodegradability and biocompatibility of spider silks make them beneficial to use in biomedical applications. Spiders cannot be farmed because of their territorial and cannibalistic nature. Hence, production of recombinant spider silks is the only feasible solution for large scale production.

Large scale production is still challenging due to the small recombinant protein size, low yield and low water solubility of bio-synthetic spider silk. Our main concern is the production of recombinant spider silks is the only feasible solution for large scale production and purification.

Our main spidersilk protein we seek to produce and purify in a large scale is MaSp2. Spider dragline silk is primarily composed of proteins called major ampullate spidroins (MaSp) that consist of a large repeat array flanked by nonrepetitive N- and C- terminal domains. All MaSp genes are co-expressed in the major ampullate gland of Nephila Clavipes.

**RESULTS**

Figure 2: Production of spider silks at (A) 5L (B) 20L (C) 100L volumes. Note: 1L bottle to scale

**MATERIALS AND METHODS**

**Protein Production Using Bioreactors**

A) TFF Column B) 500L Bioreactor C) Homogenizer

**Discussion**

Initially transgenic goats were utilized to produce spidersilk protein MaSp1 and MaSp2. Our goal of large scale protein production and purification could not be met ideally by this method. It is very time consuming and inconvenient. For example in E. Coli, it is relatively easy to make multiple constructs but for a goat it would take a long time. Goats also cost more money to maintain (food, land, resources). Use of Spidersilk protein commercially would be more difficult using a transgenic animal such as goats due to regulations and negative stigma toward GMOs, as opposed to mass producing via E. Coli. Using bacteria such as E. Coli is beneficial due to its much lower cost margin, short generation time, and well understood and researched methods of inserting genes and producing desired proteins.

**Methods of Protein purification**

Our proteins MaSp1 and MaSp2 have a polyhistidine tag, which is a string of histidine residues typically at either N terminus or the C terminus of a recombinant protein. Originally to purify our protein we used a Nickel Column. Nickel columns are used for immobilized metal affinity chromatography for the purification of recombinant proteins with a polyhistidine tag at either terminus. Our protein would bind to the nickel column and then collected via elution with a high concentration of Imidazole. This process however was too expensive for large scale protein production and purification.

For a while, we used Ammonium Sulfate precipitation. Following fermentation, cell pellets were collected via centrifugation and suspended in a lysis buffer (.5M NaCl 200mM Tris). The cell suspension was sonicated for an hour and the disrupted cells were incubated at 70-80°C for 30 minutes to precipitate E. coli proteins. The protein was precipitated by adding ammonium sulfate. The purified protein pellet was washed till the conductivity of the wash reaches 10 microsiemens and lyophilized. Ammonium sulfate precipitation was also too inconvenient and time consuming. We would also expose our protein to a lot of salt that would take a lot of time to wash.

Currently, the cheapest and most efficient method of purifying our protein, that complements large scale production is by the use of TFF columns. Tangential flow filtration (TFF) is a rapid and efficient method for separation and purification of biomolecules. Our protein is around 90 kDa. The pore size in the TFF columns we use are 750 kDa and 50 kDa. The 750 kDa column washes away larger molecules, while we further remove smaller impurities in the 50 kDa column as well as concentrating our protein. We “back flush” the 50 kDa column to collect our protein. TFF columns can be used over and over again with low maintenance and can filter samples from 10mL to thousands of liters.

**CONCLUSIONS**

- Goal is produce 250 grams of purified spidersilk protein
- Currently we reach around one gram of protein per liter fermented in our bioreactor.
- We are optimizing the TFF protocol to recover more spidersilk protein. By working with different transmembrane pressures (TMP) increase shear stress (stress induced on the liquid to degrade the protein) by trying a stronger pump, and flow rates.

**ACKNOWLEDGEMENTS**

Funding: Utah Science Technology and Research (USTAR), the Department of Biology.