January 1971

The Effect of Carbon on Algal Growth--Its Relationship to Eutrophication

Joel C. Goldman
Donald B. Porcella
Joe E. Middlebrooks
Daniel F. Toerien

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The Effect of Carbon on Algal Growth—
Its Relationship to Eutrophication

By: Daniel C. Goldman, Donald R. Porcella, E. Joe Middlebrooks, and David F. Toorien
Utah Water Research Laboratory, College of Engineering
April 1977
ERRATA

Page 10 Line 16 Reads Should Read
16 2(\text{PO}_4^{3-}) 2(\text{PO}_4^{2-})

Page 14 Figure Title Trussel (1970) Trussell (1970)

Page 39 Figure Title Coree (1970) Foree (1970)

Page 55 Line 23 antotrophic autotrophic

Table 7. Alkalinity and pH data for various natural water bodies in North America.

<table>
<thead>
<tr>
<th>Water Body</th>
<th>Total Alkalinity Actual mg/l as CaCO_3 pH</th>
<th>Equilibrium pH</th>
<th>Free CO_2 Eq CO_2</th>
<th>Total Inorganic Carbon mg/l</th>
<th>Reported Condition of Water Body</th>
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</thead>
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<tr>
<td><strong>Great Lakes</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Eutrophic</td>
</tr>
<tr>
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<td>8.60</td>
<td>1.64</td>
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<td>8.60</td>
<td>1.63</td>
<td>25.0</td>
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<td>8.0</td>
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<td>2.39</td>
<td>27.1</td>
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<td>8.51</td>
<td>1.39</td>
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<td>2.65</td>
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<td>8.27</td>
<td>6.02</td>
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<td>8.2-7.7\textsuperscript{6}</td>
<td>8.69</td>
<td>1.63-5.18 &amp; 29.3-30.5</td>
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<td>2.59-6.49 &amp; 24.3-25.2</td>
<td>Oligotrophic-Mesotrophic</td>
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<td>Big Peter Pond</td>
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<td>8.2-7.2\textsuperscript{6}</td>
<td>8.65</td>
<td>1.51-14.2 &amp; 27.1-31.6</td>
<td>Eutrophic</td>
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</tbody>
</table>

1 Data from reference cited
2 Calculated from Equation 10
3 Calculated from Equation 12
4 Calculated using Saunders et al. (1962) method based on pH and total alkalinity
5 Based upon Rawson's (1960) assumption that 85 percent of the total dissolved solids was bicarbonate
6 Surface pH - Bottom pH
THE EFFECT OF CARBON ON ALGAL GROWTH—ITS RELATIONSHIP TO EUTROPHICATION

A Review Paper

By

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April 1971
Joel C. Goldman is currently a doctoral candidate in Environmental Health Sciences at the University of California, Berkeley. Before going to Berkeley he attended the University of Minnesota where he received both his bachelors and masters degrees in Civil Engineering. Included in Mr. Goldman's engineering experience are employment with the New York City Board of Water Supply and a sanitary engineering consulting firm in Minneapolis. More recently he participated in a joint Federal Government-State of California research program to determine the feasibility of removing nitrate-nitrogen from agricultural return waters of the San Joaquin Valley in central California by growing and harvesting algae. Since then he has become involved in various problems in aquatic biology, particularly the role that inorganic carbon plays in the eutrophication process.

Donald B. Porcella is Assistant Professor of Environmental Biology at Utah State University where he is involved in water quality research at the Utah Water Research Laboratory. Before coming to USU Dr. Porcella was at the University of California at Berkeley as an assistant research zoologist at the Sanitary Engineering Research Laboratory. He was a Fulbright post graduate fellow at the Norwegian Institute for Water Research at Oslo, Norway, and he was also a research zoologist for the U.S. Public Health Service at the R.A. Taft Sanitary Engineering Center in Cincinnati, Ohio. He received the A.B. and M.A. degrees in Zoology and the Ph.D. degree in Environmental Health Science at the University of California at Berkeley. He is the author or co-author of some 25 publications.

E. Joe Middlebrooks is a Professor of Civil Engineering at Utah State University where he is involved in water quality research at the Utah Water Research Laboratory. He came to USU from the University of California at Berkeley where he was an Associate Research Engineer and Assistant Director of the Sanitary Engineering Research Laboratory. From 1962 to 1968 he was Assistant and Associate Professor at Mississippi State University. Dr. Middlebrooks received the BCE and the MSE degrees from the University of Florida, and the Ph.D. degree from Mississippi State University. He is a registered professional engineer, and the author or co-author of some 50 publications.

Danie F. Toerien is currently a post doctoral scholar in the Sanitary Engineering Division, Department of Civil Engineering at the University of California, Berkeley. Before going to Berkeley he attended the University of Pretoria (South Africa) where he received both his masters and doctorate degrees in Microbiology. After employment in the South African Department of Agriculture and at the University of Pretoria, Dr. Toerien, in 1966, joined the National Institute for Water Research of the South African Council for Scientific and Industrial Research. There he was a member of a research team investigating the biochemistry and microbiology of anaerobic digestion. At the University of California, Berkeley, he is participating in a project aimed at developing algal assays, sponsored by the Environmental Protection Agency.
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INTRODUCTION


Effective control of cultural eutrophication must involve the manipulation of those factors which affect algal growth, i.e. light, temperature, nutrients, mixing, predation, etc. (Toerien et al., 1970). Because at the present time man can only effectively control the discharge of nutrient concentrations into aquatic systems, most past and current research on the remediation of eutrophication effects has been concerned with nutrient control. The cost of removal (or exclusion) of specific nutrients varies, so from an economic standpoint it becomes important to identify which nutrients limit algal growth for a given situation.

Algae require carbon, hydrogen, oxygen, nitrogen, phosphorus, sulfur, magnesium, iron, potassium, various other cations, and a number of trace elements to carry out the metabolic processes necessary for growth. Only a few of these elements can be removed from water through treatment at this time. Although Goldman (1960) and Skulberg (1967) have implicated magnesium, iron and molybdenum, and other trace elements as limiting factors for algal growth in some waters, most attention has been directed towards nitrogen and phosphorus as limiting nutrients. Several reasons for this interest in nitrogen and phosphorus follow:

(1) Nitrogen and phosphorus are relatively major constituents in algae; a typical stoichiometric formula for algal biomass being \( \text{C}_{106}\text{H}_{181}\text{O}_{45}\text{N}_{16}\text{P} \) (McCarty, 1970);

(2) Geochemical considerations suggest that phosphorus is probably the most frequently limiting nutrient (Hutchinson, 1957);

(3) The considerably detailed information on the behavior of nitrogen and phosphorus in nature (Hutchinson, 1957, Task Group 2610P (AWWA), 1967);

(4) The relative ease and familiarity of chemical analysis (e.g. Amer. Publ. Health Assoc., 1965) at the relatively high concentrations observed in waste waters (McGauhey, 1968):
(5) the vast amount of research performed on the physiological and biochemical utilization of nitrogen and phosphorus (e.g. Fogg, 1959, Lewin, 1962, Kuhl, 1968);

(6) at least for phosphorus, the relative ease with which it can be removed from waste waters by chemical treatment (Wuhrmann, 1957, Rohlich, 1961).

Because phosphorus can be removed relatively easily in both economical and technical terms (e.g. Culp and Moyer, 1969) and because it has been considered to be the most probable limiting nutrient in most natural waters, proposals for constructing tertiary treatment plants for removing phosphorus from waste waters, elimination of phosphate builders in detergents, and limitations on the use of phosphorus fertilizers have all been advanced as aids in controlling eutrophication.

Recently the concept that phosphorus is the most probable limiting nutrient in natural waters has been questioned (Legge and Dingledein, 1970) and several investigators have suggested that carbon is really the most important limiting nutrient in natural waters (Lange, 1967, Kuentzel, 1969, Kerr et al., 1970, King, 1971). These considerations have now entered the realm of controversy (Kuentzel, 1969, 1970, Legge and Dingledein, 1970, Sawyer, 1970, Shapiro, 1970, Abelson, 1970, Bowen, 1970, Likens, 1971). The implication of carbon as being a major factor controlling cultural eutrophication has significant consequences. Currently removal of phosphorus from detergents has been advocated (Environmental Science and Technology, 1970, Dawson, 1970). The economic aspects of phosphorus removal and replacement with other materials for the consumer and manufacturer and the environmental effects of possible replacements (e.g. substitutes similar to the recently banned nitrito-triacetic acid (NTA)) on aquatic ecosystems have not been evaluated. Therefore, serious questioning of the role of phosphorus in the eutrophication of natural waters may be warranted. However, the possible role of other factors which can limit algal populations should be considered in light of the extensive literature which exists on the subject of algal growth.

It is the goal of this review to clarify and obtain a perspective on the role of carbon in controlling algal growth in natural waters. Information on carbon cycling and metabolism, inorganic carbon chemistry, algal utilization of carbon, and concepts of nutrient limitation will be discussed and reviewed to gain this perspective. This information will provide for a more complete understanding of the role of carbon in eutrophication.
CARBON TRANSFORMATIONS IN THE AQUATIC ENVIRONMENT

The Carbon Cycle

To define the role of carbon in the growth of algae and hence as a factor affecting eutrophication, it is first necessary to consider the carbon cycle in aquatic environments (Figure 1). In the aquatic habitat carbon changes revolve around synthesis and degradation of organic matter, i.e. reduction of inorganic carbon into organic carbon and then oxidation of organic carbon into inorganic carbon, with a strong interdependence of these reactions upon each other. In the conversion of inorganic carbon into organic carbon the photosynthetic reactions of plants, algae and bacteria, are important, while microbial degradation of organic carbon is important in the reverse reaction.

Synthesis of Organic Carbon Compounds

Organic carbon compounds may be synthesized from CO$_2$ in the aquatic environment through three types of reactions, (1) photosynthesis by plants and algae, (2) photosynthesis by certain bacteria, and (3) incorporation of carbon by other mechanisms.

Plant and algal photosynthesis

In its simplest form the photosynthetic reaction can be expressed as:

$$2H_2O + CO_2 \xrightarrow{\text{LIGHT}} (CH_2O) + H_2O + O_2$$  \hspace{1cm} (1)

While much is known about the intricacies of the specific biochemical reactions involved, Equation (1) only indicates the initial reactants required and the resulting products formed. The process of photosynthesis is considered to occur in two separate stages (Rabinowitch and Govindjee, 1969) designated in Figure 2. First, through the action of light (radiant energy) on the chlorophyll pigment, a high energy oxidant and a high energy reductant are formed. Then the reductant is used in the conversion of CO to carbohydrates (formation of potential chemical energy) while the oxidant gives rise to molecular oxygen.

Bacterial photosynthesis

In bacterial photosynthesis oxygen is not generated nor is water photometabolized, but other reduced inorganic or organic compounds are utilized
Figure 1. Carbon cycle in aquatic ecosystems.

(Pfennig, 1967). The Thiorhodaceae (purple sulfur bacteria) and the Chlorobacteriaceae (green sulfur bacteria) photometabolize reduced sulfur compounds, while the Athiorhodaceae (non-sulfur purple bacteria) photometabolize simple organic compounds (Pfennig, 1967). The simplest reaction describing bacterial photosynthesis is:

\[
\text{CO}_2 + 2\text{H}_2\text{A} \xrightarrow{\text{LIGHT}} \text{BACTERIOCHLOROPHYLL} \rightarrow (\text{CH}_2\text{O}) + 2\text{A} + \text{H}_2\text{O} . \quad (2)
\]

where \(\text{H}_2\text{A}\) is either a reduced sulfur compound or a simple organic compound.

The photosynthetic bacteria occur in the anaerobic zones of all aquatic environments (Pfennig, 1967). Although always present, the non-sulfur purple bacteria are usually not visible. Purple and green sulfur bacteria are often abundant, giving rise to pink and green layers or blooms (Kaiser, 1966a, 1966b), especially below the thermocline in lakes (Bavendamm, 1924, Genovese, 1963).

Because the photosynthetic bacteria photometabolize all end products of fermentation reactions (Pfennig, 1967), they are important in the carbon cycle in aquatic habitats. In relation to the growth of green and blue-green algae and to their effect in producing unsightly masses, the photosynthetic bacteria are probably not
Figure 2. Mechanisms of plant and bacterial photosynthesis.

too important in natural waters. They can be the dominant biotic form in oxidation ponds which receive high organic loading and thus have a prevailing anaerobic environment (Holm and Vennes, 1970).

Non-photosynthetic incorporation of CO₂

Non-photosynthetic incorporation of CO₂ takes place in two broad categories, namely that by chemoautotrophic bacteria and that by heterotrophic organisms. The chemoautotrophic bacteria include a number of specialized groups which are able to use inorganic oxidizable substrates as energy sources for growth (Stanier et al., 1970). Included under the chemoautotrophic bacteria are the hydrogen bacteria, the colorless sulfur bacteria, the nitrifying bacteria, the iron bacteria, and some of the methane bacteria. Most of these bacteria are obligate autotrophs, capable of using CO₂ as sole source of carbon. Because the chemoautotrophic bacteria rarely constitute a significant portion of the biomass in natural waters, incorporation of CO₂ into organic material through activities of these bacteria possibly never is important in surface waters.

Some non-algal heterotrophic organisms incorporate CO₂ into organic matter during their growth. For instance, the continued oxidation of acetyl coenzyme A by means of the citric acid cycle in many heterotrophs requires the simultaneous presence of oxaloacetate. If the constituents of the citric acid cycle are used in cellular synthesis, as indeed they are, a drain is put on the dicarboxylic acids in the cycle. To
replenish this drain, anaplerotic enzymatic pathways are used (Kornberg, 1966). These anaplerotic sequences can make use of carboxylation reactions (incorporation of carbon dioxide). In such a way interconversions of pyruvate and dicarboxylic acids of the citric cycle are brought about (Mahler and Cordes, 1966). Enzymes participating in such reactions are malic enzyme (which enjoys a wide distribution in nature), pyruvate carboxylase, phosphoenolpyruvate carboxykinase, and phosphoenol pyruvic carboxylase (Mahler and Cordes, 1966). However, the incorporation of CO₂ by heterotrophic organisms (other than algae) in most natural surface waters probably constitutes an insignificant portion of the total incorporation of CO₂ in these waters.

Degradation of Organic Matter

Every biologically important element can be considered to pass through a continuous cycle from the non-living environment to the living environment and back to the non-living environment (Stanier et al., 1970). This cycle of matter occurs in the aquatic environment as well, giving rise to the phenomenon termed the self-purification of water in which moderate amounts of organic pollution are transformed into stable end products (Pelczar and Reid, 1965). These transformations are brought about by the activities of many types of microorganisms present in the aquatic habitat.

The degradation of organic matter in the aquatic habitat appears to be basically similar to that in any other habitat in which organic matter is degraded, e.g. the soil, the intestinal tracts of various animals, sewage treatment plants, etc. Under anaerobic conditions different end products are formed than under aerobic conditions, but many of these compounds formed under anaerobic conditions are further oxidized under aerobic conditions. In general, CO₂ is the end product for all organic carbon compounds. For the purposes of this review a generalized scheme of organic matter degradation is presented in Table 1. The product compounds viz. CO₂, NH₃, NO₃, and PO₄ can again be used for growth of photosynthetic organisms.
Table 1. Generalized scheme of microbial degradation of organic waters.

<table>
<thead>
<tr>
<th>Substrate + Microbial Activity</th>
<th>Yields</th>
<th>Representative End Products&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Anaerobic</th>
<th>Aerobic</th>
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<tr>
<td>Proteins and other nitrogenous compounds</td>
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<tr>
<td></td>
<td>Amino acids +</td>
<td>Amino acids</td>
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<tr>
<td></td>
<td>Ammonia +</td>
<td>Nitrites + Nitrates</td>
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<td></td>
<td>Hydrogen sulfide +</td>
<td>Sulfates</td>
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<td>H&lt;sub&gt;2&lt;/sub&gt;</td>
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<td></td>
<td>Alcohols +</td>
<td>CO&lt;sub&gt;2&lt;/sub&gt; + H&lt;sub&gt;2&lt;/sub&gt;O</td>
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<td></td>
<td></td>
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<td>Neutral compounds +</td>
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<td></td>
<td></td>
<td>Glycerol +</td>
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</tr>
<tr>
<td></td>
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<td></td>
<td></td>
<td>Alcohols +</td>
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</tr>
<tr>
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<td></td>
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<td></td>
</tr>
<tr>
<td>purines, pyrimidines</td>
<td></td>
<td></td>
<td>Amino acids</td>
<td>Amino acids</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Lower fatty acids +</td>
<td>CO&lt;sub&gt;2&lt;/sub&gt; + H&lt;sub&gt;2&lt;/sub&gt;O</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>NH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>NO&lt;sub&gt;2&lt;/sub&gt; + NO&lt;sub&gt;3&lt;/sub&gt;</td>
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<td></td>
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<td>CO&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
</tbody>
</table>

<sup>a</sup> Indicates that compound can be further oxidized under aerobic conditions by other microbes.
THE CHEMISTRY OF INORGANIC CARBON IN NATURAL WATERS

The CO$_2$-HCO$_3^-$-CO$_3^{2-}$ equilibrium system, a major component of the buffering system of most natural waters (Weber and Stumm, 1963a, 1963b), is affected to a large degree through changes in pH resulting from algal growth. The extraction of CO$_2$ from an algal growth system through assimilation into algal biomass at a rate faster than it can be replaced through atmospheric CO$_2$ diffusion, respiration, fermentation processes, and readjustment of solid carbonate equilibria leads to an increase in pH level. This rise in pH can affect algal growth in a number of ways. These include:

a. A change in the carbon species: Aqueous CO$_2$ is reduced in concentration and the equilibrium system is shifted so that the HCO$_3^-$ and CO$_3^{2-}$ ions predominate. Research to date indicates that certain algae growing autotrophically can use only aqueous CO$_2$ as their carbon source, while others, notably blue-green algae, dominate in higher pH environments and may utilize the HCO$_3^-$ and even CO$_3^{2-}$ ion directly (Jackson, 1964). A considerable controversy exists today over whether or not these ions are used directly (Holm-Hansen, 1967).

b. Solubility: The solubility of other essential nutrients, particularly phosphorus, iron and trace elements is affected. Algal growth could then become limited by one or more of these precipitated and unavailable elements.

c. Metabolic effects: Extreme pH values affect the metabolic mechanisms of all living organisms and algal growth rates could be altered at the higher pH values found in some active algal systems.

CO$_2$-HCO$_3^-$-CO$_3^{2-}$ Equilibrium System

The pH of the most natural waters is greatly influenced by the CO$_2$-HCO$_3^-$-CO$_3^{2-}$ equilibrium system. The system is governed by both the content of atmospheric CO$_2$ and by the total alkalinity of the water. The entire equilibrium system can be described by the following stoichiometric equations (all constants used to describe the CO$_2$-HCO$_3^-$-CO$_3^{2-}$ system are from Kern (1960)):

Ionization of water

\[
H_2O \rightleftharpoons H^+ + OH^-; \quad K_w = (H^+) \cdot (OH^-) = 10^{-14} \text{ @} 25^\circ \text{C} \quad \ldots \quad (3)
\]

Solubility of CO$_2$ in H$_2$O

\[
CO_2 (\text{gas}) \rightleftharpoons CO_2 (\text{aq}); \quad CO_2 = k_{CO_2} P_{CO_2} \quad \ldots \quad (4)
\]
\[ k_{\text{CO}_2} = 10^{-1.5} \text{ moles/atmosphere @} 25^\circ \text{C} \]
\[ P_{\text{CO}_2} = 10^{-3.5} \text{ for atmospheric air} \]

Hydration of CO\(_2\)

\[ \text{CO}_2(\text{aq}) + \text{H}_2\text{O} \rightleftharpoons \text{H}_2\text{CO}_3; \quad K_q = \frac{(\text{CO}_2)}{(\text{H}_2\text{CO}_3)} = 600 \quad \ldots \ldots \ldots \ldots (5) \]

Since \((\text{CO}_2)\) (aq) at equilibrium is much greater than \((\text{H}_2\text{CO}_3), (\text{H}_2\text{CO}_3)^* = (\text{H}_2\text{CO}_3) + (\text{CO}_2)\) (aq) can be considered valid and \((\text{H}_2\text{CO}_3)^*\) will hereafter be considered to be equal to the sum of the dehydrated and hydrated forms of CO\(_2\).

First ionization of \((\text{H}_2\text{CO}_3)^*\)

\[ \text{H}_2\text{CO}_3^* \rightleftharpoons \text{H}^+ + \text{HCO}_3^-; \quad K_1 = \frac{(\text{H}^+)(\text{HCO}_3^-)}{(\text{H}_2\text{CO}_3)^*} = 4.45 \times 10^{-7} \text{ @} 25^\circ \text{C} (6) \]

\((K_1' = \text{apparent dissociation constant})\)

Second ionization of \((\text{H}_2\text{CO}_3)\)

\[ \text{HCO}_3^- \rightleftharpoons \text{H}^+ + \text{CO}_3^{2-}; \quad K_2 = \frac{(\text{H}^+)(\text{CO}_3^{2-})}{(\text{HCO}_3^-)} = 4.6 \times 10^{-11} \text{ @} 25^\circ \text{C} (7) \]

Total alkalinity of most natural waters (in equiv./liter)

\[ \text{Tot. Alk. (ALK)} = (\text{HCO}_3^-) + 2(\text{CO}_3^{2-}) + (\text{OH}^-) + (\text{NH}_3) + (\text{H}_2\text{PO}_4^-) + 2(\text{PO}_4^{3-}) + 3(\text{PO}_4^{3-}) + (\text{B(OH}_4^-) - (\text{H}^+) \ldots (8) \]

\((\text{H}^+)\) can be neglected for \((\text{H}^+) < 10^{-4} \quad (\text{pH} > 4)\)

Temperature and ionic strength greatly influence these equations. Morton and Lee (1968a) and Park et al. (1970) have shown the importance of considering activity rather than concentration when describing the CO\(_2\)-HCO\(_3^-\)-CO\(_3^{2-}\) system. Morton and Lee (1968a) in their analysis of Lake Mendota in Wisconsin calculated an activity coefficient of 0.74 for the CO\(_3^{2-}\) ion, based on the water's ionic strength of 0.0045. Similarly, Park et al. (1970) found the ionic strength of the Columbia River to be 0.0018 giving an activity coefficient for the CO\(_3^{2-}\) ion of 0.83. Although most natural waters are relatively dilute solutions, these results demonstrate that activity rather than concentration must be considered in any generalized formulation of the CO\(_2\)-HCO\(_3^-\)-CO\(_3^{2-}\) system, particularly for the divalent ions such as CO\(_3^{2-}\) which are
greatly influenced by ionic strength. The importance of this concept can be seen in Figure 3 where activity coefficients for the various components of the CO$_2$-HCO$_3$-CO$_3^-$ system, as a function of ionic strength, are plotted according to the data of Klotz (1964). These curves show the strong dependence of the activity of the CO$_3^-$ ion on ionic strength. The mono-valent ions have less of a dependency. The need to consider activity rather than concentration in highly mineralized waters has been considered by Berner (1965), who has reviewed the role of activity coefficients in the CO$_2$-HCO$_3$-CO$_3^-$ system of sea water.

Temperature as well as ionic strength greatly influences this system. Both Langelier (1946) and Dye (1952) have discussed its importance in great detail. To demonstrate this effect the thermodynamic equilibrium constants, K$_1$, K$_2$, and K$_w$ are plotted in Figure 4 as functions of temperature according to the data of Harned and Scholes (1941), Harned and Bonner (1945), and Harned and Owen (1958). The constants increase with increasing temperature. Also, plotted in this figure are the solubility coefficients for CO$_2$ in water, k$_{CO2}$ in moles per atmosphere, as a function of temperature according to Harned and Davis (1943). In contrast to the equilibrium constants this coefficient decreases with increasing temperature.

![Figure 3. Effect of ionic strength on activity coefficients for components of CO$_2$-HCO$_3$-CO$_3^-$ system.](image)
By combining and rearranging Equations (3) through (8) the equilibrium pH of a natural water can be described. The following equations, as proposed by Weber and Stumm (1963a) and by Thomas and Trussell (1970) show this relationship:

$$
\begin{align*}
(H^+) &= \frac{K_w + K_1' (H_2CO_3)^*}{2(ALK)} \\
&= \sqrt{\frac{[K_w + K_1' (H_2CO_3)^*]^2 + 8(ALK) K_1' K_2 (H_2CO_3)^*}{2(ALK)}} \\
&\quad \ldots (9)
\end{align*}
$$

Figure 4. Effect of temperature on equilibrium constants and solubility coefficient in CO$_2$-HCO$_3$-CO$_3^-$ system.
or

\[
\text{pH} = -\log_{10} \left[ \frac{K_w + K_1' \left( k_{CO_2} \right)^2 \left( P_{CO_2} \right)}{2(ALK)} \right] + \sqrt{\left[ \frac{K_w + K_1' k_{CO_2} \left( P_{CO_2} \right)}{2(ALK)} \right]^2 + 8(ALK) K_1' K_2 k_{CO_2} \left( P_{CO_2} \right)} \] ...

(10)

These equations hold true only for those situations where the free CO\(_2\) concentration in solution is in equilibrium with atmospheric CO\(_2\). For most natural waters this situation is rarely ever found. Stumm (1964) indicates that most natural waters are supersaturated with CO\(_2\). Morton and Lee (1968a) have made a similar claim, showing that thermal stratification plays an important role in maintaining vertical CO\(_2\) gradients in natural waters. Higher CO\(_2\) concentrations, and corresponding lower pH values are found nearer the bottom where bacterial activity predominates. Hutchinson (1957) in his classical work has extensively described the CO\(_2\)-HCO\(_3^-\)-CO\(_3^-\) equilibrium system in lakes and has made a similar claim that natural water bodies are often supersaturated with CO\(_2\). He has greatly stressed the point that continually occurring complex interactions such as biological activity and heterogeneous chemical reactions, greatly affect the CO\(_2\) concentration present in a water body.

Horne (1969) has reviewed the latest work on CO\(_2\) equilibrium in the oceans and reports that while the Pacific Ocean is undersaturated to a large degree, the Indian Ocean and the South Atlantic near the equator are supersaturated. Keeling (1968) has developed a map showing the distribution of CO\(_2\) in the oceans at the surface. His conclusions are that while supersaturation occurs at the equator, undersaturation occurs toward the poles. Also, Kelley and Hood (1969) have reported that CO\(_2\) concentrations in the North Pacific Ocean and the Bering Sea are greatly affected by the currents and the river discharges. The supersaturation with dissolved CO\(_2\) at the entrance to Puget Sound in Washington was equivalent to an atmospheric CO\(_2\) concentration of 0.09 percent. Park et al. (1969) also showed that the dissolved CO\(_2\) concentration of the Columbia River was greatly in excess of equilibrium values. Other rivers in the region displayed a similar characteristic (Park et al., 1970).

Referring back to Equation (10), the equilibrium pH of a natural water increases with an increase in total alkalinity. This relationship is shown in Figure 5 in which curves are plotted for varying temperatures (Thomas and Trussell, 1970).

The molar concentration of all three carbon species making up the equilibrium system can be described in terms of the pH and alkalinity of a given system by the following equations:
\[ C_T = (H_2CO_3)^* + (HCO_3^-) + (CO_3^{2-}) \] (11)

in which \( C_T \) equals the total molar concentration of carbon.

Figure 5. Effect of alkalinity on equilibrium pH for aquatic system exposed to atmosphere (\( CO_2 = 0.03 \) percent). From Thomas and Trussel (1970).
\[(\text{H}_2\text{CO}_3)^* = \frac{(\text{H}^+)}{K_1} \left[ \frac{(\text{ALK}) - (\text{OH}^-) + (\text{H}^+)}{1 + \frac{2K_2}{(\text{H}^+)}} \right] \] .................................. (12)

\[(\text{HCO}_3^-) = \frac{(\text{ALK}) - (\text{OH}^-) + (\text{H}^+)}{1 + \frac{2K_2}{(\text{H}^+)}} \] .................................. (13)

\[(\text{CO}_3^--) = \frac{K_2}{(\text{H}^+)} \left[ \frac{(\text{ALK}) - (\text{OH}^-) + (\text{H}^+)}{1 + \frac{2K_2}{(\text{H}^+)}} \right] \] .................................. (14)

A distribution diagram based upon Equations (11) through (14) showing the relative proportions of all three species as a function of pH (independent of alkalinity concentration) is presented in Figure 6 in which the relative proportions of the three carbon species change as a function of pH. However, when the system is in equilibrium with atmospheric \(\text{CO}_2\) the concentration of free \(\text{CO}_2\) in solution remains constant for all pH values while the concentration of the bicarbonate and carbonate species changes as a function of the total carbon alkalinity.

Figure 6. Effect of pH on distribution of inorganic carbon species in \(\text{CO}_2\)-\(\text{HCO}_3^-\)-\(\text{CO}_3^--\) system.
Alkalinity and Buffering Capacity

The operational definition of alkalinity for most natural waters, as defined by Equation (8), implies that the $\text{CO}_2^{-}$-$\text{HCO}_3^{-}$-$\text{CO}_3^{2-}$ equilibrium system is the major buffering system of fresh waters. As contrasted to sea water, where the silicate and borate concentrations are high and are major contributors to the alkalinity and buffering capacity, particularly the silicates (Sillen, 1961; Garrels, 1965), fresh waters are normally quite low in these constituents. Also, phosphates and ammonia (at high pH), which can also contribute to a water's total alkalinity, are found only in relatively minute concentrations in fresh waters. They usually make up such a minor component of the total alkalinity that they are neglected in making such a determination.

Thus, the apparent major buffering system of a fresh water is composed of the $\text{CO}_2^{-}$-$\text{HCO}_3^{-}$-$\text{CO}_3^{2-}$ system. However, as pointed out by Weber and Stumm (1963a), the homogeneous $\text{CO}_2^{-}$-$\text{HCO}_3^{-}$-$\text{CO}_3^{2-}$ buffering system, although an important component of the overall buffering capacity of a natural water, is itself greatly affected by the complicated network of heterogeneous and biological reactions continually occurring in the aquatic environment as the whole system strives toward, but seldom reaches, equilibrium. Stumm (1964, 1967), Bricker and Garrels (1967), and Bostrom (1967) have suggested that heterogeneous reactions involving solid phases, such as clays and other minerals, may be the principal buffering agents in fresh waters. They point out that there is a dearth of information in this area and stress the need for further research.

Biological Effects on pH

Biological activity can alter the pH of a natural water in many ways. Table 2, taken from Weber and Stumm (1963a), has depicted the various biological reactions that may alter pH. As an example, Berner et al. (1970), has recently demonstrated the effect of sulfate reduction in marine sediments on increasing alkalinity and hence buffering capacity. Since many of these reactions are localized in the sense that they occur only in certain portions of an aquatic ecosystem (i.e. photosynthesis in the photic zone, reduction processes in anaerobic portions of the bottom sediments), vertical gradients of decreasing pH with increasing depth are very often found in natural waters. These gradients are maintained through thermal stratification. With the onset of spring and fall overturn, relatively equal pH levels are produced for short times throughout the volume of the water body. The formation of new thermal gradients together with the continuous biological activity once more causes gradients to form. (See review by Lee and Hoadley, 1967.)

The effect of actively growing algae on the pH of surface waters can be better understood by an analysis of the complete photosynthetic reaction. This reaction should show the assimilation of all nutrients and the formation of complete products. Equation (1) should thus be expanded from its simplified form.

As an example, the assimilation of nitrogen affects the pH of a water in either direction depending on the form of nitrogen being assimilated. If $\text{NH}_4^+$-$\text{N}$ is used the pH will decrease, while the utilization of $\text{NO}_3^{-}$-$\text{N}$ will cause a rise in pH. This phenomenon has been observed by a number of researchers (Trelease and Trelease,
Table 2. Biologically mediated reactions affecting pH in natural water systems.*

<table>
<thead>
<tr>
<th>Process</th>
<th>Reaction</th>
<th>Effect on pH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Photosynthesis</td>
<td>$6(\text{CO}_2) + 6(\text{H}_2\text{O}) \rightarrow (\text{C}_6\text{H}_12\text{O}_6 + 6\text{O}_2)$</td>
<td>Increase</td>
</tr>
<tr>
<td>Respiration</td>
<td>$(\text{C}_6\text{H}_12\text{O}_6 + 6\text{O}_2) \rightarrow 6(\text{CO}_2) + 6(\text{H}_2\text{O})$</td>
<td>Decrease</td>
</tr>
<tr>
<td>Methane Fermentation</td>
<td>$(\text{C}_6\text{H}_12\text{O}_6) + 3(\text{CO}_2) \rightarrow 3(\text{CH}_4) + 6(\text{CO}_2)$</td>
<td>Decrease</td>
</tr>
<tr>
<td>Nitrification</td>
<td>$(\text{NH}_4^+) + 2(\text{O}_2) \rightarrow (\text{NO}_3^-) + \text{H}_2\text{O} + 2(\text{H}^+)$</td>
<td>Decrease</td>
</tr>
<tr>
<td>Denitrification</td>
<td>$5(\text{C}_6\text{H}_12\text{O}_6) + 24(\text{NO}_3^-) + 24(\text{H}^+) \rightarrow 30(\text{CO}_2) + 12(\text{N}_2) + 42(\text{H}_2\text{O})$</td>
<td>Increase</td>
</tr>
<tr>
<td>Sulfide Oxidation</td>
<td>$(\text{HS}^-) + 2(\text{O}_2) \rightarrow (\text{SO}_4^{2-}) + (\text{H}^+)$</td>
<td>Decrease</td>
</tr>
<tr>
<td>Sulfate Reduction</td>
<td>$(\text{C}_6\text{H}_12\text{O}_6) + 3(\text{SO}_4^{2-}) + 3(\text{H}^+) \rightarrow 6(\text{CO}_2) + 3(\text{HS}^-) + 6(\text{H}_2\text{O})$</td>
<td>Increase</td>
</tr>
</tbody>
</table>

* From Weber and Stumm (1963a)
1935. Cramer and Myers. 1948. Davis et al. 1953. Cramer and Myers (1948), working with Chlorella, proposed the following stoichiometric equations to describe this occurrence:

Nitrate Assimilation

\[
\begin{align*}
1.0 \left(\text{NO}_3^-\right) + 5.7(\text{CO}_2) + 5.4(\text{H}_2\text{O}) \xrightarrow{\text{LIGHT}} (C_{5.7H9.8O2.3N1.0}) \\
+ 8.25(\text{O}_2) + 1.0(\text{OH}^-) 
\end{align*}
\]  

(15)

Ammonia Assimilation

\[
\begin{align*}
1.0(\text{NH}_4^+) + 7.6(\text{O}_2) + 17.7(\text{H}_2\text{O}) \xrightarrow{\text{LIGHT}} (C_{7.6H8.1O2.5N1.0}) \\
+ 7.6(\text{O}_2) + 15.2(\text{H}_2\text{O}) + 1.0(\text{H}^+) 
\end{align*}
\]  

(16)

These equations explain the opposite effects on pH caused by either \(\text{NO}_3^-\) or \(\text{NH}_4^+\) assimilation. Every mole of \(\text{NO}_3^-\) assimilated results in the formation of one mole of \(\text{OH}^-\), thus raising the pH. \(\text{NH}_4^+\) assimilation, on the other hand, leads to the production of one mole of \(\text{H}^+\) for every mole of \(\text{NH}_4^+\) assimilated, with a resulting decrease in pH.

As previously stated, algae can utilize \(\text{CO}_2\) from four major sources in a natural water: (1) From \(\text{CO}_2\) diffused from the atmosphere; (2) from the respiration of heterotrophic forms; (3) from anaerobic fermentation; and (4) from bicarbonate alkalinity. The first three sources provide a direct supply of \(\text{CO}_2\). The alkalinity, on the other hand, provides \(\text{CO}_2\) by a continual readjustment of the concentrations of the various carbon sources making up the \(\text{CO}_2\)-HCO\(_3^-\)-CO\(_3^-=\) system as shown by the following equations:

\[
\begin{align*}
2(\text{HCO}_3^-) &\leftrightarrow (\text{CO}_3^-) + (\text{H}_2\text{O}) + (\text{CO}_2) \\
(\text{HCO}_3^-) + (\text{H}_2\text{O}) &\leftrightarrow (\text{H}_2\text{O}) + (\text{CO}_2) + (\text{OH}^-) \\
(\text{CO}_3^-) + (\text{H}_2\text{O}) &\leftrightarrow (\text{CO}_2) + 2(\text{OH}^-)
\end{align*}
\]  

(17) (18) (19)

Typically, the pH of natural waters is about 8.3 where \(\text{HCO}_3^-\) is the major ion (see Figure 6). Thus, as \(\text{CO}_2\) is extracted from solution by growing algae at a pH around 8.3, additional carbon dioxide is provided through these reactions. Both Equations (17) and (18) describe the principal reactions, with the reaction in Equation (17) being the dominant of the two. As the pH rises the \(\text{CO}_3^-\) form becomes the major carbon species and it, too, can be converted directly to \(\text{CO}_2\) by a hydration process as shown in Equation (19). This reaction similarly results in a pH rise. It is not uncommon to have pH values as high as 10 to 11 in active algal systems such as waste stabilization ponds. This phenomenon has been observed by a number of researchers (Golueke et al., 1962, Pipes, 1962b, Beck et al., 1969). This rise in pH
gives evidence that the CO₂ supplied from the first three sources mentioned is either unavailable (i.e. diffusion gradients) or insufficient to meet the demands of the growing algae and that a further demand is placed on the bicarbonate alkalinity through a readjustment of the CO₂-HCO₃⁻-CO₃²⁻ system. It is obvious then that even when the free CO₂ content of a water is insufficient to meet the demand of the algae, the HCO₃⁻ and CO₃²⁻ forms can continually supply free CO₂ for algal utilization. Deuser (1970) showed that the carbon utilized by the diatom Chaetoceros curvisetum during a heavy bloom condition in the Black Sea was derived from inorganic carbon species other than free CO₂ after the initial free CO₂ was depleted.

Sawyer and McCarty (1967) point out that no changes in the total alkalinity of the system occur when free CO₂ is utilized by algae since there is no change from electrical neutrality. However, when and if the HCO₃⁻ ion is used directly there would be a decrease in alkalinity proportional to its uptake. Similarly, as the pH rises due to algal growth CO₃²⁻ and eventually OH⁻ will begin to precipitate out of solution with a corresponding reduction in alkalinity. Also the concentration of other alkalinity components such as phosphates could be affected by algal growth and significant effects on the total alkalinity could occur depending on the relative concentrations of all the alkalinity components of the system.

CaCO₃ Equilibrium

While the literature dealing with the purely chemical aspects of CaCO₃ equilibrium in natural waters is extensive, surprisingly little attention has been given to the role of CaCO₃ and phytoplankton activity. The chemistry of CaCO₃ formation is extremely complex. As pointed out by Bricker and Garrels (1967), CaCO₃ exists in a number of polymorphic and hydrated forms, although the two principal forms found in the sediments of natural waters appear to be calcite and aragonite. Many natural waters are supersaturated with CaCO₃, both in the calcite and aragonite forms (Bricker and Garrels, 1967) and many areas of the oceans are saturated with calcite (Weyl, 1961, Schmalz and Chave, 1963, Dietrich, 1963, Siever et al., 1965). Peterson (1966) has shown that waters in the Pacific are unsaturated with respect to calcite except at the surface. Morton and Lee (1968b) have demonstrated the importance of ion pair formation, particularly with magnesium, on CaCO₃ formation in ocean waters.

In natural fresh waters, CaCO₃ saturation is not as common as it is in ocean waters. Kramer (1967) has studied CaCO₃ formation in the Great Lakes and indicates that saturation is very temperature dependent, and for the observed temperatures Lake Erie and Lake Ontario are mostly unsaturated. Morton and Lee (1968a) have found Lake Mendota in Wisconsin to be unsaturated with respect to CaCO₃ in the bottom layers while supersaturated in the surface waters.

Weber and Stumm (1963a, 1963b) and Kleijn (1965) have shown the effect of CaCO₃ saturation on the buffering capacity of a natural water. A water saturated with CaCO₃ is considerably more buffered than the same water which is unsaturated. It follows that the water in its heterogeneous natural environment is more strongly buffered than the same water studied in the laboratory. Thus, factors other than the homogeneous CO₂-HCO₃⁻-CO₃²⁻ equilibrium system help control the buffering
capacity of these waters. The role of this phenomenon in supplying or denying carbon for the growth of algae is still unclear. For example, the observation that Charophytes become incrusted with CaCO₃ during growth and yet are dominant in waters saturated with CaCO₃ seems to indicate that at least some plants are able to extract sufficient CO₂ from CaCO₃-saturated waters (Forsberg, 1965). Thermodynamically, it would take considerable energy to cause the CO₂ to become available from solid phase CaCO₃ for algal growth without the addition of H⁺. Therefore the utilization of CO₂ would lead to a rise in pH and actually lead to a decrease in available inorganic carbon for growth as in the following reaction which would predominate at a pH greater than 6.5:

\[ \text{Ca}^{++} + 2\text{HCO}_3^- = \text{CO}_2 + \text{CaCO}_3 + \text{H}_2\text{O} \]

(algal uptake) (precipitate)

Hutchinson (1957) applying the fact that the carbonate in natural limestone contains no ¹⁴C has observed that only a relatively few hard water lakes have lower ¹⁴C content than the atmosphere. This indicates that most of the carbon in lake systems is of modern origin and thus enters the aquatic system as allochthonous material or directly from the air. This further suggests that little CO₂ would be available from CaCO₃.

In the time span of a transient algal bloom the solid phase CaCO₃ system may not have any effect on the availability of carbon since the rate at which CaCO₃ goes into and out of solution is slow. Thus, the maintenance of the equilibrium system may not keep pace with the changes in the CO₂-HCO₃⁻-CO₃²⁻ system due to photosynthetic activity. Also, other factors present in a natural water may accelerate or hinder CaCO₃ precipitation. Chave (1965) and Chave and Suess (1967, 1970) have shown that organic compounds coat CaCO₃ precipitates, thus preventing their return to solution under otherwise favorable conditions. Others (Oppenheimer, 1961, Greenfeld, 1963) have shown accelerated CaCO₃ precipitation in the presence of bacteria. Simkiss (1964) has shown the inhibitory effect of certain organophosphates on the precipitation of CaCO₃ in sea water. Eyster (1958) has shown a similar effect on CaCO₃ precipitation by inorganic phosphates. He suggests that the formation of marl deposits is associated with phosphate deficiencies. Stumm and Leckie (1970) have demonstrated the buildup of hydroxyapatite on solid CaCO₃ surfaces, stressing the point that this phenomenon is a major factor controlling the resolubilization of phosphates in natural waters. Paasche (1963, 1964) and Steemann Nielsen (1966) have demonstrated in detail the mechanisms governing CaCO₃ precipitation due to the growth of the marine coccolithophorid, Coccolithus huxleyi.

Because of the great complexity involved in CaCO₃ chemistry, it is difficult to ascertain its role in the total CO₂-HCO₃⁻-CO₃²⁻ system with respect to the availability of inorganic carbon for algal growth. One would surmise that CaCO₃ would have minimal effect, but further research is required before CaCO₃ chemistry can be related to the problems of eutrophication.

20
Reaction Rates

If carbon were limiting in an algal bloom, the slowest reaction involved in the chemical transformation of the ionic forms of inorganic carbon to free CO$_2$ could be rate limiting and could be the controlling factor governing the utilization of inorganic carbon in photosynthesis. This concept is a key point in any discussion of carbon limitations in natural waters; yet there is almost a complete absence of information on this subject in the literature.

Chemical reaction rates

Kern (1960) has reviewed the research on CO$_2$ hydration and dehydration and points out that while the reactions described in Equations (6) and (7) are instantaneous, the hydration and dehydration of CO$_2$ as described in Equation (5) is a relatively slow step. The slowness of the dehydration step was claimed by Hood and Park (1962) to be the factor favoring direct bicarbonate utilization by certain marine phytoplankton. Watt and Paasche (1963) and Steemann Nielsen (1963) have effectively disputed this claim.

Dehydration of carbon dioxide can occur in one of two ways, depending on the pH. For pH values below 8 the reaction described in Equation (5) occurs. At pH values greater than 10, dehydration can occur via the following reaction (Kern, 1960):

\[
\text{HCO}_3^- \rightleftharpoons \text{CO}_2 + \text{OH}^-. \quad (20)
\]

Between pH values of 8 and 10 both methods of dehydration are significant. Also in this pH range the normal dehydration of H$_2$CO$_3$ to CO$_2$ and H$_2$O is catalyzed by the presence of OH$^-$ ions (Kiese and Hastings, 1940).

Considerable research has been performed on determining the rate constants for the dehydration reaction in Equation (5) (Faurholt, 1925, Brinkman et al., 1933, Roughton, 1941, Scheurer et al., 1958, Sirs, 1958). Roughton (1941), using a thermometric method, determined the rate constant for the dehydration process according to Equation (5) for various temperatures. These results are plotted in Figure 7 and show a strong temperature dependency of the rate constant. At a temperature of 25°C the rate constant is 26.6 sec$^{-1}$. Rabinowitch (1951) has reported the rate constant at 25°C for the dehydration reaction in Equation (20) to be 0.47x10$^{-4}$ sec$^{-1}$, considerably slower than in Equation (5), while Kern (1960) reviewed the literature and found it to be 2x10$^{-4}$ sec$^{-1}$. Thus, can a situation occur where the rate of CO$_2$ assimilation by photosynthesizing algae during a bloom condition be so rapid that the rate of dehydration of H$_2$CO$_3$ becomes a rate-limiting step? Rabinowitch (1951) has attempted to answer this question by developing a hypothetical situation where the following conditions prevailed: 1) A pH of over 10; 2) a HCO$_3^-$ concentration of 0.02 M; and 3) that only the reaction in Equation (20) took place. He calculated that a maximum of 9x10$^{-7}$ mole CO$_2$/l sec would be available for algal growth. He then considered that a 0.1 percent (by volume) concentration of algae would, under strong light conditions, be capable of photosynthesizing at a maximum rate of 3.3x10$^{-7}$ mole CO$_2$/l sec. Thus almost three times more CO$_2$ would be available than would be required for this concentration of
algae. It could be argued that the numbers used by Rabinowitch for these calculations are completely unrealistic when applied to a typical bloom condition in an eutrophic natural water. In order to show that Rabinowitch's calculation was actually conservative when applied to a bloom condition in a eutrophic water a similar calculation will be made based on the following assumptions:

1) That the total bicarbonate alkalinity as CaCO$_3$ is equal to 50 mg/l;
2) That the bloom has already progressed to the point where the pH has been raised to 10, where CO$_2$ is derived from HCO$_3^-$ directly (Equation (20));
3) That the algal concentration, X, at this point in the bloom is 50 mg/l;

Figure 7. Effect of temperature on dehydration rate constant for carbonic acid.
(4) That the specific growth rate of the algae, $u$, is 0.3 day$^{-1}$; and
(5) That the carbon content of the algal cells is 50 percent of the total algal dry weight.

Thus the change in algal concentrates per unit time, $dX/dt$, can be described as

$$\frac{dX}{dt} = \mu X = 15 \text{mg} \cdot \text{l}^{-1} \cdot \text{day}^{-1} = 1.74 \times 10^{-4} \text{mg} \cdot \text{l}^{-1} \cdot \text{sec}^{-1} \quad \text{(21)}$$

and the change in carbon, $C$, per unit time transformed into algal biomass is

$$\frac{dC}{dt} = -0.5(\mu X) = 8.7 \times 10^{-5} \text{mg} \cdot \text{sec}^{-1} \quad \text{(22)}$$

Converting the carbon content to an equivalent molar concentration of bicarbonate, $(HCO_3^-)$

$$\frac{d(HCO_3^-)}{dt} = 7.24 \times 10^{-9} \text{moles} \cdot \text{sec}^{-1} \quad \text{(23)}$$

Fifty mg/l of CaCO$_3$ alkalinity equals 61 mg/l of HCO$_3^-$ alkalinity, or $1 \times 10^{-3}$ M of HCO$_3^-$. Thus, using Rabinowitch's value of $0.47 \times 10^{-4}$ sec$^{-1}$ for the dehydration rate constant, the ratio of carbon dioxide available to carbon dioxide utilized at 25$^\circ$C is approximately 6:1. With Kern's dehydration rate constant of $2 \times 10^{-4}$ sec$^{-1}$ the ratio is approximately 28:1. Under the conditions described the dehydration step is definitely not rate limiting. These calculations are very crude because other than the arbitrary nature of the values chosen no accounting was made for temperature and activity effects, nor was carbon dioxide diffusion from the atmosphere considered. However, the values chosen would constitute a severe algal bloom situation (e.g. Mackenthun et al., 1968). Also it was assumed that the pH had already reached 10. Most blooms start at a considerably lower pH value when the faster dehydration reaction, $H_2CO_3 + CO_2 + H_2O$, would predominate. The lower pH values would make more HCO$_3^-$ readily available for conversion to CO$_2$ than indicated by the calculation for a pH value of 10.

Another important factor which strengthens the argument against $H_2CO_3$ dehydration being a rate-limiting step for carbon utilization by growing algae in a bloom condition is the presence of the enzyme carbonic anhydrase which catalyzes the dehydration reaction.

The role of the enzyme carbonic anhydrase

Carbonic anhydrase is the enzyme responsible for catalyzing the dehydration of $H_2CO_3$ during human respiration (Meldrum and Roughton, 1933). Edsall and Wyman (1958) have reported that carbonic anhydrase has a very high activity, being able to double the rate of dehydration of $H_2CO_3$ when present in minute concentrations of 1 $\mu$g/l or less. While its role in human respiration is quite well defined, little information is available on how it affects $H_2CO_3$ dehydration in algal systems. Both Steemann Nielsen and Kristiansen (1949) and Osterlind (1950b) tried to explain the direct utilization of HCO$_3^-$ in certain aquatic plants and algae by demonstrating a lack of this enzyme. However, this enzyme was found in all the species they examined, regardless of the species' ability to use HCO$_3^-$, and they could not draw
positive conclusions regarding its role. Litchfield and Hood (1964) demonstrated that the enzyme was present in 11 fresh water and marine algae. They showed that the enzyme was located within the soluble portions of the inner cell and that heating had an adverse affect on activity. Nelson et al. (1969) found that the carbonic anhydrase concentration in *Chlamydomonas* was 20 times greater when the cells were grown on 0.03 percent CO₂ as compared to 1 percent CO₂. This suggests that carbonic anhydrase participation in the dehydration of H₂CO₃ becomes more important when the pH of an algal system not at equilibrium with atmospheric CO₂ increases. The H₂CO₃ is readily derived from the ionic inorganic forms, but also the enzyme may mediate the direct dehydration of HCO₃⁻ via Equation (20).

While there is no direct evidence that the enzyme is extracellular, Berger and Libby (1969) suggested that the presence of this enzyme in the oceans could be a significant factor in equilibrating atmospheric CO₂ with the CO₂ content of sea water. Also, Krishnamurty (1969) reported the possibility of metal carbonate complexes aiding in this enzymatic process. Forster and Edsall (1969) reviewed a recent conference on the chemical, biological, and physiological aspects of CO₂ and stated that CO₂ dissolved most readily in solvents such as acetone. These compounds contain both hydrophobic and polar groups. Enzymes such as carbonic anhydrase could have a similar effect on CO₂ which could help in explaining their effect on the kinetics of CO₂ assimilation by algae.

It is known that zinc is required for activation of carbonic anhydrase (Edsall and Wyman, 1958). Riepe and Wang (1967) partially explained the role of zinc with the hypothesis that HCO₃⁻ is dehydrated to CO₂ when there is a breaking of a C-O bond in HCO₃⁻. A resulting proton transfer causes an OH⁻ to be coordinated to Zn²⁺ with the result that the OH⁻ is separated from the enzyme.

What specific role the enzyme has in relation to the uptake of inorganic carbon by algae in natural waters still remains to be answered. The very fact that its presence has been demonstrated in many algal systems suggests that it is intimately involved in providing carbon for algal growth when the free CO₂ content of a water is low. However, it would appear that to be effective in the uptake of inorganic carbon the carbonic anhydrase would have to operate externally to the cell. Further research on this aspect of inorganic carbon utilization is necessary.
CARBON UTILIZATION BY ALGAE

The photosynthetic reaction described in Equation (1) shows the formation of a carbohydrate product which represents a carbon content, on a dry weight basis, of over 50 percent. Generally, most values given in the literature indicate a carbon content for algal cells very close to this value. Ketchum (1954) reported values of 51 to 56 percent for various algae grown under continuous illumination. Parsons et al. (1961) showed a range from 15.9 to 53.2 percent for a variety of marine phytoplankton. Thus, the reported carbon content of various freshwater and marine algae does vary (Table 3). This variation is a function of the species and the environment of cultivation.

Sources of Carbon

CO₂, HCO₃⁻, CO₃²⁻, and organic compounds can all serve as the source of carbon for algae under specific conditions. Under normal conditions where free CO₂ is available it is often the preferred form of carbon used by the photoautotrophic algae (Myers, 1951). A number of researchers indicate that bicarbonates and carbonates may also be used directly (Holm-Hansen, 1967). Also, there is extensive information to support the notion that heterotrophic and chemotrophic activity by algae is widespread.

Organic carbon utilization by algae

Many phytoplankton algae exhibit heterotrophic, chemotrophic, and autotrophic types of metabolism (Ketchum, 1954). Some species may exhibit all three metabolic types under appropriate conditions. Chemotrophic assimilation of carbon by algae has been discussed by Bristol-Roach (1926, 1927), Barker (1935), Pearsall and Bengry (1940), Doyle (1943), Algeus (1946), Hunter and Provasoli (1951), Myers (1951), Fogg (1953), Lewin (1963).

Myers (1951) reported that although many researchers had shown heterotrophic growth of algae through the addition of organic carbon, none was able to demonstrate that this growth occurred when CO₂ was not limiting. With adequate CO₂ it appears that the photosynthetic mechanism is the predominant one. Lange (1967) suggested that organic matter converted to CO₂ through bacterial respiration enhanced algal activity, and that this process was a major factor related to algal blooms in natural waters. However, his work was performed under conditions of relatively high nutrient and low CO₂ concentrations and was not comparable to natural conditions nor to a control where CO₂ was added in excess.
Many other workers have demonstrated both the uptake and release of organic carbon compounds by algae (Ward et al., 1964, Fogg, 1965, Fogg et al., 1965, Ward and Moyer, 1966, Vaccaro and Jannasch, 1967, Smith et al., 1968a, 1968b, Litch-

Table 3. Variation in carbon content of algal cells of various species.

<table>
<thead>
<tr>
<th>Specie</th>
<th>Carbon Content % - ut. basis</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Fresh Water Algae</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Chlamydomonas</td>
<td>47.46</td>
<td></td>
</tr>
<tr>
<td>Anagaenopsis</td>
<td>49.57</td>
<td></td>
</tr>
<tr>
<td>Oikomonas termo</td>
<td>51.76</td>
<td></td>
</tr>
<tr>
<td>Stichococcus bacillaris</td>
<td>52.65 57.14</td>
<td>Milner (1953)</td>
</tr>
<tr>
<td>Chlorella pyrenoidosa</td>
<td>49.51 70.17</td>
<td></td>
</tr>
<tr>
<td>Stichococcus bacillaris</td>
<td>53.82</td>
<td></td>
</tr>
<tr>
<td>Chlorella pyrenoidosa</td>
<td>54.89</td>
<td></td>
</tr>
<tr>
<td>Chlorella vulgaris</td>
<td>53.37</td>
<td></td>
</tr>
<tr>
<td>Scenedesmus obliquus No. 1</td>
<td>54.55 54.87</td>
<td>Ketchum and Redfield (1949)</td>
</tr>
<tr>
<td>Scenedesmus obliquus No. 2</td>
<td>54.55 54.87</td>
<td></td>
</tr>
<tr>
<td>Scenedesmus basilensis</td>
<td>54.82</td>
<td></td>
</tr>
<tr>
<td>Nitzschia closterium</td>
<td>54.98</td>
<td></td>
</tr>
<tr>
<td><strong>Marine Algae</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tetraselmis maculata</td>
<td>35.1, 36.8</td>
<td></td>
</tr>
<tr>
<td>Dunaliella salina</td>
<td>39.6, 45.4</td>
<td></td>
</tr>
<tr>
<td>Monochrysis lutheri</td>
<td>53.2, 44.6</td>
<td></td>
</tr>
<tr>
<td>Syracosphaera carterae</td>
<td>34.6, 35.2</td>
<td></td>
</tr>
<tr>
<td>Chaetoceros sp.</td>
<td>31.0</td>
<td>Parsons et al. (1961)a</td>
</tr>
<tr>
<td>Skeletonema costatum</td>
<td>26.0, 31.3</td>
<td></td>
</tr>
<tr>
<td>Coscinodiscus sp.</td>
<td>14.9, 18.8</td>
<td></td>
</tr>
<tr>
<td>Phaeodactylum tricornutum</td>
<td>37.5, 49.5</td>
<td></td>
</tr>
<tr>
<td>Amphidinium carteri</td>
<td>40.7, 47.1</td>
<td></td>
</tr>
<tr>
<td>Exuviella sp.</td>
<td>44.5, 47.4</td>
<td></td>
</tr>
<tr>
<td>Agmenellum quadruplicatum</td>
<td>42.0, 44.9</td>
<td></td>
</tr>
<tr>
<td>Gigartina agardhii</td>
<td>42.90</td>
<td>Milner (1953)</td>
</tr>
<tr>
<td>Ulva sp.</td>
<td>44.36</td>
<td></td>
</tr>
<tr>
<td>Amphilpleura rutilans</td>
<td>46.52</td>
<td></td>
</tr>
<tr>
<td>Macroystis pyrifera</td>
<td>46.02</td>
<td></td>
</tr>
<tr>
<td>Navicula torquatum</td>
<td>48.45</td>
<td></td>
</tr>
<tr>
<td>Egregia menziesii</td>
<td>49.97</td>
<td></td>
</tr>
</tbody>
</table>

a Samples analyzed by two different methods
field et al., 1969). Fogg (1965) and Fogg et al. (1965) showed that glycolic acid was a major excretory product of phytoplankton and that production of this acid increased under high light intensity and carbon dioxide deficiencies. Glycolic acid could be used as a carbon source by bacteria and algae and thus could be a carbon source in the regeneration of CO$_2$ for photosynthetic growth. This concept suggests that a portion of the CO$_2$ utilized by algae during bloom conditions is actually derived from bacterial activity. This CO$_2$ may be cycled or introduced from organic carbon sources (breakdown of allochthonous material, dissolved organics, decay of autochthonous production). Kuentzel (1969) presented the hypothesis that bacterial production of CO$_2$ is the major causative factor in many algal blooms. Although Kuentzel's argument was based primarily on the work of Lange (1967), he failed to acknowledge the significance of other sources of inorganic carbon, primarily atmospheric CO$_2$ and bicarbonate. Also no mention was made of the apparent ability of certain algae to utilize the HCO$_3^-$ directly.

Smith et al. (1960) reported that complexed CO$_2$ in carboxylic acids was more readily available for assimilation by algae than was dissolved CO$_2$. Steemann Nielsen (1963) criticized this on the basis that under the conditions of the experiment, $^{14}$CO$_2$ assimilation was inhibited, not by the preferential use of CO$_2$ from the complex, but rather by the release of NH$_3$ from the complex at the high pH values experienced; the NH$_3$ thus released could act as a poison to affect cell metabolism.

The significance of the role of organic carbon compounds in the occurrence of algal blooms following cultural eutrophication is at present uncertain, as evidenced by the many conflicting reports in the literature. Careful consideration will be given to the utilization of inorganic carbon by algae because the use of this form of carbon by photosynthesizing algae is better understood and it obviously has a significant role in algal growth during the eutrophication process.

**Inorganic carbon utilization by algae**

The early work on inorganic carbon utilization by algae from the late 1930's through the early 1950's was performed by researchers investigating the phenomena governing the photosynthetic mechanism. The main theme of this work was to determine which forms of carbon preferentially stimulated algal growth. At this time eutrophication was an unrecognized problem in world wide terms.

As mentioned previously in this paper, CO$_2$ activity rather than concentration would provide more accurate interpretation of results especially for some of the high nutrient concentrations which are used as culture media. None of the investigations described in this section have measured the activity of the chemical constituents described and thus the results cannot be utilized unequivocally. Although the results are not invalidated, activity of nutrients should be considered in such investigations as one might suspect that growth rates (e.g. see Monod, 1949) would be a function of activity rather than molar concentrations. Perhaps, the advent of selective ion electrodes will help in further investigations.

Emerson and Green (1938) working with *Chlorella*, a favorite test alga of early researchers, found in manometric studies that saturation with CO$_2$ in concentrations
ranging from 0.05 percent to 5 percent of atmospheric air caused little change in the rate of photosynthesis over a pH range from 4.6 to 8.9. These results were criticized by Steemann Nielsen (1952) on the basis that allowances were not made for carbon reserves present in the algae cells at the start of the experiment.

In a most convincing series of experiments Osterlind (1949, 1950a, 1950b) and Steemann Nielsen (1951, 1952, 1953, 1955a, 1963) demonstrated that HCO₃⁻ was preferred by Scenedesmus; whereas dissolved CO₂ was the only inorganic carbon source available to certain other species, notably Chlorella. These and other results were reviewed quite thoroughly by Fogg (1953) in his monograph on algal metabolism and by Rabinowitch (1945, 1951, 1956) in his three volume treatise on photosynthesis. Osterlind (1949, 1950a, 1951a) demonstrated that Chlorella was capable of utilizing only dissolved CO₂ while young Scenedesmus cultures could utilize both dissolved CO₂ and HCO₃⁻ and seemed to utilize the latter more effectively. Old Scenedesmus cultures lost their ability to utilize HCO₃⁻ and showed a response similar to Chlorella. Further experiments (1950b) showed that there was little difference in the concentration of the enzyme, carbonic anhydrase, responsible for dehydration of H₂CO₃ in both groups of algae, leading Osterlind to the conclusion that a true utilization of HCO₃⁻ by Scenedesmus took place. Osterlind (1951b) later proposed the theory that young Scenedesmus uniquely possessed an enzyme system which upon “photoactivation” catalyzed direct HCO₃⁻ utilization. Activation either took place at the cell wall surface or within the cell itself with the enzyme in the latter case being simply carbonic anhydrase.

Briggs and Whittingham (1952) substantiated Osterlind’s claim that Chlorella could only utilize dissolved CO₂. Steemann Nielsen (1952) working with higher aquatic plants demonstrated a similar action in certain species. He showed a definite utilization of HCO₃⁻ on one side of an aquatic leaf and liberation of OH⁻ ions on the other side according to the equation HCO₃⁻ = OH⁻ + CO₂. He later presented evidence (Steemann Nielsen and Jensen, 1958) that the HCO₃⁻ was capable of penetrating the Chlorella cell surface but that the rate of this penetration was so low as to be insignificant. Felfoldy (1960a) demonstrated HCO₃⁻ utilization by Chlorella but noted that utilization occurred only after long periods of adaptation. He also indicated there was a possible direct utilization of CO₃⁻ by certain algae at very high pH values although no concrete evidence to support this notion was presented (1960b).

Later attempts by Hood and Park (1962) to show that certain marine phytoplankton including a marine Chlorella sp. could also utilize HCO₃⁻ were severely criticized by Watt and Paasche (1963) and by Steemann Nielsen (1963). Hood and Park claimed that, aside from respiratory CO₂ production, CO₂ derived from the dehydration of H₂CO₃ and from diffusion of atmospheric CO₂ were the only sources of carbon available to algae for autotrophic growth. They felt that diffusion from the atmosphere was so slow as to be an insignificant source of carbon. Also they made the claim that dehydration was slow enough to likewise cause carbon limitations. On the basis of the latter hypothesis they purported to show that HCO₃⁻ was the main source of carbon for certain algal species at very low dissolved CO₂ concentrations. Both Watt and Paasche (1963) and Steemann Nielsen (1963) showed by theoretical and experimental work that the “slow hydration rate” was indeed fast enough to prevent distinguishing between CO₂ and HCO₃⁻ utilization. Thus no valid conclusions could be drawn from Hood and Park’s work. Briggs (1959)
claimed that in certain instances the apparent direct utilization of $\text{HCO}_3^-$ could really be explained by its affect on the $\text{CO}_2$ concentration at the point where consumption was taking place. $\text{HCO}_3^-$ was thus acting solely as a reservoir for $\text{CO}_2$. Paasche (1963, 1964) in other work showed that coccolith formation in the coccolithophorid *Coccolithus huxleyi* resulted solely from utilization of $\text{HCO}_3^-$. The rate of photosynthesis, on the other hand, was a function of $\text{HCO}_3^-$ and/or the dissolved $\text{CO}_2$ present. Steemann Nielsen (1966) further amplified these conclusions by showing that a naked clone of the same specie utilized only $\text{CO}_2$ for photosynthesis but that $\text{HCO}_3^-$ was made directly available through the coccolith formation mechanism. Swift and Taylor (1966) showed an optimum pH of 7.8 for cell division of the coccolithophorid *Cricosphaera elongata*.

Witt and Borchardt (1960) and Gates and Borchardt (1964) working with continuous algal cultures showed that when the $\text{CO}_2$ supply was turned off to a relatively small concentration of growing *Scenedesmus* the pH rose from 8.3 causing an immediate lag phase in the production rate while the algae readjusted to using $\text{HCO}_3^-$ as their source of carbon. After this readjustment the production rate continued as before until the culture failed at a pH of 10. In a similar experiment with a much more concentrated culture of *Scenedesmus* no lag phase occurred when the $\text{CO}_2$ supply was stopped and the algae had to convert to using $\text{HCO}_3^-$. The authors explained this occurrence by stating that with the low algal concentration carbon never was limiting and the algae were using only dissolved $\text{CO}_2$ before the supply was stopped. In the latter case carbon was limiting even before the $\text{CO}_2$ supply was stopped and the algae were forced to use both dissolved $\text{CO}_2$ and $\text{HCO}_3^-$. No adjustment was thus necessary for conversion to $\text{HCO}_3^-$ once the $\text{CO}_2$ supply was removed. However, Soltero and Lee (1967), demonstrating an automatic pH control device for algal cultures, gave evidence that at a pH of 7 optimum growth of *Scenedesmus* occurred.

This was in contrast to the work of Witt and Borchardt (1960) and Gates and Borchardt (1964) who could show little change in the growth of *Scenedesmus* over a wide pH range although best growth was observed at a pH of 8.3. A possible explanation for these differences might be that Witt, Gates, and Borchardt used wastewater treatment plant effluent for their substrate in which the total alkalinity was 120 mg/l (as CaCO$_3$), while Soltero and Lee used a synthetic medium in which the bicarbonate alkalinity was 21 mg/l (as CaCO$_3$). Thus, the apparent difference in optimum pH for growth of *Scenedesmus* might be accounted for by the fact that in the case of Soltero and Lee, little $\text{HCO}_3^-$ was available over the entire pH range, and that as the pH was lowered more carbon in the form of $\text{CO}_2$, which could also be used by the *Scenedesmus*, was present since pH adjustment was controlled through $\text{CO}_2$ addition. At a pH of 6, however, less growth was observed. Although more dissolved $\text{CO}_2$ would be present at a pH of 6, the poorer growth at this pH value as compared to that at a pH of 7 could be explained by an adverse physiological effect on the algae of this lower pH. In the case of Witt, Gates, and Borchardt the high alkalinity contributed the maximum amount of $\text{HCO}_3^-$ at a pH of 8.3, which is the point in the $\text{CO}_2$ equilibrium system where essentially all the inorganic carbon is $\text{HCO}_3^-$. Thus, the algae were utilizing primarily $\text{HCO}_3^-$ and growing best at this pH. Brown (1969) growing algae on agricultural drainage water in cultures in which the pH was controlled with “Good” organic acid buffers (Good et al., 1966), and which had a total alkalinity of 350 mg/l (as CaCO$_3$), achieved best growth at a pH of 8.4 (as...
compared to other tested pH values of 6.15, 7.5, and 10.5). His results are in close agreement with those obtained by Witt, Gates, and Borchardt.

In most of the experiments dealing with carbon effects on *Chlorella* it was found that a CO₂ concentration of from 0.01 to 0.03 percent was adequate for maintaining the maximum rate of photosynthesis (Emerson and Green, 1938, Briggs and Whittingham, 1952, Steemann Nielsen, 1953, 1955a). Since approximately 0.03 percent CO₂ is found in atmospheric air, sufficient carbon for maximum photosynthesis of *Chlorella* should be present when a culture is exposed to and in equilibrium with atmospheric CO₂. Obviously then for most culture devices in which little or no turbulent mixing is maintained, carbon is extracted from solution at a faster rate than it can diffuse into solution from the atmosphere. A carbon limitation is then created, placing an upper limit on the rate of photosynthesis.

The term “rate of photosynthesis” refers to the rate at which a given algal cell produces oxygen or fixes carbon dioxide, while the term “rate of growth,” refers to the rate of algal mass produced or the more commonly used term “specific growth rate,” refers to the rate of algal mass production per cell concentration.

The percent CO₂ in air is a relatively meaningless term in trying to describe the amount of CO₂ required for maximum photosynthesis in culture work if no accounting is made for the concentration of CO₂ in solution which is really available to the algae. The amount really available is a function of the sparging rate and its effect on the CO₂ tension.

Steemann Nielsen (1955a) in referring to the high CO₂ concentrations (0.2 - 1.0 percent) required by Osterlind (1951a) and by Rosenberg (1954) to achieve maximum photosynthetic rates with *Scenedesmus* and *Chlorella* claimed that in both cases a dense algal concentration caused enough competition for the available carbon so that maximum photosynthetic rates could be measured only at these high CO₂ concentrations. Thus, the cultures were actually carbon limited at the low CO₂ concentrations and no true measurements of maximum photosynthetic rates were possible.

Rosenberg (1957) answered Steemann Nielsen’s criticism by stating that CO₂ limitations were only important under high light intensities. With dense algal cultures the effect of light shading should have caused even greater carbon limitations than he observed. He did point out, however, that discrepancies among researchers as to optimum CO₂ concentrations for maximum photosynthesis might be explained by the fact that long adaptation periods seem to be required when switching from high to low CO₂ concentrations. Rosenberg’s experiments only lasted for minutes, which might explain his failure to note high photosynthetic rates at low CO₂ concentrations. Steemann Nielsen and Willemoes (1966) did not observe any differences in the photosynthetic rate of *Chlorella* with either air or 5 percent CO₂ in air as the carbon source.

Myers (1953) reported that CO₂ concentrations between 0.1 and 5 percent were adequate to prevent the growth rate from becoming carbon limited. It appears from the various reports reviewed on the subject of CO₂ concentration in algae cultures that often optimum levels of CO₂ for maximum photosynthetic rates and
for maximum growth rates are claimed to be the same (Steemann Nielsen, 1955a). Obviously the required concentration of carbon dioxide in solution is a function of the desired algal production, the denser the culture, the more CO₂ required. Myers (1953) states that a 5 percent CO₂ concentration is adequate for growth situations in which the CO₂ uptake is high, and as a result there is a large diffusion gradient between the culture and the gas phase. The 5 percent CO₂ added will maintain enough CO₂ in solution so that carbon is not limiting. Davis et al. (1953) substantiated Myers' claim and found no difference in algal growth rate at CO₂ concentrations in air from 0.56 to 4.43 percent.

Based upon the work of Myers (1953), Davis et al. (1953), and other researchers who contributed to the classic Carnegie Institute publication on the mass culturing of algae, most subsequent studies on algae culturing indicate that CO₂ concentrations from 1 to 5 percent are adequate. The work of Pipes and Koutsoyannis (1962) and Miller (1968) are representative examples.

Myers (1953), Steemann Nielsen (1953, 1955a), Fogg and Than-Tun (1960, 1965) have all reported toxic effects at high CO₂ concentrations. Myers and Fogg (1953) claimed that CO₂ concentrations over 5 percent were to be avoided. Steemann Nielsen (1953) showed that in saturated light, CO₂ concentrations greater than 1 percent were toxic, while in unsaturated light CO₂ assimilation rates increased at CO₂ concentrations greater than 1 percent. Further work by Steemann Nielsen (1955a) demonstrated that under certain saturated light conditions, pH affected CO₂ assimilation as a function of CO₂ concentration. Lower pH values tended to give increasing CO₂ assimilation rates for CO₂ concentrations greater than 1 percent. Fogg and Than-Tun (1960) and Fogg (1965) have demonstrated the existence of a toxic effect on a blue-green alga, Anabaena, at a CO₂ concentration over 1 percent. However, Tew et al. (1962) in trying to reduce volume requirements for a photosynthetic gas exchanger for space application by utilizing 100 percent CO₂ as the carbon source, were able to sustain a culture for a considerable period of time by providing the CO₂ at a flow rate corresponding to the algae production rate such that all the carbon was assimilated. Although the unit eventually failed, the authors attributed this failure to the physical design of the continuous culture device and not necessarily to any toxic effects resulting from the use of 100 percent CO₂.

Pipes (1962a) along with Tew et al. (1962) have performed the only meaningful experiments in the United States with continuous cultures which describe the kinetic relationships of carbon dioxide limited growth of Chlorella. Pipes, using the kinetic growth model for chemostatic cultures proposed by Monod (1950) and Novick and Szilard (1950), and reviewed in detail by Herbert, Elsworth, and Telling (1956), showed that when a constant supply of CO₂ is provided the algal production rate within a range of dilution rates (or residence times) was independent of the dilution rate and linearly proportional to the rate of CO₂ supply (by doubling the flow rate of a fixed concentration of CO₂-enriched air the algal production rate was doubled).

In Czechoslovakia extensive work by the Institute of Microbiology of the Czechoslovak Academy of Sciences on the mass culturing of algae has been in progress for a number of years (Setlik et al., 1966, Annual Report, 1967, 1968). A major concern has been the solution to problems associated with carbon limitations on
growth. Various techniques for efficiently supplying CO$_2$ have been investigated. Work in Russia dealing with carbon effects on algal growth also is reported in the literature (Levshina, 1965, Semenenko, 1966).

**Carbon Utilization in Special Systems**

The incorporation of carbon by algae in the aquatic environment occurs in three general systems: wastewater treatment systems such as oxidation ponds, natural water bodies (i.e. the oceans, lakes, impoundments, rivers, etc.), and controlled laboratory facilities (e.g. batch and chemostat experiments). In natural systems, visible concentrations of algae are usually undesired and when present in sufficient numbers constitute a nuisance condition leading to serious impairment of beneficial uses of the waters. In the treatment systems the growth of algae is encouraged since treatment efficiency is dependent on the oxygen supplied by photosynthetic organisms. In both natural and waste treatment systems the introduction of chemical nutrients induces algal growth which will be limited by the one factor in shortest supply. It appears from reports in the literature that carbon is often the growth limiting nutrient in oxidation ponds but that its role as a limiting nutrient for algal growth in natural waters is not as well documented.

**Wastewater treatment systems**

In the early work performed at the University of California in developing algal-bacterial systems for wastewater treatment, studies (Ludwig et al., 1951) dealing with the algae species *Euglena* showed that the addition of supplementary CO$_2$ to a settled, sterilized sewage medium had a positive effect on cell concentration and yield. The addition of air enriched with 2.3 percent CO$_2$ increased the cell concentration and yield considerably as compared to the addition of atmospheric air (0.03 percent CO$_2$). This study indicated that carbon might be the growth limiting factor for sewage-grown algae. Other studies in this research endeavor confirmed the suspicion that the lack of carbon had limited algal growth in oxidation ponds (Oswald et al., 1953a, 1953b). Further work by Oswald (1963) demonstrated that maximum light conversion efficiencies were obtained with CO$_2$ at a concentration of 1 to 2 percent. However, nitrogen was depleted at a CO$_2$ concentration of 2 percent and thus became limiting in cultures grown in higher CO$_2$ concentrations. It is possible that even greater efficiencies would have been attained at the higher CO$_2$ concentrations had enough nitrogen been provided.

Although the work at the University of California was the first to elucidate the need for supplementary carbon in waste treatment systems, other researchers have since reached a similar conclusion. Studies by Allen (1955) on algal growth with sewage effluents demonstrated a marked increase in growth when 5 percent CO$_2$ was bubbled through the cultures. Allen noted that both *Chlorella* and *Scenedesmus* were the prominent algae species found in stabilization ponds and that, while in batch cultures, *Chlorella* would at first be the predominant species, it was superseded by *Scenedesmus*. Although Allen did not offer an explanation for this particular succession, it would seem that results obtained in the work of Osterlind, Steemann Nielsen and others on the preferential use of HCO$_3^-$ as a carbon source for *Scenedesmus*, as compared to that for *Chlorella*, offer a logical explanation for this changeover. Since Allen's cultures were experiencing a pH rise (up to values of 8 to 10, levels at which
the predominant carbon form is $\text{HCO}_3^-$), the possibility exists that Scenedesmus replaced Chlorella due to its ability to utilize $\text{HCO}_3^-$ directly.

Meffert (1955) working with algae grown on sewage also added $\text{CO}_2$ to improve algal growth, although her reasoning for this action is open to question. She claimed that because $\text{CO}_2$ production from bacterial respiration was not sufficient, a rise in pH resulted in the predominance of $\text{HCO}_3^-$ and $\text{CO}_3^{2-}$. The possibility that a simple carbon limitation existed which bacterial production of $\text{CO}_2$ could not overcome, regardless of whether the Scenedesmus were utilizing $\text{HCO}_3^-$, as claimed by others was not explored. This limitation was possibly corrected once supplementary $\text{CO}_2$ was added to the cultures.

In their work with algal nutrient removal systems for sewage effluents Fitzgerald (1961) and Fitzgerald and Rohlich (1962) showed that carbon usually becomes limiting before nitrogen, and that the addition of extra carbon as $\text{CO}_2$ increased yields and shortened the time for nitrogen assimilation in batch cultures. Brown and Arthur (1969) achieved similar results in trying to remove nitrate-nitrogen from agricultural tile drainage through assimilation into algal cellular material. However, the drainage water also required phosphorus and iron additions. Bush et al. (1961) working with a continuous outdoor algal growth system added 8 percent $\text{CO}_2$ in order to achieve optimum nutrient removal from sewage effluents.

Fitzgerald and Rohlich (1962) also demonstrated the effect of $\text{CO}_2$ addition on the maintenance of phosphorus solubility in algal cultures by preventing the removal of the phosphorus through precipitation at elevated pH values. Bogan et al. (1960), Bogan (1961), and Hemens and Mason (1968) likewise demonstrated the relationship between rise in pH level and phosphorus removal in algal wastewater treatment systems.

Mattoni et al. (1967), working with integrated algal systems which included wastewater treatment, water reclamation and protein production, controlled pH through acid addition. They observed alkalinity losses in semi-continuous algal reactors which they attributed to ammonia assimilation by algae. Pipes (1962b) reported that a similar reaction took place in algal waste treatment ponds, but only at a relatively high initial pH and alkalinity. At a lower initial pH and alkalinity a reverse reaction occurred, with the alkalinity in the effluent being higher than in the influent, although in both cases the pH rose over the initial value. No hypothesis was offered to explain this phenomenon. Pipes and Gotaas (1960) also showed that Chlorella, using sewage as a substrate, utilized organic carbon as a carbon source and that this utilization became more pronounced when the inorganic carbon supply was limited. Residence time and hence age of cells appeared to be the contributing factor in the excretion of dissolved organic material by the algae. At a residence time less than 3 days a net decrease in dissolved organic material took place in the growth units, while at a residence time greater than 3 days excretion exceeded assimilation of organic material.

Both Witt and Borchardt (1960) and Gates and Borchardt (1964) in their studies on nitrogen and phosphorus removal from wastewater treatment plant effluents by the controlled culture of algae, realized the need for adding supplementary $\text{CO}_2$. King (1970) has also shown the necessity of carbon supplementation in waste-
water oxidation ponds. Beck et al. (1969) showed that CO₂ addition was necessary for complete NO₃-N assimilation by algae grown on agricultural tile drainage in the San Joaquin Valley of Central California.

**Natural water bodies**

Sawyer (1954) in an early paper suggested that algal blooms could be controlled through carbon limitation if atmospheric CO₂ were the only source of inorganic carbon available to algae. However, as he pointed out, additional carbon from the alkalinity and from bacterial CO₂ production provides an ample reservoir of inorganic carbon. Sawyer also recognized the importance of pH on algal growth, and mentioned that pH values up to almost 10 were observed in water bodies where algal growth was active.

Ketchum (1954) made a similar claim that carbon in natural waters was rarely, if ever, limiting. However, Steemann Nielsen (1955b) through his work on eutrophic Danish lakes, felt that high pH values resulting from CO₂ assimilation would limit algal activity during summer conditions. Jackson (1964) stated that blue green algal blooms occurred only when the pH value was greater than 8.5. He disagreed with the long held theory that blooms actually started at lower pH values and that a pH rise above 8.4 was only a consequence of the bloom condition and not a precipitating factor. He felt that blue green algae required an elevated pH value in order to proliferate. Since at a pH greater than 8.4 very little inorganic carbon is present as CO₂, perhaps the blue greens utilize HCO₃⁻ (as claimed for Scenedesmus) or even CO₃²⁻. He also felt that there might be a preferential use of dibasic orthophosphate (HPO₄²⁻) over the monobasic form (H₂PO₄⁻), since the former is essentially the only inorganic phosphate form present at a pH greater than 8.4. He concluded by offering a possible solution for preventing blue green algae from blooming: By keeping a lake water well buffered or so low in ions that a pH of 8.4 is never reached, blue green algae would never have the proper environment required for their profuse growth. A recent Russian paper by Merezhko (1968) claimed that several blue green algae utilized HCO₃⁻ as the carbon source.

**Controlled laboratory facilities**

Gerloff, Fitzgerald, and Skoog (1952) working with the blue green alga *Microcystis aeruginosa* cultured in an unbuffered medium showed that maximum growth occurred at a pH of 10. McLachlan and Gorham (1961) and McLachlan (1962) working with the same algal species, observed little change in growth in a pure culture over a pH range of 6.5 to 10 in a well-buffered medium. However, when they tried to grow this alga together with the green alga *Scenedesmus* at a pH of 7.4 they obtained less than one-third of the growth reached by the blue green alga in pure culture. Thus, there was a definite competitive effect at the lower pH, indicating that only at the higher pH values would the blue green alga predominate. Eberly (1967), working with *Oscillatoria agardhii*, another blue green algae grown in batch cultures, reported that the cultures with the highest initial pH values (up to 10) reached the exponential phase earliest, but that all cultures eventually reached the same level of maximum biomass. Holm-Hansen (1967) raised the question that since most blue green algae grow best at the higher pH values (8-11), and since at a pH of 11 most of
the carbon is in the form of \( \text{CO}_3^{2-} \), are the algae really utilizing this ion preferentially?

In a recent conference held in Berkeley, California, on eutrophication and biostimulation (Middlebrooks et al., 1969) the need was stressed in several papers for a more detailed and quantitative look at the role of \( \text{CO}_2 \) in relation to the problems of algal blooms. McIntire (1969) developed a laboratory model of a riffle area of a natural stream with water provided by a spring-fed creek. He found that the addition of \( \text{CO}_2 \) above that normally present in solution had a dramatic effect on algal growth at light intensities around 1000 foot-candles and greater. This would tend to indicate that either carbon was limiting at this high light intensity or else the addition of the \( \text{CO}_2 \) suppressed the pH enough so that certain nutrient salts (phosphorus or iron?) became more soluble thus stimulating growth. Wright (1969) demonstrated that \( \text{CO}_2 \) was the growth limiting factor for primary production in the Firehole River complex in Wyoming. Using the upstream-downstream \( \text{CO}_2 \) measurement method as described by Odum (1956) for determining primary productivity, Wright showed that changes in the photosynthetic rate were directly related to changes in the \( \text{CO}_2 \) content in the water along the stretch of river studied. He concluded by stating that streams receiving effluents with high organic loads could produce enough \( \text{CO}_2 \) through bacterial respiration to make this nutrient as important in causing algae blooms as any other nutrient.

The recent Provisional Algal Assay Procedures (1969) has stimulated a considerable amount of research into finding a suitable method for measuring the algae growth potential of a water body. In trying to develop a test with which the effect of one nutrient can be studied at a time it is important that all other nutrients be in excess. Porcella (1969) has shown the difficulty in trying to interpret data which indicate that more than one nutrient may be limiting at a given time. He found that a sizable increase in algal growth in continuous culture devices took place at a given phosphorus level when supplementary \( \text{CO}_2 \) was added. In the unit without \( \text{CO}_2 \) the pH was maintained around 10.3. Obviously \( \text{CO}_2 \) was limiting in this case and not phosphorus. Pearson et al. (1969), in comparing the batch type assay with the continuous culture assay, purported to show that the former was characterized by a maximum specific algal growth rate four times greater than that of the latter, in deference to theoretical considerations which would indicate that an opposite effect should occur. On the basis of these results it was concluded that the batch test was unreliable. In Pearson et al. tests, nitrogen was supposed to be the growth limiting factor. However, the maximum specific growth rate for the continuous culture test was around 0.2 day\(^{-1}\), considerably lower than theory would indicate for the type of organism and substrate tested, while the batch assay maximum growth constant was around 0.9 day\(^{-1}\), a rate closer to a typical value. The question then arises, was nitrogen really limiting in the continuous culture test, and if not, could a lack of \( \text{CO}_2 \) have caused this depressed growth constant? The continuous culture unit was closed to the atmosphere and relied upon slow air bubbling for agitation. The batch flasks, on the other hand, had a relatively large surface area to volume ratio (150 mL of medium in a 250 mL flask) and were continually swirled on a shaker table. Brown and Arthur (1969), as previously mentioned, demonstrated the effect of \( \text{CO}_2 \) addition and culture volume to flask size on algal growth. It would appear then that the results obtained in the Pearson study are inconclusive and further refinement
of the test procedures is in order so that there is assurance that only the nutrient of concern is truly limiting.

An important consideration in developing a standard algal assay procedure, which was not included in the original PAAP report, would be to have pH control through CO₂ addition over the growing cultures. Since reproducibility is a desired product of such a test it would appear that pH control would be an essential requirement when trying to compare data from different laboratories. Galloway and Krauss (1961) demonstrated this point and indicated that aside from physiological changes in algae due to pH changes, optimal density measurements for growth determinations could be affected by precipitate formation, which in turn would be a function of pH.

**Nutrient Utilization by Algae**

This discussion has been abbreviated because of the considerable literature available to the reader and because of the great variety of possible conditions which can exist. The list of critical (i.e., most likely to be limiting) nutrients was defined on the basis of an algal growth medium (Algal Assay Procedures, 1971) which can be used for culturing most phytoplankton (Table 4). Some of the cations (sodium, potassium, magnesium) and anions (chloride, sulfate) necessary as elements for algal growth in that medium are generally in plentiful supply in natural waters and thus have been excluded from the list. Obviously, there is considerable variation in the composition of natural waters and these estimates can only be a comparative guide. There are numerous stoichiometric formulas for the elemental composition of algae (e.g. Ketchum, 1954, Stumm and Morgan, 1962, McCarty, 1970) but as shown in Tables 5 and 6, it is impossible to assign an exact elemental relationship to plant life in general. Moreover, within a single species of algae maintained under different environmental conditions, the elemental composition can be quite variable. For example, Porcella et al., 1970, have shown that the utilization of nitrogen and phosphorus per unit of cells produced in batch cultures of *Selenastrum capricornutum* varies by a factor of 5 for nitrogen and 30 for phosphorus for conditions of high growth rate to zero growth rate and nitrogen limiting conditions to phosphorus limiting conditions. Therefore, definition of a limiting nutrient based on estimates of nutrient composition in water or on estimates of nutrient composition in the cells is not feasible. Actual definition of those factors responsible for controlling algal blooms in a specific body of water requires considerable study of all the factors which can govern growth of algal population: light, temperature, macronutrients and micronutrients, mixing, predation, settling and resuspension, and the diurnal and seasonal variation in those factors.

**Role of Carbon in Eutrophication Processes**

Kuentzel (1969), in attempting to demonstrate the general lack of inorganic carbon in natural waters, quotes Hutchinson (1957) as saying that the free carbon dioxide content of a natural water in equilibrium with atmospheric CO₂ will normally be between 0.4 and 1.0 mg/l. While this statement is true, it already has been pointed out that many natural waters are supersaturated with CO₂ as evidenced by their low pH values relative to their total alkalinitities. As an example, pH and alkalinity values for a variety of lakes throughout North America are listed in Table 7.
Table 4. Critical nutrient concentrations of various waters.  

<table>
<thead>
<tr>
<th>Nutrients (µg/l)</th>
<th>Estimated Averages</th>
<th>Specific Examples of Waters Where Algal Growth Occurs</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>World River Waters¹</td>
<td>Selected, Temperate World Lakes³</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Lake Tahoe, California (Oligotrophic)⁴</td>
</tr>
<tr>
<td></td>
<td></td>
<td>River (near New Orleans)⁵</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Raw Sewage⁶</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Algal Growth Medium⁷</td>
</tr>
<tr>
<td>Total inorganic carbon</td>
<td>12000</td>
<td>23000⁶</td>
</tr>
<tr>
<td></td>
<td>10000</td>
<td>10000</td>
</tr>
<tr>
<td></td>
<td>4000</td>
<td>38000</td>
</tr>
<tr>
<td></td>
<td>2120</td>
<td></td>
</tr>
<tr>
<td>Total nitrogen</td>
<td>220⁵</td>
<td>200</td>
</tr>
<tr>
<td></td>
<td>33⁴</td>
<td>22⁵</td>
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<td></td>
<td>29000</td>
<td>42000</td>
</tr>
<tr>
<td></td>
<td>42000</td>
<td></td>
</tr>
<tr>
<td>Total phosphorus</td>
<td>70²</td>
<td>20</td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>--</td>
</tr>
<tr>
<td></td>
<td>11000</td>
<td>186</td>
</tr>
<tr>
<td>Total iron</td>
<td>670</td>
<td>50-250</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>60</td>
</tr>
<tr>
<td></td>
<td>132</td>
<td>33</td>
</tr>
<tr>
<td>Silica (as SiO₂)</td>
<td>13000</td>
<td>2000</td>
</tr>
<tr>
<td></td>
<td>---</td>
<td>7000</td>
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<tr>
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<td>1320</td>
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<td>Trace Elements</td>
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<td></td>
</tr>
<tr>
<td>Boron</td>
<td>20</td>
<td></td>
</tr>
<tr>
<td>Cobalt</td>
<td>0.02-2.0</td>
<td></td>
</tr>
<tr>
<td>Copper</td>
<td>100</td>
<td>0.35</td>
</tr>
<tr>
<td>Manganese</td>
<td>20-140</td>
<td>114</td>
</tr>
<tr>
<td>Molybdenum</td>
<td>&lt;1.0</td>
<td>2.88</td>
</tr>
<tr>
<td>Zinc</td>
<td>50-250</td>
<td>15</td>
</tr>
</tbody>
</table>


b Nitrate - N only

c Average of values presented in Table 7.

d Inorganic nitrogen only
Table 5. Chemical composition of some algae from ponds and lakes in southeastern
United States. 1

<table>
<thead>
<tr>
<th>Analysis</th>
<th>Chara</th>
<th>Pithophora</th>
<th>Spirogyra</th>
<th>Spirogyra</th>
<th>Rhizoclonium</th>
<th>Oedogonium</th>
<th>Mougeotis</th>
<th>Anabaena</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ash percent</td>
<td>43.4</td>
<td>27.77</td>
<td>13.06</td>
<td>13.86</td>
<td>17.36</td>
<td>12.69</td>
<td>14.54</td>
<td>5.19</td>
</tr>
<tr>
<td>C percent</td>
<td>29.3</td>
<td>35.38</td>
<td>42.40</td>
<td>41.16</td>
<td>39.10</td>
<td>40.84</td>
<td>40.74</td>
<td>49.70</td>
</tr>
<tr>
<td>N percent</td>
<td>2.46</td>
<td>2.57</td>
<td>3.01</td>
<td>2.35</td>
<td>3.46</td>
<td>2.64</td>
<td>1.77</td>
<td>9.43</td>
</tr>
<tr>
<td>P percent</td>
<td>0.25</td>
<td>0.30</td>
<td>0.20</td>
<td>0.23</td>
<td>0.43</td>
<td>0.08</td>
<td>0.25</td>
<td>0.77</td>
</tr>
<tr>
<td>S percent</td>
<td>0.55</td>
<td>1.42</td>
<td>0.27</td>
<td>0.24</td>
<td>0.27</td>
<td>0.15</td>
<td>0.36</td>
<td>0.53</td>
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<tr>
<td>Ca percent</td>
<td>8.03</td>
<td>3.82</td>
<td>0.57</td>
<td>0.84</td>
<td>0.52</td>
<td>0.44</td>
<td>1.68</td>
<td>0.36</td>
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<tr>
<td>Mg percent</td>
<td>0.92</td>
<td>0.20</td>
<td>0.45</td>
<td>0.30</td>
<td>0.21</td>
<td>0.16</td>
<td>0.57</td>
<td>0.42</td>
</tr>
<tr>
<td>K percent</td>
<td>2.35</td>
<td>3.06</td>
<td>0.92</td>
<td>0.99</td>
<td>1.90</td>
<td>3.03</td>
<td>1.20</td>
<td>1.20</td>
</tr>
<tr>
<td>Na percent</td>
<td>0.13</td>
<td>0.07</td>
<td>1.42</td>
<td>1.43</td>
<td>0.09</td>
<td>0.06</td>
<td>0.49</td>
<td>0.18</td>
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<tr>
<td>Fe mg/l</td>
<td>2,520</td>
<td>2,836</td>
<td>1,368</td>
<td>1,793</td>
<td>1,820</td>
<td>1,643</td>
<td>60</td>
<td>80</td>
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<tr>
<td>Mn mg/l</td>
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<td>829</td>
<td>1,641</td>
<td>1,658</td>
<td>1,677</td>
<td>1,729</td>
<td>1,080</td>
<td>800</td>
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<tr>
<td>Zn mg/l</td>
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<td>29</td>
<td>72</td>
<td>46</td>
<td>89</td>
<td>119</td>
<td>520</td>
<td>---</td>
</tr>
<tr>
<td>Cu mg/l</td>
<td>19</td>
<td>23</td>
<td>47</td>
<td>34</td>
<td>75</td>
<td>75</td>
<td>143</td>
<td>70</td>
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<td>B mg/l</td>
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<td>65</td>
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<table>
<thead>
<tr>
<th>Analysis</th>
<th>Cladophora</th>
<th>Euglena</th>
<th>Hydrodictyon</th>
<th>Microcystis</th>
<th>Lyngbya</th>
<th>Nitella</th>
<th>Aphaniizomenon</th>
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<tr>
<td>Ash percent</td>
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<td>4.12</td>
<td>17.94</td>
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<td>48.14</td>
<td>39.96</td>
<td>46.46</td>
<td>40.23</td>
<td>38.43</td>
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<tr>
<td>N percent</td>
<td>2.30</td>
<td>5.14</td>
<td>3.87</td>
<td>8.08</td>
<td>5.01</td>
<td>2.70</td>
<td>8.57</td>
</tr>
<tr>
<td>P percent</td>
<td>0.56</td>
<td>0.67</td>
<td>0.24</td>
<td>0.68</td>
<td>0.31</td>
<td>0.23</td>
<td>1.17</td>
</tr>
<tr>
<td>S percent</td>
<td>1.58</td>
<td>1.91</td>
<td>1.41</td>
<td>0.27</td>
<td>0.28</td>
<td>0.34</td>
<td>1.18</td>
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<tr>
<td>Ca percent</td>
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<td>0.05</td>
<td>0.69</td>
<td>0.53</td>
<td>0.45</td>
<td>1.89</td>
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<td>Mg percent</td>
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<td>0.17</td>
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<td>Na percent</td>
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<td>0.04</td>
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<td>Zn mg/l</td>
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<td>B mg/l</td>
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<td>3.6</td>
<td>112</td>
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</table>

1 Taken from Water Quality Criteria (1968), from data supplied by Lawrence (personal communication) to the Department of the Interior.

Also indicated in this table is 1) whether the lake is supersaturated with carbon dioxide and 2) the current lake condition. The data in this table suggest that there is little correlation between free CO₂ availability and degree of eutrophication in these lakes. For the Great Lakes there is almost an inverse relationship between free CO₂ available and degree of eutrophication. Similarly, a study of 12 lakes in Northern Saskatchewan by Rawson (1960) failed to show any correlation between free CO₂ availability and degree of eutrophication. It appears that many oligotrophic lakes are relatively rich in available inorganic carbon and that some factor other than carbon is limiting algal growth in these lakes. Lake Tahoe, on the California-Nevada border, is a good example of this situation. This lake is one of the most beautiful natural lakes in the world. Noted for its exceptional clarity, it is considered to be oligotrophic. It has a total alkalinity of 41 mg/l as CaCO₃ and a surface pH of 7.8 (McGauhey et al., 1968, 1969). Thus, it contains about 1.4 mg/l of free CO₂ in solution and has a total inorganic carbon content of about 10.2 mg/l. If carbon were indeed the growth-limiting factor in Lake Tahoe, and assuming that (1) all of this carbon were available for algal growth, and (2) 50 percent of the dry weight of the algae is carbon, then Lake Tahoe could support a growth of over 20 mg/l of algae. This concentration of biomass would easily be described as a serious nuisance condition. Lake Tahoe, although perhaps experiencing some enrichment, is still one of the lesser productive natural water bodies in the world.
Table 6. Ranges of concentrations of minor chemical constituents in Chlorella pyrenoidosa cells. (After Coree (1970).)

<table>
<thead>
<tr>
<th>Constituent</th>
<th>Concentration Range</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total Ash</td>
<td>1.4 - 20.2</td>
<td>[Scott (1943), Spoehr and Milner (1949), Ketchum and Redfield (1949)]</td>
</tr>
<tr>
<td>P</td>
<td>0.06 - 3.0</td>
<td>[Jewell and McCarty (1968), Scott (1943), Ketchum and Redfield (1949), Knauss and Porter (1954), Foree and McCarty (1968), Borchardt and Agad (1968)]</td>
</tr>
<tr>
<td>Ca</td>
<td>0.0 - 1.6</td>
<td>[Scott (1943), Knauss and Porter (1954)]</td>
</tr>
<tr>
<td>Mg</td>
<td>0.3 - 1.5</td>
<td>[Scott (1943)]</td>
</tr>
<tr>
<td>K</td>
<td>0.04 - 1.4</td>
<td>[Scott (1943)]</td>
</tr>
<tr>
<td>Na</td>
<td>0.07 - 0.7</td>
<td>[Scott (1943)]</td>
</tr>
<tr>
<td>S</td>
<td>0.4 - 0.8</td>
<td>[Knauss and Porter (1954)]</td>
</tr>
<tr>
<td>Fe</td>
<td>0.02 - 3.4</td>
<td>[Knauss and Porter (1954)]</td>
</tr>
<tr>
<td>Mn</td>
<td>0.02 - 2.6</td>
<td>[Knauss and Porter (1954)]</td>
</tr>
<tr>
<td>Sr</td>
<td>0.0004 - 0.05</td>
<td>[Knauss and Porter (1954)]</td>
</tr>
<tr>
<td>Cu</td>
<td>0.0008 - 0.03</td>
<td>[Knauss and Porter (1954)]</td>
</tr>
<tr>
<td>Zn</td>
<td>0.0004 - 0.009</td>
<td>[Knauss and Porter (1954)]</td>
</tr>
</tbody>
</table>

Edmondson (1970) has presented the chemical and biological changes that occurred when secondary effluent was diverted from Lake Washington producing major changes in nutrient input. Approximately three years following the diversions, the lake has improved markedly as evidenced by the reduction in chlorophyll in surface phytoplankton and transparency of the lake. The summer mean of the chlorophyll content of the phytoplankton in the epilimnion was definitely related to the concentration of phosphate in the surface water during the previous winter but not to nitrate, CO₂ or alkalinity. The CO₂ and nitrate concentrations decreased much less than the phosphate after the diversions. The free CO₂ concentrations fluctuated around 75 percent of the values present prior to diversion, and the alkalinity increased by about 20 percent after diversions. Based upon this information, it appears to be unlikely that changes in inorganic or organic carbon concentrations contributed to the improvements in Lake Washington.

Similar results on the benefits of diversion have been reported by Liebmann (1970) and Laurent et al. (1970) for several European lakes.

Kerr et al. (1970) have reported that CO₂ is the limiting nutrient in the natural waters they studied. This conclusion was supported primarily by laboratory data with samples containing surpluses (1-2 mg/l) of phosphate and limited quantities of
Table 7. Alkalinity and pH data for various natural water bodies in North America.

<table>
<thead>
<tr>
<th>Water Body</th>
<th>Total Alkalinity Actual (mg/l as CaCO₃)</th>
<th>pH @20°C</th>
<th>Equilibrium CO₂ @20°C</th>
<th>Free CO₂ @20°C</th>
<th>Total Inorganic Carbon (mg/l)</th>
<th>Condition of Water Body</th>
<th>Report Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Great Lakes</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Erie</td>
<td>97</td>
<td>8.1</td>
<td>8.60</td>
<td>1.64</td>
<td>24.3</td>
<td>Eutrophic</td>
<td></td>
</tr>
<tr>
<td>Ontario</td>
<td>96</td>
<td>8.1</td>
<td>8.60</td>
<td>1.63</td>
<td>25.0</td>
<td>Eutrophic</td>
<td></td>
</tr>
<tr>
<td>Michigan</td>
<td>113</td>
<td>8.0</td>
<td>8.65</td>
<td>2.39</td>
<td>27.1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Huron</td>
<td>82</td>
<td>8.1</td>
<td>8.51</td>
<td>1.39</td>
<td>19.7</td>
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<td></td>
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<tr>
<td>Superior</td>
<td>46</td>
<td>7.4</td>
<td>8.27</td>
<td>3.88</td>
<td>12.0</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lake Tahoe, Calif.</td>
<td>41</td>
<td>7.8</td>
<td>8.24</td>
<td>1.38</td>
<td>10.2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Goose Lake, Wis.</td>
<td>107</td>
<td>7.9</td>
<td>8.63</td>
<td>2.65</td>
<td>26.8</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sylvan Lake, Wis.</td>
<td>110</td>
<td>9.1</td>
<td>8.64</td>
<td>0.17</td>
<td>26.4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Green Lake, Wash.</td>
<td>32.6</td>
<td>7.7</td>
<td>8.17</td>
<td>1.29</td>
<td>8.2</td>
<td></td>
<td></td>
</tr>
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<td></td>
<td></td>
</tr>
<tr>
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<td>7.9</td>
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<td>2.65</td>
<td>26.8</td>
<td></td>
<td></td>
</tr>
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<td>110</td>
<td>9.1</td>
<td>8.64</td>
<td>0.17</td>
<td>26.4</td>
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<tr>
<td>Green Lake, Wash.</td>
<td>32.6</td>
<td>7.7</td>
<td>8.17</td>
<td>1.29</td>
<td>8.2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Canadian Lakes</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cree Lake</td>
<td>48.5</td>
<td>7.2</td>
<td>8.27</td>
<td>6.02</td>
<td>13.4</td>
<td>Oligotrophic</td>
<td></td>
</tr>
<tr>
<td>Larange Lake</td>
<td>122.5</td>
<td>8.2-7.7</td>
<td>8.69</td>
<td>1.63-5.18</td>
<td>29.3-30.5</td>
<td>Mesotrophic</td>
<td></td>
</tr>
<tr>
<td>Waskesiu Lake</td>
<td>154.5</td>
<td>8.2-7.7</td>
<td>8.78</td>
<td>2.06-6.53</td>
<td>37.0-38.5</td>
<td>Eutrophic</td>
<td></td>
</tr>
<tr>
<td>Hunter Bay Lake</td>
<td>97.5</td>
<td>7.9-7.5</td>
<td>8.60</td>
<td>2.59-6.49</td>
<td>24.3-25.2</td>
<td>Oligotrophic</td>
<td></td>
</tr>
<tr>
<td>Big Peter Pond</td>
<td>113.5</td>
<td>8.2-7.2</td>
<td>8.65</td>
<td>1.51-14.2</td>
<td>27.1-31.6</td>
<td>Eutrophic</td>
<td></td>
</tr>
</tbody>
</table>

1. Data from reference cited
2. Calculated from Equation 11
3. Calculated from Equation 13
4. Calculated using Saunders et al. (1962) method based on pH and total alkalinity
5. Based upon Rawson's (1960) assumption that 85 percent of the total dissolved solids was bicarbonate
Based upon the concentrations of phosphates used, it should have been obvious that CO$_2$ would become limiting. This has been demonstrated many times in waste-water stabilization ponds where carbon is known to limit the total production of algae. Also other possible limiting nutrients (for example, iron and trace elements) were not evaluated, nor was the effect of reducing phosphorus concentrations considered. Edmondson's (1970) results on Lake Washington indicate that reductions in phosphate inputs have a dramatic effect on the primary productivity of a natural lake with similar geochemical properties. (See Oglesby, 1968.)

In view of the above studies, it can be concluded that carbon limits productivity only under certain well defined conditions that occur relatively infrequently in nature.

**SUMMARY**

Algae can obtain carbon for the synthesis of new cells from a variety of sources: inorganic (CO$_2$, HCO$_3^-$, CO$_3^{2-}$) and organic (bacterial degradation, direct utilization). The availability of carbon from these sources depends on the chemistry (pH, buffering capacity, activity of the different ions, ionic composition) and the biology of the aquatic systems (organisms present and nutrient composition). These factors may have as much effect on the kinds of algae present, i.e., succession, as on the mass of algae. The role of rates of growth in relation to rates of nutrient utilization and relationships between growth parameters obtained from such rate studies in natural or semi-natural environments may eliminate some of the confusion which exists concerning which nutrient (carbon, nitrogen, phosphorus, etc.) is limiting. The role of CaCO$_3$ in waters saturated with Ca$^{++}$ and CO$_3^{2-}$ ions and those rate-governing factors which might affect the rate of carbon uptake (e.g., temperature, carbonic anhydrase) are still unclear but may not materially affect the mass concentration of natural algal populations.

It is possible that carbon could be growth rate limiting and possibly controlling the algal mass in systems where other nutrients are considerably in excess (relatively short residence time, sewage lagoons; lakes which are already eutrophic; laboratory flask studies with artificial media) or in special situations such as very low alkalinity lakes or extremely hard water lakes. However, in most oligotrophic and mesotrophic waters and in many eutrophic lakes the carbon supply from inorganic sources in the water, from the daily increment from the atmosphere, bacterial degradation of autochthonous and allochthonous materials, would be more than adequate to force some other factor to be limiting (light, nitrogen, phosphorus, etc.). To gain practical perspective on the carbon question, one might ask which factor one would try to control (hence force to be limiting) in order to prevent, reverse, or check the effects of eutrophication. Light, temperature, and carbon would not be among the answers and in terms of practical technology, one would almost have to answer that it would be necessary to control phosphorus.
REFERENCES CITED


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