Global Evaluation of Biofuel Potential from Microalgae

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GLOBAL EVALUATION OF BIOFUEL POTENTIAL
FROM MICROALGAE

by

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ABSTRACT

Global Evaluation of Biofuel Potential from Microalgae

by

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Traditional terrestrial crops are currently being utilized as a feedstock for biofuels but resource requirements and low yields limit the sustainability and scalability. Comparatively, next generation feedstocks, such as microalgae, have inherent advantages such as higher solar energy efficiencies, larger lipid fractions, utilization of waste carbon dioxide, and cultivation on poor quality land. The assessment of microalgae-based biofuel production systems through lifecycle, technoeconomic, and scalability assessments has been forced to extrapolate laboratory-scale data due to the immaturity of the technology. This type of scaling leads to large uncertainty in the current near-term productivity potential and ultimately the results from modeling work that rely on this type of modeling. This study integrated a large-scale validated outdoor microalgae growth model that utilizes 21 species and reactor-specific inputs that accurately account for biological effects such as nutrient uptake, respiration, and temperature with hourly historical meteorological data from around the world to determine the current global productivity potential. A global map of the microalgae lipid and biomass productivity has
been generated based on the results of annual simulations at 4,388 global locations spread over the seven continents. Maximum annual average yields between 24-27 m³·ha⁻¹·yr⁻¹ are found in Australia, Brazil, Colombia, Egypt, Ethiopia, India, Kenya, and Saudi Arabia with the monthly variability (minimum and maximum) yields of these locations ranging between 14 and 33 m³·ha⁻¹·yr⁻¹. A scalability assessment that leverages geographic information systems data to evaluate geographically realized microalgae productivity, energy consumption, and land availability has been performed highlighting the promising potential of microalgae-based biofuels compared to traditional terrestrial feedstocks. Results show many regions can meet their energy requirements through microalgae production without land resource restriction. Discussion focuses on sensitivity of monthly variability in lipid production compared to annual average yields, biomass productivity potential, effects of temperature on lipid production, and a comparison of results to previous published modeling assumptions.
PUBLIC ABSTRACT

Global Evaluation of Biofuel Potential from Microalgae

The objective of the proposed work is to determine the productivity potential of microalgae around the world based on the current large uncertainty of the productivity potential found in literature. To achieve this objective, a validated thermal and biological growth model was utilized coupled with weather data files from weather stations around the world. This enabled a realistic assessment of the productivity potential based on actual climatic variables. Sensitivity of microalgae lipid productivity to biomass production, temperature, and variability was performed illustrating the importance of biological and temporal resolution.

Results from modeling work were leveraged for a scalability assessment based on transportation fuel consumption and land availability statistics from around the globe. A comparison of the results from this study to the current assumption in literature shows the community has dramatically overestimated the current near term productivity potential in literature.

As research into microalgae continues to grow, many studies are being performed to understand the energetic and economic feasibility of converting microalgae to biofuel. The large variability as to the true current productivity potential of microalgae has detrimentally affected large-scale assessments of the microalgae-to-biofuels system. The large uncertainty has resulted in favorable skewing of the environmental impact, economic feasibility, and resource requirements of the microalgae-to-biofuel process. The results from this study offer a better understanding of the true large-scale near term productivity of microalgae based on light, nutrient, and temperature factors to better inform future energetic and economic assessments of biofuel from microalgae.

Jeffrey W. Moody
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<td>photosynthetic rate (carbon)</td>
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<td>-</td>
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<td>$\varphi_T$</td>
<td>temperature efficiency factor</td>
<td>-</td>
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INTRODUCTION

Global oil consumption is rising at an unprecedented rate primarily due to increasing consumption in developing nations. From 1990 to 2012, China, India, and Brazil have increased oil consumption by 311%, 186%, and 100%, respectively, and currently consume 19.6% of the 3,590 million tonnes (Mt) of the oil produced annually on the planet [1]. Recent volatility in oil prices, attributed to increased demand and limited resource, has led to the development of unconventional petroleum reserves such as oil sands and increased exploration of alternative and renewable fuel sources. Alternative renewable energy sources have been investigated as a way to alleviate dependency on fossil fuels while positively impacting the environment. Scalability limitations associated with traditional terrestrial biofuel feedstocks have renewed interest in next generation feedstocks such as microalgae. Some of the advantages that microalgae offer over traditional terrestrial oil crops are: higher lipid productivities, lack of competition for arable land, year-round cultivation, integration with saline and low quality water sources, and higher solar energy yields [2, 3]. These scalable advantages make microalgae a promising feedstock for biofuel production and a potential sustainable alternative to traditional petroleum fuels.

Compared to the yields from traditional terrestrial crops such as corn (0.172 m$^3$·ha$^{-1}$·yr$^{-1}$), soybeans (0.636 m$^3$·ha$^{-1}$·yr$^{-1}$), and rapeseed (0.974 m$^3$·ha$^{-1}$·yr$^{-1}$), microalgae are reported to have productivities that are orders of magnitude higher [4-6]. Consensus on the true near term productivity potential of microalgae is proving challenging due to the immaturity of the industry. As a results, the productivity potential for microalgae at large-scale is being estimated through the linear scaling of laboratory based growth and lipid data which has led to a large variance in reported values [7, 8]. This type of scaling has been integrated into various lifecycle,
technoeconomic, and resource assessments of the microalgae to biofuels process. The simplistic scaling of small-scale laboratory based data for large-scale microalgae biofuel assessments leads to erroneous assumptions about industrial function, and is a source of large uncertainty [3]. Values reported in lifecycle, technoeconomic, and resource modeling literature range from 2.3 m³·ha⁻¹·yr⁻¹ reported by Ramachandra et al. [7] to 140 m³·ha⁻¹·yr⁻¹ reported by Pate [8] with Lam et al. [9], Resurreccion et al. [10], Quinn et al. [11], Scott et al. [6], Vasudevan et al. [12], Guieysse et al. [13], Clarens et al. [14], Schenk et al. [15], ANL et al. [16], Lardon et al. [17], Campbell et al. [18], Davis et al. [19], Moheimani et al. [20], Chisti et al. [4, 21, 22], Rodolfi et al. [23], Wijffels and Barbosa [2], Williams and Laurens [24], Yeang [25], Liao et al. [26], Batan et al. [27], and Mata et al. [5] reporting values between these extremes. The higher productivities reported by Pate [8] and Mata et al. [5] are within a factor of 3 of the thermodynamic theoretical maximum lipid yield of 372 m³·ha⁻¹·yr⁻¹ [28, 29]. Large uncertainty in the above reported productivity potential stems from the use of simplistic growth modeling either through simple solar conversion efficiency or linear scaling of laboratory data, which both fail to incorporate biological function and geographic diversity. Errors in microalgae production modeling at large-scale have been propagated throughout to skew economic, lifecycle, and scalability assessments as lipid yield typically represents the functional unit in these assessments.

Certainty in the near-term microalgae productivity is required to better inform large-scale assessments. Decreasing the uncertainty in the current productivity potential from microalgae requires increased fidelity in growth modeling through temporal and biological resolution combined with geographically specific climatic and resource data [30]. This study integrates a microalgae growth model with hourly historical metrological data from various global locations for the assessment of the lipid and biomass productivity potential of microalgae
cultivated in photobioreactors. The microalgae growth and lipid content is simulated on an hourly time scale over the course of 1 year at 4,388 global locations through the use of a minimum of 12 and a maximum of 25 (depending on site) years of metrological data. Results from annual simulations were interpolated to produce a dynamic global map of microalgae lipid productivity and intended to represent the current large-scale production potential based on a photobioreactor architecture. Discussion focuses on the effects of temperature on productivity, a geographically specific scalability assessment, monthly variability in productivity, and a comparison of modeled results to productivity potentials reported in microalgae biofuel lifecycle, technoeconomic, and scalability literature.
MATERIALS AND METHODS

The following sections present details on the thermal and biological growth models representative of *Nannochloropsis* cultivated in a photobioreactor architecture including an overview of validation efforts, meteorological data, and simulation architecture, Fig. 1. Integrating the validated thermal and biological growth models with historical hourly meteorological data enables simulation of microalgae growth at the various observed locations. Results from simulations are leveraged to produce a dynamic global map of the lipid productivity through the surface interpolation of annual results. A sensitivity analysis of the lipid yield to biomass productivity, annual variability, and temperature was performed. Thermal and biological modeling were performed in MatLab with results processed and land availability statistics generated using Esri ArcGIS [31].
Photobioreactor Thermal and Biological Growth Models

Model validation was done utilizing data from Solix photobioreactors. A schematic of the overall geometry is presented in Fig. 2. The Solix reactor test bed is located in Fort Collins Colorado adjacent to the Engines and Energy Conversion laboratory at Colorado State University. The basin measures 3 meters by 18 meters and has sixteen reactors with dimensions detailed in Fig. 2. The outer most reactors receive more light and grow at a slightly elevated rate and were not included in the data set used for validation. The system was operated in two groups of 8 reactors. Growth was monitored continuously in one of the eight reactors and manual samples were taken from all reactors to verify uniform growth.

Fig. 2 Diagram and photograph of the Generation 3 Solix photobioreactor used to validate bulk growth model. Reactors are evenly spaced in a structural/thermal water basin in Fort Collins, Colorado, USA
Reactors were harvested as a group. All of the culture was removed from the reactors, mixed for homogeneity and the required inocula were removed. The remainder of the culture was harvested by centrifugation. New nutrient rich media was prepared, filtered, and added to the inocula. It is noted that media recycling (centrate from the centrifuge) could be done but was not standard practice. The required culture volume was then re-injected into the reactors to complete the inoculation process.

The biological growth model requires 21 inputs with light and temperature serving as the primary inputs [3]. A thermal model of the shallow pool is used to accurately represent the temperature of the system. The temperature of the shallow pool (assumed to be equivalent to the photobioreactor temperature) and corresponding meteorological conditions serve as the primary inputs to the growth model which predicts biomass and lipid content.

The thermal model of the growth system accounts for heat loss and gain from conduction, convection, evaporation, and solar incidence as presented in Fig. 3. The model incorporates the climatic variables of atmospheric pressure, cloud cover, wind speed, wind direction, dry-bulb temperature, dew-point temperature, and solar radiation to predict the temperature of the shallow basin.
Fig. 3  Photobioreactor water basin thermal resistance model incorporated into the thermal pool model to predict the temperature at each of the 4388 global locations

The water basin where the photobioreactors resided was separated into 16 equally spaced vertical nodes. The approach used to calculate the temperature of the water was adapted from Weyer-Geigel [30]. The heat balance equation applied at each node is:

\[ E_{in} - E_{out} + E_{generated} = E_{stored} \]  \hspace{1cm} (1)

It is assumed the temperature of the water in the basin will be equivalent to the microalgal culture temperature as the polyethylene does not represent a thermal barrier. The heat balance equation for the temperature at any given internal node, \( n \), becomes:

\[ G_n - \frac{k}{L_n} (T_n - T_{n-1}) - \frac{k}{L_n} (T_n - T_{n+1}) = m_n c_p \frac{\Delta T_n}{\Delta t} \]  \hspace{1cm} (2)

where \( G_n \) represents solar radiation incident at node \( n \), \( k \) is the thermal conductivity of water, \( L_n \) is the distance between nodes, \( m_n \) is the mass of node \( n \), \( c_p \) is the specific heat of water,
and $T$ is the temperature. To represent the surface and ground node the following two equations were utilized:

$$G_{\text{sur}} - \frac{k}{L_n} (T_{\text{sur}} - T_1) - h_r (T_{\text{sur}} - T_{\text{sky}}) - h_i (T_{\text{sur}} - T_{\text{ambient}}) = \frac{m_n}{2} c_p \frac{\Delta T_{\text{sur}}}{\Delta t}$$  \hspace{1cm} (3)

$$G_{\text{bottom}} - \frac{k}{L_n} (T_n - T_{n-1}) - Q_i = \frac{m_n}{2} c_p \frac{\Delta T_n}{\Delta t}$$  \hspace{1cm} (4)

where $G_{\text{sur}}$ and $G_{\text{bottom}}$ represent the solar radiation at the surface and bottom of the reactor, respectively, $h_r$ is the net radiation coefficient with the sky, $Q_i$ is the stored or released energy from the ground, and $h_i$ is the convection coefficient.

The thermal model is used to calculate the culture temperature and input to the biological growth model where it is leveraged to calculate a temperature efficiency factor that effects growth and nutrient uptake. Cultivation is assumed to shut down and stays dormant when the basin temperature falls below freezing with cultivation resuming upon thaw.

The time resolved biological growth model incorporates 21 species and reactor specific inputs to accurately model the biomass and lipid productivity based on real-world climatic and thermal conditions [30]. The model captures growth through carbon uptake and includes photosynthetic rate ($P_c$), respiration rate ($rR_c$), and biosynthetic energy required for nitrogen uptake ($\Psi$) to calculate the growth rate ($\mu$):

$$\mu = P_c - rR_c - \Psi$$  \hspace{1cm} (5)

The biological growth model incorporates maximum photosynthetic rate based on a maximum carbon uptake ($P_{C,\text{max}}$), available photoactive radiation (PAR) ($E_{\text{av}}$-average PAR intensity in photobioreactor, $\varphi_m$-photon efficiency) including attenuation and losses based on Lambert-Beer absorption assumptions ($\alpha$-species specific absorption coefficient), temperature efficiency ($\varphi_T$), and nitrogen efficiency ($\varphi_{N,X_{\text{int}}}$) to calculate the carbon fixation rate:
The photosynthetic productivity is a non-linear function of PAR. The model accurately captures two of the three regions of the photosynthetic productivity curve, 1) a photon flux density dominated region where the photosynthetic rate increases linearly with increasing light intensity and 2) a light saturation photosynthetic region characterized by constant photosynthetic rate with increasing PAR. The model does not include the third region, a photo-inhibition region, characterized by a decrease in photosynthetic rate with increasing photon flux based on an assumption of high culture densities and mid to low level light intensities (sunlight sourced) incident on the photobioreactors [32-35]. Consistent with biological experimentation, the second region of the photosynthetic curve is effected by temperature and nitrogen efficiency terms with the initial photosynthetic slope not effected [32, 33, 36, 37].

The temperature dependency of photosynthesis is modeled through the corresponding effect on ribulose-biphosphate carboxylase (Rubisco) activity. The enzyme activity as a function of temperature is modeled based on a bell shaped curve with the optimal temperature at the peak corresponding to the maximum temperature efficiency ($\varphi_T$) [38]. Deviating from the ideal temperature in either direction corresponds to a decrease in temperature efficiency, with the temperature efficiency factor being a dimensionless number between 0 and 1, as presented in Eq. (7):

$$
\varphi_T = \frac{2^{-\frac{E_a}{RT_{opt}}} \frac{E_a}{R_T}}{1+e^{-\frac{E_a}{RT_{opt}}} \frac{E_a}{R_T}}
$$  (7)

From Eq. (7), the optimum temperature (23 C for the modeled species) is captured in $T_{opt}$. $R$ is the ideal gas constant, $E_a$ is the activation energy of carboxylation Rubisco, and $T$ is the culture temperature calculated in the thermal model. The effect of the temperature efficiency
factor on growth is illustrated through simulating the biomass growth over the course of one week in Learmonth, Australia. Temperature efficiencies of 1, 0.8, and 0.5 were simulated with biomass yields presented in Fig. 4. As expected, the maximum biomass is attained under optimum thermal conditions ($\varphi_T = 1$) with lower or higher temperatures decreasing the overall yield. Temperature indirectly affects the total lipid content in the cell through the effects on nitrogen uptake and thus does not have a linear effect on lipid productivity. Biomass results integrated with lipid modeling results yield 22.45, 20.41, and 15.62 m$^3$·ha$^{-1}$·yr$^{-1}$ for the temperature efficiency scenarios of 1, 0.8, and 0.5 respectively. For global modeling the temperature efficiency is dynamic and calculated on an hourly basis at each simulation location.

![Fig. 4](image_url)  
**Fig. 4**  Simulation of one week of growth in Learmonth, Australia with three different temperature efficiencies
A lipid model was created and incorporated into the biological growth model to accurately predict the microalga lipid production based on nitrogen, temperature, and light intensity affects. This model is based on a nitrogen trigger with cell function switching from protein to lipid synthesis while maintaining a mass balance between the lipid, carbohydrate, and protein:

\[ m_B = m_L + m_C + m_P \]  

(8)

where \( m_B \) is the mass of the biomass and \( m_L, m_C, \) and \( m_P \) are the mass of the lipid, carbohydrate, and protein, respectively.

The thermal and microalga growth model incorporates light and temperature as primary inputs to model carbon fixation based on the activity of the Rubisco enzyme while considering respiration losses and energy requirements for nutrient uptake. Validation of the model was performed through the collection of 9 weeks of biological growth and meteorological data at a large-scale outdoor growth facility. All model assumptions, validation, and details for the thermal and biological growth models are presented in Quinn et al. [30].

Weather Data

Historical hourly meteorological weather data serves as a primary input to the thermal and biological growth model. EnergyPlus Weather data files (EPW) from the Department of Energy (DOE) and the American Society of Heating, Refrigeration and Air Conditioning Engineers (ASHRAE) were obtained for 4,388 global locations [39, 40]. The EPW data files are derived from hourly weather data originally archived at the U.S. National Climatic Data Center. The databases contain weather observations of wind speed and direction, sky cover, visibility, ceiling height, dry-bulb temperature, dew-point temperature, atmospheric pressure, and liquid precipitation for a minimum of 12 and maximum of 25 years depending on location. Observed data was
processed statistically to generate standard one year data sets representative of historically observed weather data or EPW data files [40]. Hourly solar radiation is calculated through a model based on the sun-earth geometry, observed cloud cover, relative humidity, and wind speed [40]. Total solar radiation is converted to PAR for biological modeling [41]. The climatic variables from the EPW data files are used as the primary inputs to the thermal and biological growth models.

**Simulation Architecture**

**Harvest Technique and Reporting Metrics**

The biological growth model was operated with a time-based harvest schedule. Cultures were inoculated at a density of 1 g L\(^{-1}\) at hour zero and harvested after 160 hours or a biomass density of 3 g L\(^{-1}\) (whichever occurs first). Each harvest retains biomass that is used as inoculum for the next growth cycle. This harvest technique is representative of the operation of the research and development and pilot plant facility used for model validation [11]. The productivity potentials reported are presented based on a photosynthetic area and do not include infrastructure required for large-scale cultivation such as roads, processing facilities, or mechanical systems. Additionally, the reported potentials represent the biological lipid production and do not include losses associated with lipid extraction and conversion to fuel. The scalability assessment assumes a packing factor of 0.8 [27] to account for facility infrastructure and a 90% efficiency for the extraction process and a 90% efficiency for the conversion of oil to fuel [42, 43] to more accurately represent the microalgae to biofuel process.
**Temperature Sensitivity**

The effects of temperature on potential lipid productivity were further examined as a part of this study. Typical lab based studies regulate temperature to optimum conditions to maximize production or remove it as a variable [44-46]. However, temperature control at large-scale represents an economically unfavorable option based on the amount of raw-energy required [47]. Some pilot plant facilities are utilizing waste heat from co-located industrial processing facilities or raw energy to maintain optimum temperatures, but resource limits the scalability [11, 48]. In an effort to understand the impacts of maintaining optimum temperature, the temperature efficiency in the biological growth model was fixed to optimum conditions \((\rho_T=1)\). The corresponding biomass and lipid productivities are compared to the non-ideal case. Realistically, temperature will not be controlled at large-scale based on resource and economic limitations. Large-scale systems could use waste heat to support inoculation systems thus the results from the optimum scenario can be applied in select modeling applications or pilot-plant siting.

**Productivity Variability**

Results from a harmonization effort from ANL et al. [16] show economic viability and environmental impact associated with a microalgae biofuel system decline with increasing variability. Previous assessments have focused on reporting the annual average productivity potential and failed to investigate and report the minimum and maximum productivities throughout the year [30, 49]. The majority of modeling efforts do not have the fidelity to investigate variability as a fixed growth rate and lipid percentage is typically assumed. Productivity in a biological system is dependent on resource availability, specifically sunlight. Global positioning over the course of a year will cause variability in this resource and impact
growth. In locations with freezing temperatures, the productivity will drop to zero. This study presents a variability assessment for each simulated location with results for the minimum and maximum productivities based on monthly statistics. Variability results were extrapolated to annual values for comparative purposes. The maximum lipid productivity was found by extrapolating the month that produced the highest yield. Calculations of minimum yields were complicated due to locations with months which do not produce a harvest due to the freezing of the water basin. The productivity results from the month with the lowest yield that contained at least one full harvest coming out of a water basin thaw, was extrapolated to one year for the minimum yield.

**Geographical Information System (GIS)**

The results from the 4388 simulations were interpolated over the globe using a surface interpolation. Surface interpolation is a methodology used to develop values for unmeasured locations using measured or simulated data values from known neighboring locations [31]. Many Interpolation algorithms have been developed to account for spatial and statistical differences of data; however, the central principal of surface interpolation algorithms is the application of the weighted sum of data values from nearby locations to produce an estimated value for any given location within the maximum spatial constraints of the data. Interpolation algorithms have been used to calculate surface values that depict current and historic climate values [50, 51], extraction and estimates land surface features [52], evaluate the subsurface dispersion of spilled pollutants [53, 54], and to evaluate the impacts of noise from aircraft and industry on the surrounding environment [55].

This research used the surface interpolation tools available in the Esri ArcGIS 10.2 Geostatistical Analyst extension. ArcGIS was also used to reduce the land cover from the
GLC2000 database to obtain the non-arable land statistics for each country around the world [56]. The classifications found in the GLC2000 database are tree cover (broadleaved, evergreen), tree cover (broadleaved, deciduous, closed), tree cover (broadleaved, deciduous, open), tree cover (needle-leaved, evergreen), tree cover (needle-leaved, deciduous), tree cover (mixed leaf type), tree cover (regularly flooded, fresh water), tree cover (regularly flooded, saline water), mosaic (tree cover, other natural vegetation), tree cover (burnt), shrub cover (closed-open, evergreen), shrub cover (closed-open, deciduous), herbaceous cover (closed-open), sparse herbaceous or sparse shrub cover, regularly flooded shrub and/or herbaceous cover, cultivated and managed areas, mosaic (cropland, tree cover, other natural vegetation), mosaic (cropland, shrub, or grass cover), bare areas, water bodies, snow/ice, and artificial surfaces. Of these 22 land classifications available sparse herbaceous or sparse shrub cover and bare areas were used for the non-arable land. The Geostatistical Analyst extension provides a suite of spatial modeling and data exploration utilities that may be used to develop surface models based on known or simulated spatial point values. This tool set facilitates the exploration and evaluation of data quality, and uses a host of surface interpolation models to develop the most appropriate spatial model for the available data. It also provides analysis methods for deterministic, geostatistical, and barrier influenced interpolation. Included within these broad categories are specific interpolation functions such as Inverse Distance Weighting, Radial Basis Functions, Kriging and CoKriging, Empirical Bayesian Kriging, and Kernel smoothing.

The Ordinary Kriging method was used for exploration of the data values for biomass and lipids due to the spatial point sample data of this analysis as presented in Fig. A.1 in the Appendix. As one of the most common and widely used interpolation methods, Ordinary Kriging does not assume that the mean of the data is constant over the entire spatial domain, but rather that the mean is constant in local spatial neighborhoods [31]. This further helps
compensate for the impacts of clustered groups by assigning points within a clustered group less overall weight than isolated data points. This is an important consideration given the data in this analysis where points which tended to be highly clustered in more densely populated regions and where biomass and lipid production tend to be higher in areas nearer the equator and decrease towards the Polar Regions. It should be noted that there are some cases where elevation and climate impact the general trend; however, the consistency of the data allows for the use of Ordinary Kriging methods.

The Geostatistical Analyst extension requires the input of spatially explicit data with associated point values. Once the data is loaded and the appropriate associated variable identified, the utility provides a series of data evaluation screens and outputs that describe an optimal model based on the semivariogram and covariance graphs and maps. The number of lags and lag size are determined for the Ordinary Kriging model and optimization is conducted as necessary. The lag is the sample distance used to group, or bin, pairs of points and using the optimal number of lags enables scale-dependent spatial correlations [31]. In the case of this analysis, and the global extent of the data the model selected an appropriate lag size of 7.3 with the correlating number of lags at 12. Manual verification of the lag value was conducted to verify the Geostatistical Analyst results.

After the appropriate analysis variables were selected, the neighborhood and sector type were evaluated. Both a standard neighborhood and smooth neighborhood type for surface interpolation were assessed. Standard neighborhood options assign weights based on distance from a specific target location assigned by the analyst. Alternatively, the smooth neighborhood function adjusts the weights using a sigmoidal function defined by a smoothing factor. Based on validation tests conducted, a smooth neighborhood type using a smoothing factor of 0.5 was selected. This produced smother global isolines, removing excessive stair-step effects in areas
of less sample data. The sector type is simply the geometry of the neighborhood in which the algorithm assesses nearby point values. The Geostatistical Analyst default of four sectors with a 45° offset was selected. Other options were tested with no measurable change in results.

Model validation outputs including Predicted Error, Standardized Error, and Normal QQ Plots for each surface interpolation model to verify suitable performance were evaluated. Manual accuracy testing was also conducted by withholding a random selection of 25 percent of the global data points, performing an interpolation analysis, and then sampling and regressing the observed data against the predicted interpolated surface values. The $R^2$ between the interpolated and observed data was 0.97, thus further sustaining the conclusion that the surface interpolation methods were adequately performed.
RESULTS AND DISCUSSION

Global Productivity Potential
and Variability

Hourly biomass and lipid productivity results from simulation locations were averaged for an annual geographically resolved result. Results from the 4,388 simulation locations were surface interpolated to produce the dynamic map illustrating the lipid productivity potential from microalgae across the globe, Fig. 5. As expected, locations situated in the northern and southern parts of the globe where the light intensity and temperature are lower, result in decreased lipid productivity compared to regions more centrally located. Although locations around the equator are typically considered to be optimum cultivation locations due to annual temperature stability, results show these locations do not necessarily produce the largest lipid yields. This is due to other climatic phenomenon that impact biological growth. A detailed comparison of four countries, India, China, Brazil, and Australia, is presented in Fig. 6. Manaus, Brazil is located closer to the equator than Alice Springs, Australia but experiences more rain and cloud cover and as a result, Manaus produces a lower lipid yield of 18.9 m³·ha⁻¹·yr⁻¹ (corresponding to 10.2 g·m⁻²·day⁻¹) compared to 24.2 m³·ha⁻¹·yr⁻¹ (corresponding to 13.1 g·m⁻²·day⁻¹) in the Alice Springs. India and China are neighboring countries but India has better average lipid yields primarily due to the temperature difference between the two countries.
Fig. 5  World map of the lipid productivity potential from microalgae

Fig. 6  Lipid productivity maps for India, China, Brazil, and Australia
Results also illustrate a non-linear relationship between the biomass growth and lipid productivity. A biomass yield of 6,720 kg·ha\(^{-1}\)·yr\(^{-1}\) is found in both Ostrov-Vrangelja, Russia and Lomnicky-Stit, Slovakia with corresponding lipid productivities of 3.37 m\(^3\)·ha\(^{-1}\)·yr\(^{-1}\) and 3.26 m\(^3\)·ha\(^{-1}\)·yr\(^{-1}\) respectively. The 3.3% difference in lipid production can be attributed to the biological model capturing non-trivial biological effects including nitrogen quota, inoculation lipid content, light availability, and temperature. The dynamic global map of the biomass productivity is presented in Fig. A.2 of the Appendix.

Maximum average annual yields and the annual variability (high and low) from 11 locations across the globe are presented in Table 1. Table A.1 in the Appendix contains the maximum, minimum, and average yields of all 4388 global locations. The variability changes from location to location primarily due to climatic diversity. Locations with low variability and large annual yields represent areas where cultivation typically does not shut down due to cold conditions and operates year round. Learmonth, Australia has a higher annual yield but a large variability compared to Cali, Columbia. The economic impacts associated with variability suggest Cali, Columbia would be a superior location even though it has a slightly lower annual average productivity compared to Learmonth, Australia [16]. The maximum, average, and minimum lipid productivity maps are presented in Fig. A.3, Fig. A.4, and Fig. A.5 in the Appendix. The lipid scale for these maps ranges from 0 to 34.25 m\(^3\)·ha\(^{-1}\)·yr\(^{-1}\) so a direct comparison can be drawn from the three maps.
Table 1  Average microalgae lipid yields in m$^3$·ha$^{-1}$·yr$^{-1}$ of various regions around the world with respective high and low variability

<table>
<thead>
<tr>
<th>Location</th>
<th>Maximum Lipid Yield*</th>
<th>Average Lipid Yield</th>
<th>Minimum Lipid Yield*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kisumu, Kenya</td>
<td>29.66</td>
<td>27.42</td>
<td>23.42</td>
</tr>
<tr>
<td>Learmonth, Australia</td>
<td>33.43</td>
<td>25.88</td>
<td>16.78</td>
</tr>
<tr>
<td>Trivandrum, India</td>
<td>29.07</td>
<td>24.91</td>
<td>16.36</td>
</tr>
<tr>
<td>Cali, Columbia</td>
<td>27.62</td>
<td>24.43</td>
<td>21.18</td>
</tr>
<tr>
<td>Hawaii, United States</td>
<td>28.40</td>
<td>23.68</td>
<td>17.70</td>
</tr>
<tr>
<td>Santiago de Cuba, Cuba</td>
<td>26.31</td>
<td>23.50</td>
<td>18.66</td>
</tr>
<tr>
<td>Yuma, AZ, United States</td>
<td>32.21</td>
<td>21.61</td>
<td>8.13</td>
</tr>
<tr>
<td>Poltavka, Russia</td>
<td>27.63</td>
<td>12.66</td>
<td>5.49</td>
</tr>
<tr>
<td>Bagaskar, Finland</td>
<td>26.31</td>
<td>9.24</td>
<td>6.57</td>
</tr>
<tr>
<td>Punta Arenas, Chile</td>
<td>22.16</td>
<td>9.26</td>
<td>5.64</td>
</tr>
<tr>
<td>Rio Gallegos, Argentina</td>
<td>20.15</td>
<td>9.02</td>
<td>6.74</td>
</tr>
</tbody>
</table>

* Monthly data extrapolated to annual yields for comparison

Locations found in the northern and southern hemispheres such as Poltavka, Bagaskar, Punta Arenas, and Rio Gallegos, whose corresponding yields are 6.8, 5.0, 5.1, and 4.9 g·m$^{-2}$·day$^{-1}$ respectively, demonstrate a large variability and low annual yield due to large swings in climate as a result of geographic location with the colder climate experienced in these locations forcing a winter shut down. Yuma, Arizona, USA, a desert location with a corresponding yield of 11.7 g·m$^{-2}$·day$^{-1}$ and which does not shut down in the winter time, reaches temperatures that far exceed the optimum temperature in the summer time and results in a lower yield than Kisumu, Kenya, whose average annual culture temperature stays more closely to optimum conditions with a corresponding yield of 14.8 g·m$^{-2}$·day$^{-1}$. Deviation from optimum temperature in either direction negatively impacts the biomass and lipid productivity of the system.
Temperature Sensitivity

A temperature sensitivity analysis demonstrates the importance of considering the effect of temperature on productivity. Pilot plant facilities are currently utilizing waste heat or raw energy to maintain culture temperature in cold months to sustain production [11]. Large-scale implementation of this technique is limited due to resource availability and the economics associated with raw energy. Waste heat and raw energy for maintaining temperature will be limited to inoculum and small-scale pilot plant facilities.

Results for simulations were ideal temperature is maintained are presented in Fig. 7. Compared to the non-ideal temperature results presented in the baseline scenario, Fig. 5, the lipid productivity across the globe is dramatically higher illustrating the non-trivial effect maintaining optimum temperature has on microalgae growth and lipid productivity. The locations situated in the northern and southern most parts of the globe are impacted the most when comparing ideal temperature results to real temperature lipid productivity. Bryce Canyon Utah, for example, has an ideal and non-ideal lipid yield of 22 m³·ha⁻¹·yr⁻¹ and 10.7 m³·ha⁻¹·yr⁻¹, corresponding to a 106% increase for maintaining optimum temperature. The dramatic increase is predominantly due to extending the growing season from 214 days to 365 days. Integrating waste heat from industrial processing plants makes it feasible for locations to achieve improved lipid productivity but heat resource will ultimately limit the scalability.
The ideal lipid productivity presented in Fig. 7 does not include temporal resolution. In the northern hemisphere maximum yields will be attained from May to July while in the southern hemisphere minimum yields will be achieved during these months.

**Scalability Assessment**

A scalability assessment of the feasibility of microalgae based biofuels to meet various energy demands of global nations was performed. The lipid productivity results from this study are combined with land availability data for a geographic realized assessment. The assessment demonstrates the potential microalgae has to supplement energy, reducing demand on traditional fossil oil and the yield advantage microalgae hold over traditional terrestrial crops such as soybeans.

The quantity of agricultural land available [57] and oil consumption for various regions [58] were used to evaluate the potential impact of microalgae. The scalability assessment
includes a 90% efficiency for the extraction of lipids from the biomass, a 90% efficiency for the conversion of oil to fuel, and a 0.8 packing factor to more accurately represent the microalgae to biofuel process at industrial scale. The amount of agricultural land required to supplement 100% of the transportation fuel consumed in each region through replacement by microalgae is directly compared to the requirements for soybean based fuels, Table 22. Assumptions for soybean yields are based on geographically based yield data and include the same efficiency assumptions for extraction and conversion as for microalgae based processing, but do not include a packing factor [5, 59-62].

<table>
<thead>
<tr>
<th>Location</th>
<th>Agricultural Land Available (hectare)</th>
<th>Oil Yield (m³·ha⁻¹·yr⁻¹)</th>
<th>Agricultural Land Required</th>
<th>Microalgae</th>
<th>Soybeans</th>
<th>Microalgae</th>
<th>Soybean</th>
</tr>
</thead>
<tbody>
<tr>
<td>Brazil</td>
<td>2.35E+08</td>
<td>20.9</td>
<td>0.43</td>
<td>5%</td>
<td>177%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Canada</td>
<td>8.46E+07</td>
<td>7.0</td>
<td>0.50</td>
<td>32%</td>
<td>352%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>China</td>
<td>5.09E+08</td>
<td>12.5</td>
<td>0.36</td>
<td>15%</td>
<td>394%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Japan</td>
<td>4.67E+06</td>
<td>12.6</td>
<td>0.23</td>
<td>665%</td>
<td>29,000%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>United States</td>
<td>4.36E+08</td>
<td>12.7</td>
<td>0.56</td>
<td>27%</td>
<td>484%</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2 shows it is feasible to supplement the petroleum oil consumption of Brazil, Canada, China, and United States based on microalgae productivity potential and agricultural land. Regions such as Japan cannot practically replace their oil needs through microalgae with over 100% of their agricultural land required. Soybean requires an average of 27 times more agricultural land than microalgae in the regions examined. Displacement of agricultural land for microalgae cultivation will impact food markets and is not considered in this scalability assessment.
An advantage microalgae cultivation holds over traditional terrestrial crops is the utilization of poor quality land for cultivation. To more accurately perform a scalability assessment for microalgae, non-arable land statistics were used, based on Global Land Cover 2000 data set [56]. The amount of non-arable land needed to supplement 100% of the transportation fuel consumed in each region is presented in Table 3. The same assumptions in terms of processing efficiencies and packing factors are included in the calculations and corresponding results as previously assumed above.

Table 3  Amount of non-arable land needed to supplement 100% of oil consumption for different regions around the world based on replacement by microalgae

<table>
<thead>
<tr>
<th>Location</th>
<th>Non-arable Land Available (hectare)</th>
<th>Biodiesel Produced (l·yr⁻¹)</th>
<th>Transportation Fuel Consumption (l·yr⁻¹)</th>
<th>Non-arable Land Required</th>
</tr>
</thead>
<tbody>
<tr>
<td>Brazil</td>
<td>2.06E+07</td>
<td>2.79E+11</td>
<td>1.46E+11</td>
<td>52%</td>
</tr>
<tr>
<td>Canada</td>
<td>3.62E+08</td>
<td>1.65E+12</td>
<td>1.21E+11</td>
<td>7%</td>
</tr>
<tr>
<td>China</td>
<td>2.22E+08</td>
<td>1.79E+12</td>
<td>6.32E+11</td>
<td>35%</td>
</tr>
<tr>
<td>Japan</td>
<td>9.16E+04</td>
<td>7.49E+08</td>
<td>2.54E+11</td>
<td>34,000%</td>
</tr>
<tr>
<td>United States</td>
<td>7.03E+07</td>
<td>5.80E+11</td>
<td>9.53E+11</td>
<td>164%</td>
</tr>
</tbody>
</table>

Results from Table 3 show it is feasible for Brazil, Canada, and China to replace their petroleum oil consumption based on the utilization of poor quality land for microalgae cultivation and processing. Japan and United States, however, require over 100% of their non-arable land to replace their transportation fuel consumption through microalgae. A table representing the amount of non-arable land needed to supplement 100% of oil consumption for other countries around the world based on microalgae yield from this study is presented in Table A.2 in the Appendix.
A number of countries around the world have implemented biodiesel goals. Brazil, China, and United States have biodiesel goals of $7.30E+09$, $1.40E+11$, and $2.86E+11$ liters per year respectively [30, 63, 64]. Using the same efficiencies and packing factors as the previous scalability assessments, the amount of non-arable land required in Brazil, China, and United States is 3%, 8%, and 49%, respectively. It is feasible for these three countries to achieve their biodiesel goals through microalgae and non-arable land use. Non-arable land utilization for microalgae production provides a benefit over traditional terrestrial crops such as soy which compete with food crops for agricultural land.

**Literature Comparison**

The large uncertainty of microalgae productivity in literature makes it difficult to perform accurate lifecycle and technoeconomic assessments, as productivity represents the functional unit of these studies and dramatically impacts results. The majority of lifecycle and technoeconomic assessments in literature use different productivities resulting in a large range of end results regardless of processing differences.

Modeling assumptions in term of yield for various lifecycle, technoeconomic, and scalability assessments are compared to the results of this study. For direct comparisons between studies and the results of this work, calculations were performed to harmonize the values in the literature surveyed with details of those calculations presented in Table 4.
Table 4  Table comparing reported productivity potentials (some calculations performed for comparison purposes with assumptions detailed in notes column) from various sources

<table>
<thead>
<tr>
<th>Source</th>
<th>Oil Yield (m³ha⁻¹yr⁻¹)</th>
<th>Article Type</th>
<th>Purpose of Scaling</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ramachandra et al.ᵃ [7]</td>
<td>2.3</td>
<td>Research-Model</td>
<td>Microalgae Potential</td>
<td>8.8-24.6% oil, Wastewater</td>
</tr>
<tr>
<td>Pateᵇ [8]</td>
<td>4.7</td>
<td>Research-Model</td>
<td>Modeling Effort</td>
<td></td>
</tr>
<tr>
<td>Lam et al.ᵇ [9]</td>
<td>5.2</td>
<td>Review</td>
<td>Microalgae Potential</td>
<td>30% oil assumed, p=860kg m⁻³</td>
</tr>
<tr>
<td>Resurreccion et al.ᵇ [10]</td>
<td>5.5</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>13.4-32.4% oil, ORP, PBR</td>
</tr>
<tr>
<td>Ramachandra et al.ᵇ [7]</td>
<td>7.6</td>
<td>Research-Data</td>
<td>Microalgae Potential</td>
<td>8.8-24.6% oil, Wastewater</td>
</tr>
<tr>
<td>Scott et al.ᵇ [6]</td>
<td>8.2</td>
<td>Review</td>
<td>Microalgae Potential</td>
<td>46% oil</td>
</tr>
<tr>
<td>Lam et al.ᵇ [9]</td>
<td>8.7</td>
<td>Review</td>
<td>Microalgae Potential</td>
<td>30% oil assumed, p=860kg m⁻³</td>
</tr>
<tr>
<td>Vasudevan et al.ᵇ [12]</td>
<td>9.0</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>ORP</td>
</tr>
<tr>
<td>Guieysse et al.ᵇ [13]</td>
<td>11.8</td>
<td>Research-Model</td>
<td>Modeling Effort</td>
<td>25% oil assumed, ORP</td>
</tr>
<tr>
<td>Clarens et al.ᵇ [14]</td>
<td>11.8</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>9.45 g m⁻² d⁻¹, 30% oil assumed, ORP</td>
</tr>
<tr>
<td>Schenk et al.ᵇ [15]</td>
<td>12.0</td>
<td>Review</td>
<td>Microalgae Potential</td>
<td>10 g m⁻² d⁻¹, 30% TAG</td>
</tr>
<tr>
<td>ANL;NREL;PNL [16]</td>
<td>13.1</td>
<td>Research-Model</td>
<td>Modeling Effort</td>
<td>25% oil assumed, p=920kg m⁻³</td>
</tr>
<tr>
<td>Clarens et al.ᵇ [14]</td>
<td>16.1</td>
<td>Research-Model</td>
<td>Economic-Modeling Effort</td>
<td>12.9 g m⁻² d⁻¹, 30% oil assumed, ORP</td>
</tr>
<tr>
<td>Lardon et al.ᵇ [17]</td>
<td>18.0</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>24.75 g m⁻² d⁻¹, 17.5% oil, ORP</td>
</tr>
<tr>
<td>Campbell et al.ᵇ [18]</td>
<td>18.7</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>15 g m⁻² d⁻¹, 30% oil assumed, ORP</td>
</tr>
<tr>
<td>Davis et al.ᵇ [19]</td>
<td>23.4</td>
<td>Research-Model</td>
<td>Economic-Modeling Effort</td>
<td>25 g m⁻² d⁻¹, 20% oil, p=880kg m⁻³, ORP</td>
</tr>
<tr>
<td>Moheimani et al.ᵇ [20]</td>
<td>26.7</td>
<td>Research-Data</td>
<td>Microalgae Potential</td>
<td>PBR, Growth Data, p=860kg m⁻³</td>
</tr>
<tr>
<td>Guieysse et al.ᵇ [13]</td>
<td>27.3</td>
<td>Research-Model</td>
<td>Modeling Effort</td>
<td>25% oil assumed, ORP</td>
</tr>
<tr>
<td>Resurreccion et al.ᵇ [10]</td>
<td>30.3</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>13.4-32.4% oil, ORP, PBR</td>
</tr>
<tr>
<td>Lardon et al.ᵇ [17]</td>
<td>30.8</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>19.25 g m⁻² d⁻¹, 38.5% oil, ORP</td>
</tr>
<tr>
<td>Clarens et al.ᵇ [21]</td>
<td>31.1</td>
<td>Letter</td>
<td>Microalgae Potential</td>
<td>25 g m⁻² d⁻¹, 20% oil, p=880kg m⁻³, ORP</td>
</tr>
<tr>
<td>Rodolfi et al. [23]</td>
<td>34.1</td>
<td>Research-Model</td>
<td>Economic-Modeling Effort</td>
<td>location with 20 MJ m⁻² d⁻¹</td>
</tr>
<tr>
<td>Quinn et al.ᵇ [11]</td>
<td>36.3</td>
<td>Research-Data</td>
<td>Microalgae Potential</td>
<td>PBR, Growth Data</td>
</tr>
<tr>
<td>Campbell et al.ᵇ [18]</td>
<td>37.3</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>30 g m⁻² d⁻¹, 30% oil assumed, ORP</td>
</tr>
<tr>
<td>Wijffels and Barbosa [2]</td>
<td>40.0</td>
<td>Perspective</td>
<td>Microalgae Potential</td>
<td>3% solar efficiency, 50% oil</td>
</tr>
<tr>
<td>Scott et al.ᵇ [6]</td>
<td>40.0</td>
<td>Review</td>
<td>Microalgae Potential</td>
<td>50% oil</td>
</tr>
<tr>
<td>Williams and Laurens [24]</td>
<td>40.7</td>
<td>Opinion</td>
<td>Microalgae Potential</td>
<td>28 g m⁻² d⁻¹, 35% oil</td>
</tr>
<tr>
<td>Yeang et al.ᵇ [25]</td>
<td>46.0</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>40% oil assumed, p=860kg m⁻³, ORP</td>
</tr>
<tr>
<td>Liao et al. [26]</td>
<td>46.1</td>
<td>Letter</td>
<td>Microalgae Potential</td>
<td>25 g m⁻² d⁻¹, 50% oil, p=880kg m⁻³, ORP</td>
</tr>
<tr>
<td>Chistiᵇ [21]</td>
<td>51.9</td>
<td>Research-Model</td>
<td>LCA-Modeling Effort</td>
<td>25 g m⁻² d⁻¹, 50% oil, PBR</td>
</tr>
<tr>
<td>Batan et al. [27]</td>
<td>51.9</td>
<td>Research-Model</td>
<td>Microalgae Potential</td>
<td>30% oil, PBR</td>
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</table>
Some authors reported a range of productivity potential

The low values are repeated in this table

The high values are repeated in this table

A map of the potential lipid productivity was created using the same scale as the lipid productivity results found in literature, Fig. 8. Locations with the highest lipid productivities of this study fall in the lower third of literature results while locations with the lowest productivities fall in the lower sixth. These results show that most of the productivity potentials reported in literature and integrated into the lifecycle, technoeconomic, and resource assessments of microalgae based biofuels are making optimistic assumptions in terms of productivity potential. Results from the optimistic productivity assumptions are favorably skewing the economic and environmental impact of microalgae based biofuels. The validated model in this study demonstrates that lipid productivity is a dynamic variable that cannot be assumed constant temporally or geographically. Future assessments of the microalgae to biofuels production process require site specific assessments that incorporate geographically realized biological growth modeling to improve the validity and accuracy of results.
Fig. 8  Oil yield assumptions found in lifecycle, technoeconomic, and scalability assessments compared with the results of this study presented as a global map with a scale representative of the literature values (2.3 – 140 m³·ha⁻¹·yr⁻¹). The black dashed line represents the world average lipid productivity of 17.4 m³·ha⁻¹·yr⁻¹ corresponding to 9.4 g·m⁻²·day⁻¹. Due to a range of productivities reported in some studies a (+) represents the low end of the range and (++) represents the high end.
CONCLUSION

Oil consumption is rising at a rate that is proving to be unsustainable as reflected in the recent volatility in fuel prices. Alternatives to fossil fuels such as microalgal based biofuels are being investigated due to promising productivity potential along with other sustainable advantages compared to traditional terrestrial feedstocks. However, there is a large uncertainty in the current lipid productivity potential from microalgae. This study uses a validated outdoor thermal and biological growth model to determine the lipid productivity potential of microalgae around the world through the integration of hourly meteorological data at 4,388 sites. Results are spatially interpolated to generate a global map. An analysis of biomass yield compared to lipid percentage, the effect of temperature and variability on microalgal growth was performed showing lipid is not a linear function of biomass, temperature plays a critical role in yield, and variability must be considered in large-scale assessments. The lipid productivity falls into the lower third of what is found in lifecycle, technoeconomic, and scalability assessments currently found in literature with the highest global yields based on results from this study ranging between 24-27 m$^3$·ha$^{-1}$·yr$^{-1}$. A scalability analysis incorporating GIS land classification statistics and results from this study show many regions such as Brazil, China, and United States can reach their biodiesel goals through microalgae with the utilization of 3%, 8%, and 49% of non-arable land, respectively. Future lifecycle, technoeconomic, and scalability assessments require the integration of geographically and temporally resolved biological growth modeling for increased fidelity.
FUTURE WORK

Future research based on this project is recommended. A geographically realized LCA and TEA can be completed using the GREET life-cycle model from Argonne National Laboratory coupled with the productivity results obtained from this project to demonstrate the energetic and economic feasibility of biofuel from microalgae. Further GIS work based on resource availability and land classifications can be performed to evaluate the feasibility of microalgae cultivation sites around the world.
REFERENCES


APPENDIX
Global maps detailing the simulation points and corresponding error (Fig. A.1), the biomass productivity (Fig. A.2), and variability (Fig. A.3, Fig. A.4, Fig. A.5) are presented below. The biomass map provides for a direct comparison with Fig. 5 to evaluate the nonlinear effect of biomass growth and lipid productivity. The variability maps present a further evaluation of the effects of annual variability with respect to climatic variables.

Fig. A.1  Global map of the 4388 locations coupled with standard error assessment. The standard error verifies that where the point locations are located, and indeed denser, yields a smaller error, signifying better interpolation of surface values.
Fig. A.2  World map of the biomass productivity potential from microalgae

Fig. A.3  Global map of the maximum lipid productivity potential from microalgae
Fig. A.4  Global map of the average lipid productivity potential from microalgae

Fig. A.5  Global map of the minimum lipid productivity potential from microalgae
A table of the variability assessment for all 4,388 global locations is presented below. This table shows that each location around the world has a site specific maximum, average, and minimum variability so when performing a LCA or TEA of the biofuel from microalgae process, the assessment must also be site specific.

Table A.1  Average microalgae lipid yields in m$^3$-ha$^{-1}$-yr$^{-1}$ of various regions around the world with respective high and low variability

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* Monthly data extrapolated to annual yields for comparison
The table below represents the amount of non-arable land required to supply transportation energy supply needs of different countries around the world.

**Table A.2 Amount of non-arable land needed to supplement 100% of oil consumption for different regions around the world based on replacement by microalgae**

<table>
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<tr>
<th>Country</th>
<th>Lipid Productivity (m⁻³·ha⁻¹·yr⁻¹)</th>
<th>Non-arable land (ha)</th>
<th>Biodiesel Produced (liters a year)</th>
<th>Transportation Fuel Consumption (liters a year)</th>
<th>Non-arable Land Required</th>
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<td>Algeria</td>
<td>19.7</td>
<td>1.95E+08</td>
<td>2.49E+12</td>
<td>1.94E+10</td>
<td>1%</td>
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<td>Argentina</td>
<td>16.9</td>
<td>9.96E+07</td>
<td>1.09E+12</td>
<td>3.28E+10</td>
<td>3%</td>
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<tr>
<td>Australia</td>
<td>22.2</td>
<td>2.74E+08</td>
<td>3.94E+12</td>
<td>5.43E+10</td>
<td>1%</td>
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<td>10.9</td>
<td>4.32E+05</td>
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<td>1.44E+10</td>
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<td>1.11E+06</td>
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<td>1.17E+12</td>
<td>4.09E+10</td>
<td>3%</td>
</tr>
<tr>
<td>Finland</td>
<td>8.6</td>
<td>5.65E+05</td>
<td>3.13E+09</td>
<td>1.06E+10</td>
<td>338%</td>
</tr>
<tr>
<td>France</td>
<td>12.2</td>
<td>4.20E+05</td>
<td>3.33E+09</td>
<td>9.41E+10</td>
<td>2.827%</td>
</tr>
<tr>
<td>Germany</td>
<td>9.9</td>
<td>2.16E+05</td>
<td>1.38E+09</td>
<td>1.30E+11</td>
<td>9.413%</td>
</tr>
<tr>
<td>Greece</td>
<td>16.9</td>
<td>6.20E+03</td>
<td>6.80E+07</td>
<td>1.79E+10</td>
<td>26,332%</td>
</tr>
<tr>
<td>India</td>
<td>20.4</td>
<td>6.08E+06</td>
<td>8.04E+10</td>
<td>2.00E+11</td>
<td>248%</td>
</tr>
<tr>
<td>Indonesia</td>
<td>20.3</td>
<td>4.18E+06</td>
<td>5.51E+10</td>
<td>8.33E+10</td>
<td>151%</td>
</tr>
<tr>
<td>Iran</td>
<td>19.0</td>
<td>1.18E+08</td>
<td>1.45E+12</td>
<td>1.04E+11</td>
<td>7%</td>
</tr>
<tr>
<td>Ireland</td>
<td>9.1</td>
<td>2.00E+03</td>
<td>1.18E+07</td>
<td>7.21E+09</td>
<td>61,254%</td>
</tr>
<tr>
<td>Israel</td>
<td>20.7</td>
<td>1.12E+06</td>
<td>1.50E+10</td>
<td>1.58E+10</td>
<td>105%</td>
</tr>
<tr>
<td>Italy</td>
<td>14.7</td>
<td>7.86E+05</td>
<td>7.48E+09</td>
<td>7.47E+10</td>
<td>998%</td>
</tr>
<tr>
<td>Japan</td>
<td>12.6</td>
<td>9.16E+04</td>
<td>7.49E+08</td>
<td>2.54E+11</td>
<td>33,883%</td>
</tr>
<tr>
<td>Kazakhstan</td>
<td>12.3</td>
<td>2.03E+08</td>
<td>1.62E+12</td>
<td>1.49E+10</td>
<td>1%</td>
</tr>
<tr>
<td>Kuwait</td>
<td>21.1</td>
<td>1.44E+06</td>
<td>1.97E+10</td>
<td>2.43E+10</td>
<td>123%</td>
</tr>
<tr>
<td>Lithuania</td>
<td>10.4</td>
<td>8.98E+04</td>
<td>6.05E+08</td>
<td>3.02E+09</td>
<td>500%</td>
</tr>
<tr>
<td>Malaysia</td>
<td>19.8</td>
<td>7.06E+04</td>
<td>9.05E+08</td>
<td>3.47E+10</td>
<td>3,828%</td>
</tr>
<tr>
<td>Mexico</td>
<td>20.8</td>
<td>1.18E+07</td>
<td>1.59E+11</td>
<td>1.08E+11</td>
<td>68%</td>
</tr>
<tr>
<td>New Zealand</td>
<td>14.2</td>
<td>9.59E+05</td>
<td>8.80E+09</td>
<td>8.14E+09</td>
<td>93%</td>
</tr>
<tr>
<td>Norway</td>
<td>6.6</td>
<td>1.67E+07</td>
<td>7.11E+10</td>
<td>1.26E+10</td>
<td>18%</td>
</tr>
<tr>
<td>Pakistan</td>
<td>20.1</td>
<td>2.70E+07</td>
<td>3.51E+11</td>
<td>2.33E+10</td>
<td>7%</td>
</tr>
<tr>
<td>Peru</td>
<td>19.5</td>
<td>1.65E+07</td>
<td>2.09E+11</td>
<td>1.12E+10</td>
<td>5%</td>
</tr>
<tr>
<td>Country</td>
<td>Population</td>
<td>Population Growth</td>
<td>GDP</td>
<td>GDP Growth</td>
<td></td>
</tr>
<tr>
<td>-------------------------</td>
<td>------------</td>
<td>-------------------</td>
<td>-----</td>
<td>------------</td>
<td></td>
</tr>
<tr>
<td>Philippines</td>
<td>19.2</td>
<td>3.16E+05</td>
<td>3.92E+09</td>
<td>1.51E+10</td>
<td>385%</td>
</tr>
<tr>
<td>Poland</td>
<td>10.2</td>
<td>6.08E+05</td>
<td>4.02E+09</td>
<td>2.92E+10</td>
<td>725%</td>
</tr>
<tr>
<td>Portugal</td>
<td>17.6</td>
<td>1.43E+05</td>
<td>1.63E+09</td>
<td>1.27E+10</td>
<td>776%</td>
</tr>
<tr>
<td>Qatar</td>
<td>21.2</td>
<td>9.50E+05</td>
<td>1.31E+10</td>
<td>9.30E+09</td>
<td>71%</td>
</tr>
<tr>
<td>Romania</td>
<td>12.4</td>
<td>6.49E+04</td>
<td>5.22E+08</td>
<td>1.02E+10</td>
<td>1.960%</td>
</tr>
<tr>
<td>Russian Federation</td>
<td>7.8</td>
<td>2.41E+08</td>
<td>1.22E+12</td>
<td>1.72E+11</td>
<td>14%</td>
</tr>
<tr>
<td>Saudi Arabia</td>
<td>21.5</td>
<td>1.67E+08</td>
<td>2.33E+12</td>
<td>1.51E+11</td>
<td>6%</td>
</tr>
<tr>
<td>Slovakia</td>
<td>11.3</td>
<td>9.40E+03</td>
<td>6.87E+07</td>
<td>4.07E+09</td>
<td>5.924%</td>
</tr>
<tr>
<td>South Africa</td>
<td>20.1</td>
<td>1.63E+07</td>
<td>2.13E+11</td>
<td>3.13E+10</td>
<td>15%</td>
</tr>
<tr>
<td>South Korea</td>
<td>12.9</td>
<td>2.91E+04</td>
<td>2.43E+08</td>
<td>1.27E+11</td>
<td>51.962%</td>
</tr>
<tr>
<td>Spain</td>
<td>16.3</td>
<td>7.70E+05</td>
<td>8.12E+09</td>
<td>7.42E+10</td>
<td>913%</td>
</tr>
<tr>
<td>Sweden</td>
<td>8.6</td>
<td>2.82E+06</td>
<td>1.56E+10</td>
<td>1.60E+10</td>
<td>103%</td>
</tr>
<tr>
<td>Switzerland</td>
<td>11.1</td>
<td>4.67E+05</td>
<td>3.35E+09</td>
<td>1.30E+10</td>
<td>389%</td>
</tr>
<tr>
<td>Thailand</td>
<td>19.0</td>
<td>4.65E+04</td>
<td>5.71E+08</td>
<td>6.09E+10</td>
<td>10,665%</td>
</tr>
<tr>
<td>Trinidad and Tobago</td>
<td>20.9</td>
<td>1.50E+03</td>
<td>2.03E+07</td>
<td>1.86E+09</td>
<td>9.143%</td>
</tr>
<tr>
<td>Turkey</td>
<td>15.9</td>
<td>6.55E+06</td>
<td>6.74E+10</td>
<td>3.66E+10</td>
<td>54%</td>
</tr>
<tr>
<td>Turkmenistan</td>
<td>16.7</td>
<td>3.55E+07</td>
<td>3.85E+11</td>
<td>5.58E+09</td>
<td>1%</td>
</tr>
<tr>
<td>Ukraine</td>
<td>11.9</td>
<td>1.37E+06</td>
<td>1.06E+10</td>
<td>1.53E+10</td>
<td>145%</td>
</tr>
<tr>
<td>United Arab Emirates</td>
<td>21.5</td>
<td>6.11E+06</td>
<td>8.51E+10</td>
<td>3.79E+10</td>
<td>45%</td>
</tr>
<tr>
<td>United States</td>
<td>12.7</td>
<td>7.03E+07</td>
<td>5.80E+11</td>
<td>9.53E+11</td>
<td>164%</td>
</tr>
<tr>
<td>Uzbekistan</td>
<td>15.6</td>
<td>3.05E+07</td>
<td>3.09E+11</td>
<td>4.53E+09</td>
<td>1%</td>
</tr>
<tr>
<td>Venezuela</td>
<td>21.0</td>
<td>2.51E+06</td>
<td>3.42E+10</td>
<td>4.26E+10</td>
<td>125%</td>
</tr>
</tbody>
</table>