A STUDY OF ULTRASTRUCTURAL CHANGES IN TOLERANT AND SUSCEPTIBLE LINES OF ALFALFA INDUCED BY STEM NEMATODE

(DITYLENCHUS DIPSACI KÜHN)

by

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A dissertation submitted in partial fulfillment of the requirements for the degree of

DOCTOR OF PHILOSOPHY

in

Plant Science

(Ultrastructural Plant Cytology)

Approved:

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[Signature]
Doris C. N. Chang
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ABSTRACT

A Study of Ultrastructural Changes in Tolerant and Susceptible Lines of Alfalfa Induced by Stem Nematode (Ditylenchus dipsaci Kühn)

by

Doris C. N. Chang, Doctor of Philosophy

Utah State University, 1971

Major Professor: Dr. W. F. Campbell
Department: Plant Science

Fine structural analyses of host tissue (alfalfa, Medicago sativa L.) response to infection by the stem nematode (Ditylenchus dipsaci Kühn) were conducted. Hypocotyl regions were taken on 1, 3 and 7 days after inoculation.

Electron micrographs of infected tissue indicated the types of damage were the same between Lahontan (tolerant line) and Ranger (susceptible line). Only the infection rate (in percent) and degree of damage were different between lines and among the different temperatures (15, 20 and 25 C). The higher the temperature, the more injury resulted. After 3 to 7 days of infection, the symptoms observed were swelling and broken endoplasmic reticulum (ER), distended and broken chloroplasts, loss of nuclear material and bulging and rupturing of nuclear envelopes. Heavily infected cell walls showed more osmiophilic substances on one side. Infected cytoplasm contained more ER (both rough and smooth ER), ribosomes, vesicles and Golgi apparatus, suggesting increased metabolic activities.
Lobing nuclei were observed in all treatments. Lipid content varied with temperature in one-day-old seedlings. At 15 and 25°C, electron dense substances were commonly found along the tonoplast, intercellular spaces and on the cell wall. Also some enlarged ER were noted in the non-infected controls at these temperatures.

From the fine structural studies of host tissue it is not possible at this time to determine the nature of resistance of alfalfa lines to nematode infection. More studies at both the biochemical and electron microscopical levels are needed. Further, studies on the activities of the nematodes at the various temperatures during the infection periods would be primarily important.

(89 pages)
INTRODUCTION

Alfalfa (*Medicago sativa* L.) is by far the most important forage crop in Utah as well as in the United States as a whole. In the state of Utah and the United States, respectively, alfalfa is grown on 450,000 and 29,008,000 acres from which 1,282,000 and 72,476,000 tons of hay are harvested (USDA, 1967). Alfalfa contains about twice as much digestible protein as clover, and about four times as much as timothy-clover hay or corn silage. Moreover, the alfalfa leaf protein may have potential as a source of protein for humans (Pirie, 1966, 1969). This crop is high in mineral content and contains at least ten different vitamins. It is an especially important source of vitamin A. Furthermore, alfalfa encourages the formation of a well-structured and nitrogen enriched soil layer (Martin and Leonard, 1967).

Because alfalfa is a relatively long-lived perennial, once it is established, the farmer or rancher can realize great monetary return for his investment. Many fields of alfalfa, however, are weakened each year by the alfalfa stem nematode (*Ditylenchus dipsaci* Kühn) which is a microscopic, slender roundworm approximately 1/12-inch long and 1/900-inch wide. Conservative estimates place the loss to this organism in Utah alone at approximately $1,500,000 annually (Griffin, personal communication). In the United States, this loss is about $86 million annually (Anonymous, 1971). The loss in terms of dollars of all crops to nematodes in the United States is estimated at $1.6 billion. This value plus the amount spent for nematode control approaches $1.7 billion.
annually. Once the alfalfa plants are weakened by the alfalfa stem nematodes, bacteria and other pest organisms take over the fields.

Because chemical control of the nematodes is not yet feasible (Griffin, 1967; Sherwood et al., 1967), the only practical means of control is through the breeding and developing of resistant varieties (Grundbacher and Stanford, 1962a; Sherwood et al., 1967). However, Grundbacher and Stanford (1962a) reported that resistance to the stem nematode decreased in Lahontan alfalfa (Medicago sativa L.) with increased temperatures. Griffin (1968) further reported that there was no relationship between host response and numbers of nematodes invading susceptible and resistant alfalfa. Infection occurred at temperatures between 5 and 30 °C with maximum infection at 20 °C. Ranger (a susceptible line) was susceptible at temperatures from 10 to 30 °C, but the susceptibility of Lahontan (a tolerant line) increased slightly as temperature increased from 5 to 20 °C and increased sharply at 25 and 30 °C. The hypocotyl, cotyledonary petioles, and to a lesser extent the epicotyl were invaded first. It is assumed that changes occur at the ultrastructural level long before visible symptoms are evident. Definition of these changes may elucidate basic and subtle differences associated with tolerant and susceptible alfalfa lines. This study, at the fine structural level, sought to answer the following questions:

1. What are the differences in fine structure between Lahontan (tolerant) and Ranger (susceptible) alfalfa lines?
2. What changes are induced by stem nematode infection in these lines?
3. What are the effects of number of nematodes in relation to host response?

4. Are subtle, but detectable, differences in pectin content of the middle lamellae of host cell wall tissue evident before and/or after infection?

5. Are there subtle, but detectable, temperature effects between the two alfalfa lines?

6. What effect does temperature have on the expression of nematode induced host responses?

7. What detectable changes occur in Lahontan that may account for its altered tolerance to stem nematode attacks with increased temperature?
REVIEW OF LITERATURE

Host Tissue Responses

Structural studies

It has been established that the first visible symptom of infection by the alfalfa stem nematode *Ditylenchus dipsaci* (Kühn) is the formation of a gall or a swelling of the stem (Grundbacher and Stanford, 1962a; Radewald and Hart, 1965). The hypocotyl, cotyledonary petioles, and to a lesser extent the epicotyl were invaded first (Griffin, 1968). Infected plants were dwarfed and misshapen, and underground storage structures rotted. Cells near the nematode enlarged and separated; cytoplasm withdrew, and cell walls collapsed, forming cavities in tissues. In aseptic infections with one or two nematodes, leaf palisade parenchyma of alfalfa (*Medicago sativa* L.) leaflets contained masses of cells with dense, granular cytoplasm (Krusberg, 1961). The conspicuous galling of young alfalfa stems, caused by cell hypertrophy, was visible within 24 hours after inoculation. As early as 12 hours after inoculation, cavities developed in the cotyledonary cortex, the epidermal cells had enlarged, and cell reaction to dyes had changed. Vascular damage was not conspicuous, but did occur in old infections. Necrosis was not common in the young alfalfa tissues. The outstanding feature of these infections was that only a few nematodes affect many cells.

Histopathological studies of *D. dipsaci* on alfalfa have indicated distinct varietal differences in young seedlings following infection
(Bingefors, 1961, 1962). In tolerant alfalfa, the nematodes penetrated and were found in cavities in the plant (Bingefors, 1961). Cell walls adjacent to such cavities indicated a strong affinity for safranin. The cells in surrounding tissues were intact, and the cellular structure around the cavities showed very little disruption. By contrast, in susceptible alfalfa there was a very pronounced swelling following infection (Bingefors, 1961). The cellular structures were disrupted in the vicinity of the nematodes. Moreover, the middle lamellae of each affected cell appeared to be dissolved, with a gradient of damage extending to the middle lamellae of adjacent and surrounding cells. Further, the cell walls did not exhibit an affinity for the safranin stain. Bingefors (1961, 1962) and Krusberg (1963a) further noted cell enlargement, a disappearance of chloroplasts and an increase in intercellular space in parenchyma tissue following infection. Krusberg (1963a) reported that effects often preceded direct contact of the nematodes with the cells, suggesting that salivary secretions might be diffusing in advance of the nematodes. Many phytoparasitic nematodes, e.g., Rotylenchulus, affect only those cells upon which they feed, or a limited number of cells in the immediate vicinity of the feeding site (Birchfield, 1962). D. dipsaci, however, causes changes in cells at great distances from itself (Dropkin, 1969).

All the above results are based exclusively on observations made with the light microscope and classical histological procedures. A few investigators are beginning to use the electron microscope to study the dynamic aspects of this subject. However, little data (Bird, 1961; Huang and Maggenti, 1969; Chang, Campbell and Griffin, 1970; Paulson and Webster, 1969, 1970) are available on nematode-induced changes in
ultrastructure of the host tissue as shown by electron microscopy. Bird (1961) studied nematode-induced giant cells of tomato root, and found that the giant cells contained many mitochondria, proplastids, Golgi bodies, and a dense endoplasmic reticulum; the cytoplasm resembled that in meristematic cells during interphase, and appeared to be in a metabolically-active condition. Huang and Maggenti (1969) reported that the giant cells became multinucleate as a result of karyokinesis without subsequent cytokinesis. Paulson and Webster (1970) took sequential samples of tomato roots and observed that giant cell development was characterized by changes in the cell walls, cytoplasm, and organelles. Electron dense cytoplasm and the decreasing size of the large central vacuole were first observed. Then some regions of the giant cell walls became excessively thickened, while other parts remained relatively unchanged. In old giant cells, projections of wall material extended far into the cytoplasm. Microtubules occurred adjacent to the walls in all stages of giant cell development. As the giant cell developed, the nuclei became swollen and irregularly shaped; nucleoli became enlarged, and clumps of chromatin accumulated near the distinct nuclear membrane. Plastids did not change significantly, whereas mitochondria appeared to form vesicles, and the number of dictyosomes increased during the later stages of giant cells. In still later stages of giant cell development, smooth and rough endoplasmic reticulum were extensively produced in the form of vesicles and cisternae. Other histopathological studies of host tissues have been conducted either on nematodes other than D. dipsaci or on other host tissues (Bird, 1962; Blake, 1962a, 1962b, 1966; Christie, 1936; Dijkstra, 1957; Endo, 1964, 1965; Hussey and Krusberg,
Biochemical studies


Plant parasitic nematodes have enzymes that degrade modified cellulose. These enzymes are absent or present only in low concentrations in microbivorous forms (Dropkin, 1963; Krusberg, 1960b; Myers, 1965). The alfalfa stem nematodes are thought to secrete pectinases, which dissolve the middle lamellae between cells, and other irritants, which stimulate the formation of the actual galls. It has been demonstrated that the homogenates of *D. dipsaci* contained polygalacturonase (Krusberg, 1964, 1967; Riedel and Mai, 1971) and a pectin-transeliminase (Krusberg, 1964, 1967). Extracts of a population of *D. dipsaci* from alfalfa (Krusberg, 1960b) contained pectinmethylesterase. Goffart and Heiling (1962) claimed a pectinase was secreted into water and oatmeal agar by *D. dipsaci*. Sherwood and Huisin gh (1970) found pectic enzymes in Atlantic alfalfa buds infected by *D. dipsaci*. Actual evidence that pectinases are effective in destruction of the middle lamella, however, is not yet conclusive. Krusberg (1963a) could not macerate alfalfa tissue with extracts of *D. dipsaci* nor could he find histochemical
evidence for pectin removal. Krusberg (1967) concluded that widespread degradation of pectic compounds in the middle lamellae of alfalfa plants by nematode pectinase was not a factor in pathogenesis. Also, these enzymes occurred in phytonematodes which did not cause cell separation in their hosts.

The evidence for nematode-induced changes in plant growth regulatory substances is fairly strong (Cutler and Krusberg, 1968; Viglierchio and Yu, 1965). Yet, it has not been possible to distinguish between the introduction of a growth regulator by the nematode, and the induction of changes in levels or gradients of endogenous growth regulators of the host, or inhibition of enzymes that degrade growth regulators (Dropkin, 1969).

Recent work on decapitated pea epicotyls indicated that IAA played a central role in the control of cellulase synthesis (Fan and Machlachlan, 1967). The authors postulated that IAA depressed that part of the genome which synthesized RNA for cellulase, and therefore acted on transcription. The cellulase presumably moves to the cell walls where it is rapidly inactivated. The enzyme may change wall plasticity and may influence cellulose deposition by providing sites for attachment of new cellulose. Many of the effects of nematodes are parallel to those reported in the work on IAA. Hyperplasia, hypertrophy, and the breakdown of cell walls all developed in decapitated pea epicotyls treated with endogenous IAA. The pattern of cell wall lysis in D. dipsaci infections of stems does not seem to result from passive responses to enzymes emanating from the nematodes, but rather suggests an active host participation in response to some controlling force from the parasite (Dropkin, 1969).
Many other biochemical studies of host tissues infected by nematodes have been done (Acedo and Rohde, 1970; Bird, 1961; Colotelo and Ward, 1961; Johnson and Viglierchio, 1969; Muse et al., 1970; Orion and Minz, 1968; Wilski and Giebel, 1966; Veech and Endo, 1969).

**Mechanism of galling**

The physiology of plant gall or tumor formation has intrigued investigators and has been studied by them for years. Yet no definite pattern has been revealed to explain the observed growth changes.

Sayre (quoted by Mountain, 1960) suggested a mechanism involving IAA for gall induction by root-knot nematodes. After penetration into root tissues, larvae supposedly secreted proteolytic enzymes which hydrolyzed IAA protein complexes releasing IAA to stimulate gall formation. Sayre further suggested that nematode proteases hydrolyzed plant proteins, releasing tryptophan which was transformed by plant enzymes to IAA. Leopold (1960) reported that auxins promoted hydrolysis of plant cell proteins.

Krusberg (1961) found free tryptophan in alfalfa shoot galls caused by *D. dipsaci* but none in healthy tissues and postulated that it might be involved indirectly in gall induction. Cutler and Krusberg (1968) reported a plant-growth regulator with indole-acetic methylester (IAM) characteristics in the extracts of *D. dipsaci*. They estimated it to be present at a rate of 1 μg/100 mg dry wt of nematodes. Viglierchio and Yu (1965) noted auxin inactivators in *Ditylenchus* and observed a reduction of auxins in alfalfa-shoot tips infected with *D. dipsaci*. In the case of nematodes there is no convincing evidence for the origin of auxins, whether in the reacting host cells or in the parasites.
themselves. There have been, however, speculations that the host-cell auxin level is augmented by a nematode injection fluid secreted through the stylet, which releases plant-bound auxin. Also, it has been suggested that the auxin present within nematodes occurs by the intake and accumulation of that normally occurring in the plant cell (Mountain, 1960). Recent experiments incorporating radioactively labelled IAA in culture media, however, have indicated that *D. dipsaci* and *D. trifurcata* were unable to take up significant amounts of the label (Cutler and Krusberg, 1968). It is evident, therefore, that not all auxin-containing nematodes would be able to obtain their free auxins from the host cells.

Krusberg (1963a) stated that mere hydrolyzed products of proteins would not seem sufficient to stimulate gall formation, since all plant nematodes probably secrete proteases into plant cells, but relatively few species stimulate galling.

It is tempting to explain the extensive cell lysis observed around *D. dipsaci* by suggesting that the nematodes emit hydrolytic enzymes into plant tissues. Pectinases are the enzymes most frequently proposed (Goffart and Heiling, 1962; Krusberg, 1964, 1967; Sherwood and Huisman, 1970). No strong consistent evidence, however, has yet been found to support this claim (Krusberg, 1963; Dropkin, 1969).

Huang and Maggenti (1969) noted that giant cell nuclei in the roots of *Vicia faba* infected with *Meloidogyne javanica* were derived from repeated mitoses of the original diploid cells without subsequent cytokinesis. Owens and Novotny (1960) and Littrell (1966) reported that both cell-wall dissolution and mitoses were responsible for giant cell formation.
Nature of resistance

Resistance is a characteristic of the host plant. However, various environmental factors such as temperature, soil type, host nutrition, age of the plant and previous cropping history may alter the expression of resistance (Rohde, 1960).

Temperature effects on the rate of penetration and reproduction of nematode in the host have been found in *Ditylenchus* (Griffin, 1968; Grundbacher and Stanford, 1962a; Hanna and Hawn, 1965), *Meloidogyne* (Tyler, 1933), *Pratylenchus* (Mountain, 1957), and *Trichodorus* (Rohde and Jenkins, 1957).

Several authors have noted that soil type has an effect on pathogenicity of plant-parasitic nematodes. Bessey (1911) transplanted plants infected with root-knot nematodes into pots containing susceptible hosts. In heavy clay soils, susceptible plants were not attacked.

Potassium has been found to be associated with the resistance to root-knot nematodes (Bessey, 1911; Otiefa, 1953; Crittenden, 1954). In several plant diseases, in which enzymes have been implicated, increased calcium (Ca++) content of plant tissue was correlated with increased resistance (Alghisi and Ferraza, 1967; Bateman and Lumsden, 1965; Bateman and Millar, 1966; Corden, 1965; Thomas, 1966). Probably Ca++ forms enzyme-resistant complexes with polygalacturonates in the middle lamellae.

Older plants seem to be more resistant to nematode attacks than young seedlings, such as tea plants to *Meloidogyne* infection (Loos, 1953).
Previous cropping may be a factor in selecting out more pathogenic strains of nematode species. In Australia, tomato varieties resistant to *M. javanica* break down after five years of continuous cropping in the same fields (Giles and Hutton, 1958).

According to Rohde (1960), the mechanisms of resistance to nematodes may be summarized as follows:

1. Root secretions as a mechanism of resistance (Winslow, 1955)
2. Resistance to penetration (Sasser, 1954)
3. Internal factors involved in resistance (Seinhorst, 1956a; Bingefors, 1957; Dijkstra, 1957)
4. Resistance related to a change in the sex ratio (Cobb, Steiner and Christie, 1927; Williams, 1956)
5. Production of resistant factors in plants (Riggs and Winstead, 1958; Swink and Finkner, 1956)

Studying the resistance in clover and alfalfa to the stem nematode, *Ditylenchus dipsaci*, Bingefors (1957) suggested the possibility that there were slight differences in the chemical composition of the middle lamellae in resistant red clover varieties, and a few major genes were mainly responsible for the resistance.

The resistance of Lahontan to *D. dipsaci* was expressed as reduced swelling of the host tissue and reproduction of the nematode, but not as reduced penetration. This has been agreed upon among several investigators such as Bingefors (1957), Griffin (1968), Sherwood and Huisingh (1970), and Wynne and Busbice (1968).

Sherwood and Huisingh (1970) studied calcium nutrition and resistance of alfalfa to *D. dipsaci* and found that divalent/monovalent cation content of tissues modifies the expression of resistance by
alfalfa buds. Therefore, they suggested that the differences in resistance of varieties might be related to differences in their ability to accumulate cations. The basic nature of resistance, however, remains unknown.

**Alfalfa Stem Nematode—Ditylenchus dipsaci** Kühn

The first record of *Ditylenchus dipsaci* is that of Schwertz in 1855 (Thorne, 1961), who recorded certain diseased conditions of rye, oats, clover, and other crops but did not observe the nematodes. Kühn (1857) discovered *D. dipsaci* (*Anguilla dipsaci*) in teasel, *Dipsacus fullonum* L., and recognized those in rye as being similar in form. In 1881 Kühn first recognized alfalfa as a host.

The bulb and stem nematode, *D. dipsaci*, is a complex group of populations that attack several hundred different plants varying from fuller's teasel, alfalfa, clover, and weeds to the bulbs of tulips, narcissi, hyacinths, onions, and garlic (Thorne, 1961). This nematode is very widely distributed, but it is more serious as a pest in cooler and moister climates (Winslow, 1960).

The genus *Ditylenchus* was established by Filipjev in 1934. With the erection of this genus, a large number of species previously included in *Tylenchus* or *Anguillulina* were transferred to *Ditylenchus*. *Ditylenchus dipsaci* (Kühn, 1857) was designated as the type species by Filipjev. At that time the genus comprised 14 species. In 1949, Thorne amended the generic diagnosis. At present the genus includes more than 30 species.

One of the outstanding features of *D. dipsaci* is the remarkable race specialization, there being a dozen or more known races or biotypes
morphologically indistinguishable but each having definite host preferences (Winslow, 1960). Another feature is that anabiosis developed to a high degree in D. dipsaci. Fielding (1951) reported that these nematodes were 100 percent alive after 20 years dormancy in Hypochoeris radicata. The anabiosis of D. dipsaci is probably confined to the fourth stage larvae or preadult. We have no exact information on the physiology of dormant nematodes but it is generally presumed that metabolism proceeds very slowly, the fat stored in the body being gradually used up (Winslow, 1960).

The nematode is disseminated by unclean seeds, bulbs, cuttings, machinery, and irrigation water (Thorne, 1961).

Control of the nematode can be done by: (a) crop rotation (sugar beets, grain, corn, potatoes, tomatoes and other common crops are suitable) and (b) resistant varieties (in alfalfa, Resistador, Lahontan, Talent and Kayseri).

Structural studies

Filipjév (1934) and Thorne (1949) were prominent early researchers who studied this genus of nematode. The description made by Thorne in 1949 is still the most widely used reference for this nematode.

Recently Evans and Fisher (1970) studied the excretory systems of three Ditylenchus spp. by light microscopy. Yuen (1967, 1968) is the only one who has studied D. dipsaci by electron microscopy. Other works that have been conducted on the ultrastructure of nematodes are on Rhabdites (Beams and Sekhon, 1967), Ascaris (Bird and Deutsch, 1957), Meloidogyne (Bird and Rogers, 1965), Trichodorus (Hirumi et al.,
Biochemical studies

The majority of biochemical work concerns hydrolytic enzymes present in nematode homogenates. Myuge (1957), Tracey (1958), Krusberg (1960b, 1964), Muse et al. (1970), Riedel and Mai (1971) found that the hydrolytic enzymes present in the D. dipsaci homogenates were amylase, cellulase, chitinase, glucosidases, pectinases and protease. Krusberg (1960b), Hussey and Krusberg (1971) and Dickson, Huisingh and Sasser (1971) detected other enzymes such as hexokinase, glucosephosphate isomerase, phosphoglyceromutase, enolase, lactic dehydrogenase, malate dehydrogenase, acid and alkaline phosphatase, esterase, isocitric dehydrogenase, oxalosuccinic decarboxylase and fumarase in D. dipsaci.

Eriksson and Granberg (1969) have attempted to study the D. dipsaci races by using electrophoresis in acrylamide gel. Siddigui and Viglierchio (1970) have studied the effects of gamma irradiation on the motility, infection, reproduction and morphology of D. dipsaci.
MATERIALS AND METHODS

Preparation of Nematodes

The method for nematode sample preparation described by Thorne (1961, p. 41-52) was followed:

Nematodes in alfalfa stems were collected from a field located in Cache Valley, Utah, and were stored at 5 C overnight. Leaves were removed, and stems were cut into 1/8 or 1/4 inch long sections. The pieces of stems were placed in distilled water in a Baermann-funnel on which a David Bradley milk filter disc 6 inches in diameter was placed on a 4 mesh wire. The nematodes moved from the alfalfa tissue into the water, through the filter and settled in the bottom stem of the funnel. After an hour, the distilled water in the bottom of the Baermann-funnel, which contained the nematodes, was drained off. This suspension was stored at 4 C for 3 to 4 hours to precipitate the nematodes. The number of nematodes was then determined microscopically by placing a drop of suspension on a watch glass with a sub-divided bottom. Dilutions were made until each drop contained about 10 nematodes. When stored at 5 C, the nematodes remained viable for at least 7 days.

Preparation of Alfalfa Seedlings

Seeds of alfalfa (Medicago sativa L.) cvs Ranger (a susceptible line) and Lahontan (a tolerant line) were scarified with 100 grit silicon carbide water sandpaper and germinated for 36 hours under
room temperature in petri dishes lined with filter paper and moistened with distilled water.

**Inoculation and Plantings**

The germinated alfalfa seeds were placed in a 10 x 14 inch metal container (100 seeds/container) of sterilized Provo sand, with a soil moisture of approximately 100% field capacity. One cc of nematode suspension containing either 20 or 100 nematodes was pipetted directly over the germinating seed. Seeds were covered with 0.25 inches of soil. The containers were placed in growth chambers at temperatures of 15, 20 and 25 C. Sixteen hours of light at about 3,000 ft-c were maintained.

**Fixation**

**General**

Samples were harvested at 1-, 3- and 7-day intervals. Hypocotyl sections (1 mm) of infected alfalfa seedlings showing visible symptoms and similar regions in non-infected controls were harvested and fixed by two methods:

1. Karnovsky's fixative in cacodylate buffer pH 7.2 (1965) for 4 hours at room temperature, rinsed with cacodylate buffer and kept overnight in a refrigerator. Fixed secondarily with 2% osmium tetroxide in cacodylate buffer for 1 hour and washed with two changes of cacodylate buffer at 15 minutes each.

2. One part of 5% glutaraldehyde, 1 part of 5% osmium tetroxide together with 2 parts of 0.2 M phosphate buffer, pH 7.2 (Trump and Bulger, 1966) for 1 hour on ice, and washed with phosphate buffer.
All samples were dehydrated with an ethanol-propylene oxide series for 15 minutes each and embedded in Epon 812 (Pease, 1964) with six parts of solution A and four parts of solution B. Polymerization was carried out by placing the specimens in Beem capsules and incubating at 45 and 60 °C for 24 hours each.

**Staining pectin of the cell wall**

Hypocotyl sections of alfalfa seedlings were harvested and fixed in Karnovsky's fixative for 4 hours. Sections were fixed secondarily with 2% osmium tetraoxide for 1 hour, rinsed with cacodylate buffer, and dehydrated in 35% ethanol as usual. The method for staining pectins for electron microscopy was followed (Albersheim and Killias, 1963). The samples were washed twice in 60% ethanol for 10 minutes each, and then put into 60% ethanol alkaline hydroxylamine solution for 1 hour at 4 °C. They were then washed in 0.1 M HCl for 15 minutes, put into 2% FeCl₃ for 1 hour, then dehydrated with acetone-propylene oxide series and embedded in Epon 812.

**Sectioning, Staining and Examining**

1 μ sections for light microscopy

After sufficient polymerization and cooling of the Epon, the blocks were properly trimmed to the tissue. For orientation purposes 1 μ-thick sections were cut with a glass knife on a Sorvall MT-2 ultramicrotome.

Slides previously cleaned and stored in 60-95% ethanol with a few drops of 0.1 N HCl were air-dried and a drop of distilled water was added. A camel's hair brush was wetted and used to take the section
from the water trough to the slide. The slide was placed on a hot plate (40 C). After 15 minutes, Richardson's stain (Richardson, Jarett and Finke, 1960) was applied on the section and the slide was covered with a petri dish on the hot plate. After 1 or 2 minutes, excess stain was rinsed from the slide with a stream of distilled water. The slide was dried again on the hot plate and then examined with a light microscope.

**Thin sections for electron microscopy**

Blocks were trimmed to approximately 0.5 to 1.0 mm on a side. Sections were cut with a glass or diamond knife on a Sorvall MT-2 ultramicrotome. Only light gold or silver sections were placed on acetone-cleaned 3 mm 200 mesh copper grids. Sections were stained with saturated aqueous uranyl acetate at 60 C for 12 minutes (Watson, 1958) followed by lead citrate at room temperature for 3-5 minutes (Reynolds, 1963). Sections were examined with a Zeiss EM-9A and significant observations recorded photographically.
RESULTS AND DISCUSSION

Morphology

Seedling growth was directly influenced by temperature. No visible symptoms of nematode infection, however, were observed at any temperature in either Ranger or Lahontan 1 day after inoculation. Three days after planting, swollen hypocotyls could be seen at 20°C and 25°C in both cultivars. Visible symptoms were observed in seven-day-old seedlings at all temperatures (Figures 1, 2, and 3). The degree of swelling, however, was higher in Ranger (susceptible line) than in Lahontan (tolerant line) at all ages. These observations confirm those of Griffin (1968).

Light Microscopy

Figure 4 shows the cross section of control alfalfa hypocotyl. Figure 5 indicates that the nematode infection is only in the cortical region. Light microscopy of the infected areas in both Ranger and Lahontan substantiated the histopathological studies conducted earlier by Bingefors (1961, 1962) and Krusberg (1961).

Electron Microscopy

Nematode penetration of both Ranger and Lahontan occurs at temperatures between 5 and 30°C with maximum infection at 20°C (Griffin, 1968). Thus, 20°C was the temperature chosen for the most detailed study. The infection rate was lower at 15°C than at 20°C in Ranger.
Also the susceptibility of Lahontan increased sharply at 25 C. Thus, they were chosen as the other two temperatures for the purpose of comparing the temperature effects.

20 C

The swelling of hypocotyls in susceptible Ranger was usually visible after 3 to 7 days of inoculation. This is somewhat longer than the 24 hours reported by Krusberg (1961), and may indicate a differential cultivar response.

Control plants in both Ranger and Lahontan exhibited normal cell wall and cytoplasm. The cytoplasm contained a nucleus, mitochondria, ER, ribosomes, Golgi apparatuses, and vacuoles (Figures 6 and 22). Plasmadesmata were common on the cell walls (Figures 9 and 22). Control plants also contained normally developing chloroplasts bearing stroma, thylakoid units of the grana, and starch grains. Those chloroplasts devoid of starch grains were lens-shaped (Figures 9, 12 and 22), and those containing starch grains usually became oval or nearly round in shape (Figures 9, 11, 23 and 24).

Crystalline and paracrystalline arrays of electron dense granules were observed in the chloroplasts of control and infected plants at all ages and all treatments in both Ranger and Lahontan. This complex has been referred to as phytoferritin (Hyde et al., 1963; Jacobson, Swift and Bogorad, 1963; Robards and Robinson, 1968; Robards and Hympherson, 1967). These workers suggested that the phytoferritin was an iron-protein complex that allowed the plant to store iron in a non-toxic form. This theory is in accord with the phytoferritin that is most commonly found in differentiating plastids and often in meristematic cells and may provide iron for further development in
young cells, including the elaboration of photosynthetic material (Hyde et al., 1963; Robards and Hympherson, 1967). Robards and Robinson (1968) found that phytoferritin was completely unaffected by the treatments of deoxyribonuclease or ribonuclease, thus excluding the possibility that the inclusions were viral particles.

On the first day after inoculation with *D. dipsaci*, infected Ranger and Lahontan exhibited more lipid bodies per cell than did the controls (Figures 10, 11, 23 and 24). Sheetz and Crittenden (1966) histochemically demonstrated a greater abundance of lipids in the giant cells of rootknot-susceptible soybean than in rootknot-resistant plants. The number of lipid bodies, however, was less in Lahontan than in Ranger. No other changes were noted in the fine structure at this age.

Three days after planting, however, the chloroplasts in infected cells of Ranger were swollen and the outer chloroplast membrane was ruptured. Swelling of the thylakoid units was also noted (Figure 13). Moreover, in a few cases, the nucleus was swollen and the outer nuclear membrane was ruptured. Leakage of nuclear content was also observed (Figure 15). The permeability of the membranes had obviously been altered by the infection of nematodes, probably through enzymatic reactions. Moreover, compared to the controls, the cytoplasm of the infected cells became more dense with an abundance of ribosomes and numerous cisternae of smooth ER (Figures 16 and 17) and may indicate an injury response. Further, one side of the cell wall became more osmiophilic (Figure 17), suggesting chemical changes of the cell wall have been induced by nematode infection. In Ranger, the host tissue at the infection site, i.e., next to the nematode, showed a complete
loss of cytoplasm and an osmiophilic cell wall at the side adjacent to
the nematode (Figure 18). These suggest that at the infection site,
the injury responses are caused by mechanical suction and the enzyme
secretion of the nematode.

With few exceptions, three days after planting Lahontan contained
normally developing chloroplast, cytoplasm and nucleus even in the
infected cells (Figures 25, 26 and 27). Lahontan alfalfa has been
regarded as tolerant to nematode infection; therefore, less damage
responses were expected. In a very few cases, however, when the cell
was heavily infected, the infection responses at the fine structural
level were the same as those in Ranger, i.e., ER became swollen in
sausage-like chains, numerous vesicles formed, chloroplasts swelled,
and the outer membranes of the chloroplasts ruptured. Swelling of
thylakoid units of the grana was also noted (Figure 28). One impor-
tant feature about Lahontan, even at the infection site, was that the
cell wall remained unchanged (Figure 28).

In Ranger, seven days after infection, partial to complete disrup-
tion of chloroplasts was evident (Figure 19). Abundant "vesicles,"
which may have formed from ER, were found in the cytoplasm and the cell
wall broke down causing a mixing of cytoplasm of adjacent cells
(Figure 20). In Lahontan, seven days after infection, most chloroplasts
were normal (Figures 30 and 31). Heavily infected areas, however,
showed symptoms, such as leaking of nuclear content, swollen ER (Figure
32), and broken chloroplasts (Figures 32 and 33), as were observed in
Ranger. But most cell walls still remained normal (Figure 32).

At all temperatures and all treatments, especially in one-day-old
seedlings, large amounts of electron dense masses were present in both
cytoplasm and vacuoles (Figures 34 and 35). They varied in size and shape and have been identified as protein bodies (Bagley et al., 1963; Horner and Arnott, 1965). As the seed germinated, the protein bodies swelled and developed cavities within. Later these swollen bodies broke up into fragments which were digested and disappeared, and in seven-day-old seedlings very few protein bodies were visible. Lipid and protein bodies are the two major reserve foods stored in alfalfa seeds. Both were digested and disappeared gradually with increased age of the seedlings.

Myelin figures were found in some treatments (Figure 35) and apparently have no relation with the temperature treatment and nematode infection. Curgy (1968) suggested that myelin figures were seen only after aldehyde fixation or aldehyde/OsO₄ double fixation and indicated that fixation with OsO₄, with KMnO₄ or with a combined glutaraldehyde-OsO₄ mixture did not reveal such structures.

At 20°C, some chloroplasts were observed containing numerous small vesicles along the periphery of the chloroplasts in control and treated of both cultivars (Figure 35). Laetsch (1968) observed tubules of the peripheral reticulum on the chloroplasts in dicotyledons which possess the C₄-dicarboxylic acid pathway of photosynthetic CO₂ fixation. According to J. H. Hillard (personal communication, 1971), the presence (C₄) or absence (C₃) of parenchymatous bundle sheath on a leaf cross-section is the most reliable structural criterion for C₃ and C₄ plants. On this basis, then, alfalfa should be classified as a C₃ plant. However, tubules of peripheral reticulum were observed in the alfalfa chloroplasts. Thus, peripheral reticulum or vesicles may have no relation with type of photosynthetic pathways.
All of the above treated plants were inoculated with 20 nematodes per seedling. For a comparison of the host response and numbers of nematode each seedling was inoculated with 100 nematodes. Fine structural studies agreed with the statement made by Griffin (1968), i.e., the response of host tissue inoculated with 100 nematodes per plant did not show any more severity of infection than those inoculated with only 20 nematodes (Figures 37, 38 and 39). Griffin (1968) speculated that this response is due to the ability of *D. dipsaci* to invade resistant as well as susceptible alfalfa plants. He also found that the nematode population gradually declined in tolerant alfalfa, which agrees with the findings of Bingefors (1951) and Grundbacher and Stanford (1962a). In Ranger, the cytoplasm at the infected site was plasmolyzed (Figure 37). In Lahontan, slightly infected cells showed slight swelling of the ER only. In heavily infected cells, osmiophilic cell walls, broken and swollen ER, and numerous vesicles were observed (Figure 39).

The comparison of pectin content in the cell wall is shown in Figures 40 to 47. No obvious difference was noted in the pectin content of cell walls in treated or non-treated control plants at all ages in either Ranger or Lahontan. This finding agrees with the statement made by Krusberg (1963b, 1967) that there is no histochemical evidence from light microscopy for pectin removal, and the degradation of pectic compounds in the middle lamellae of alfalfa by nematode pectinase was not a factor in pathogenesis.

In summary, at 20°C, except for the lower degree of responses in Lahontan than in Ranger, almost no difference in structure has been observed between Lahontan and Ranger. The organelles showing damages
after infections were ER, chloroplasts, and nucleus. The lobing of nuclei was often observed in both treated and controls; therefore, it is not due to nematode infection and fits the interpretation made by Õpik (1966) that nuclear lobing is an indication of intense nuclear activity. Since lipid content is also found to be associated with nematode resistance in soybean (Sheetz and Crittenden, 1966), more investigations on lipids should be done. Studies on protein bodies at the same time also should not be neglected, for both protein and lipid are important components of the membranes (Davson and Danielli, 1952; Robertson, 1959). In light microscopic studies, Bingefors (1961, 1962) stated that the middle lamellae of each affected cell appeared to be dissolved. These fine structural studies, however, show that up to seven days after infection, the middle lamellae were not dissolved, and one side of the cell wall becoming more osmiophilic was the only response that was observed (Figures 13, 17, 18 and 39). Observations longer than seven days after inoculation should be carried out. In Lahontan, even in the infection sites, most cell walls remain unchanged. It has been speculated by Bingefors (1961, 1962) and Seinhorst (1956b) that differences in the chemical composition of the middle lamellae might account for resistance. Thus, these differences should be investigated in the future.

15°C

The basic fine structure of alfalfa hypocotyl tissue in plants grown at 15°C was similar to those grown at 20°C. The differences noted, however, were as follows. The lipid content in one-day-old non-infected plants was higher than in infected plants of both Ranger and Lahontan. This was a reverse from that observed at 20°C, and may
represent a temperature effect. Chemical analysis should further elucidate this point. Also, electron dense substances were often found in a somewhat continuous manner along the tonoplast (Figure 48), or in a semi-continuous manner in intercellular spaces (Figure 49). Further, some of the electron dense substances became separate spots distributed along the cell wall (Figure 50). At 20 °C, these electron dense substances were only observed in a very few cases, but in 15 °C they occurred rather frequently. Further investigation into their chemical nature and function would be most interesting.

The degree of damage in both Ranger and Lahontan to stem nematode infection at 15 °C was less than that observed at 20 °C. Most chloroplasts and nuclei were normal. Chloroplasts were, however, occasionally observed with peripheral vesicles formed from invagination of the inner membrane (Figure 51). This agrees with the chloroplast structure observed by Laetsch (1968) and Klein and Pollock (1968). Swollen ER were observed in both Ranger and Lahontan at all ages (Figures 53 and 62). Since there is no osmometer on campus, whether the swollen ER is due to unfavorable osmolarity of the fixating solution or due to the cool temperature effect is not distinguishable. It is entirely possible that any abnormal environmental condition (e.g. 15 °C is somewhat cold for the growth of alfalfa seedlings) is able to cause the swelling of ER.

Generally, none of the nematode infected plants showed any visible symptoms in one-day-old seedlings at 15 °C. Light and electron micrographs likewise showed normal cell structure.

In three-day-old seedlings, most chloroplasts near the infection sites were free of damage (Figures 55, 56, 57 and 58). In both Ranger
and Lahontan, cell walls usually were normal (Figures 50, 55 and 57). Endoplasmic reticulum in the infected areas were slightly swollen (Figures 56 and 58), and some were broken into short pieces (Figure 56). Numerous vesicles were also present in the cytoplasm (Figure 56). Chloroplasts contained peripheral vesicles and phytoferritin complexes in both infected and non-infected plants of all ages (Figures 56, 57, 58 and 59), indicating that the nematode infection has no effect on the peripheral vesicles and phytoferritin of chloroplasts. Lobed nuclei were observed in seedlings of all treatments (Figure 57). At the infection sites, the chloroplasts showed the same responses in both cultivars as those grown at 20°C (Figures 13 and 59). The nuclei in cells at the infection sites, however, exhibited less damage than those from plants grown at 20°C (Figures 15 and 59). Endoplasmic reticulum in this area exhibited exactly the same symptoms as those observed at 20°C (Figures 28 and 59).

Seven days after inoculation, the infected nucleus contained less nuclear material than the control (Figures 60 and 61). Chloroplasts and ER showed similar symptoms previously observed at 20°C. Slight swelling of ER was observed in control plants at this temperature (Figure 62), indicating possible low temperature effect. The cytoplasm of infected plants usually contained more actively secreting Golgi apparatus and vesicles (Figure 63). Since infected plants showed visible galling, it is logical to find more Golgi apparatus and vesicles in infected cytoplasm because Golgi vesicles are thought to be participating in cell wall formation (Larson, 1965; Rosen et al., 1964).
In Ranger, the severity of nematode infection at 15 C was slightly lower than that at 20 C, while Lahontan was much lower than Ranger (infection rate was 20% and 92%, respectively).

At 15 C, fine structural studies showed no qualitative difference between Lahontan and Ranger as compared to those at 20 C. Lipid content was the only quantitative difference found between 15 and 20 C. Quantitative histochemical or chemical analysis would be more reliable for the measurement of lipid quantity than electron micrographs.

Since few cell walls showed any change upon nematode infection at this temperature, it, therefore, emphasizes the need for determining the differences of chemical composition in cell walls of Lahontan and Ranger, and how this relates to different temperatures of the host tissues.

When the cells were infected, however, the type of damage was the same in both lines and not much different from those observed at 20 C, indicating cell walls may play a very important role in the tolerance of nematode infection in alfalfa.

25 C

At 25 C, the tolerance of Lahontan to nematode infection was much lower than at 20 C, the infection rate being 70% and 20%, respectively. Ranger showed nearly 100% infection (Figure 2) at this temperature. This agrees with the work of Griffin (1968). The degree of damage was slightly lower in Lahontan than in Ranger. With few exceptions, most of the damage responses were similar between both lines. One striking feature of the high temperature effect was the exhibition of partial to an all-round heavily swollen nuclear envelope in one-day-old
nematode infected Ranger seedlings (Figures 65 and 66). This suggests that secretion is emitted by the invading nematodes. Together with the cellular symptoms observed in the areas far removed from the invading nematodes, it presents strong evidence that the damages are caused by the salivary secretions diffusing in advance of the nematodes (Krusberg, 1963a). Numerous plasmodesmata were present (Figures 66 and 67). These symptoms were not observed in infected Lahontan plants.

In one-day-old Ranger seedlings, swollen ER in sausage-like chains were observed in many cells of the treated samples and some cells of non-infected control (Figures 68 and 69), suggesting a possible high temperature effect. Configurations of the swollen ER were similar to those of nuclear envelopes (Figures 65, 66 and 69), indicating a close relation between these two organelles. Separate and continuous electron dense substances similar to those observed at 15°C were also present quite often on the cell walls and tonoplasts in all treatments (Figures 65, 66, 68 and 70). Their function and chemical nature remain unidentified at the present time. Zee and O'Brien (1970) found similar electron-dense adcrusting substance deposited in the regions of the wall that lay between the primary pit fields in cells of wheat. Whether or not only special areas of the cell have these substances is not known.

At 25°C, more Golgi apparatuses and Golgi vesicles were noted in all treatments (Figure 71). Morphologically, the hypocotyls of 25°C treated seedlings were much bigger than the controls and those treated at 15 or 20°C (Figures 2 and 3). Thus, it is logical to find more Golgi apparatuses, Golgi vesicles and more cell wall surfaces in the differentiating young seedlings. In one-day-old Lahontan seedlings,
swollen ER (Figure 76) was the only symptom that had been detected. Lipid content was higher in Lahontan than Ranger in one-day-old samples. At three temperatures, three different patterns of lipid contents have been observed. Whether these are due to the true temperature effect remains to be determined.

Three days after planting, infected tissues in both Ranger and Lahontan exhibited the same responses as those at 20 C, such as swollen chloroplasts (Figure 72). At this age, the appearance of the cell wall showed the same characteristics as those at other temperatures. No swollen nuclear envelopes were observed at this age. Whether or not the swollen nuclear envelopes recover from the damage or break into pieces such as swollen ER is not known.

Seven days after inoculation, round and partially broken or completely disrupted chloroplasts and swollen ER were noted (Figures 74, 75 and 76). Mitochondria were abnormal in infected Ranger (Figure 74). Vascular bundle cells were not affected by the nematode infection in any treatment. The size of these cells was much smaller than the cortical cells (Figure 78). They also contained nucleus, mitochondria, chloroplasts, ribosomes, ER, cell walls, Golgi apparatuses and vacuoles.
CONCLUSION

In conclusion, at a given temperature, no qualitative differences in the fine structure of Lahontan and Ranger have been observed. Quantitatively, the amount of lipid bodies, ER, ribosomes, Golgi apparatuses and Golgi vesicles showed differences between treated and control within and between temperatures. At all temperatures, ER, chloroplasts and nuclei were the organelles that revealed recognizable symptoms after nematode infection at 3 and 7 days. Some of the cell walls in the heavily infected areas also became moreosmiophilic.

Except for infection rates (in percent) and the varying degree of damage at 15, 20 and 25 C, the heavily infected tissues showed the same types of damages.

This fine structural study does not provide a satisfactory explanation as to why Lahontan is tolerant and Ranger is susceptible or why the infection rates change at different temperatures. It is possible that the changes in infection rates are due to changes in nematode invading abilities at different temperatures.

For further investigations, many approaches can be applied to evaluate the tolerance of plants to stem nematode attacks, such as studying the changes in chemical composition of cell wall with various temperatures, analyzing the lipid and protein content of host tissues, and—most important of all—examining the nematode activities and amount of enzyme secretion under different environmental conditions. Longer infection times (over seven days) also need to be conducted.
Explanation of figures

Abbreviations are as follows:

- C: Chloroplast
- CW: Cell wall
- CY: Cytoplasm
- EDS: Electron dense substance
- ER: (Rough) Endoplasmic reticulum
- G: Grana
- GA: Golgi apparatus
- GV: Golgi vesicle
- IS: Intercellular space
- L: Lipid
- M: Mitochondrion
- MF: Myelin figure
- N: Nucleus
- Ne: Nematode
- NE: Nuclear envelope
- Nu: Nucleolus
- Ph: Phytoferritin
- Pl: Plasmadesmima
- Pr: Protein body
- PV: Periferal vesicle
- R: Ribosomes
- S: Starch
- SER: Smooth endoplasmic reticulum
- T: Tonoplast
- V: Vesicle
- Va: Vacuole

Unless specified, all the treated samples were inoculated with 20 nematodes per plant.
Figure 1. Morphology of seven-day-old control alfalfa seedlings.

Figure 2. Morphology of seven-day-old nematode infected alfalfa seedlings. Note the swollen hypocotyl region.
Figure 3. Morphology of seven-day-old alfalfa seedlings--(a) control and (b) infected plant. Note the swollen hypocotyl of the infected plant.

Figure 4. Cross section of hypocotyl region of control alfalfa seedling (1 μm).

Figure 5. Cross section of hypocotyl region of nematode infected alfalfa seedling (1 μm). Note the infection is only in the cortical region.
Figure 6. One-day-old Ranger alfalfa control plant, bearing normally developed cell wall, mitochondria, Golgi apparatus, endoplasmic reticulum, ribosomes, nucleus, vacuoles and chloroplasts. X 24,000

Figure 7. One-day-old nematode infected Ranger alfalfa plant. X 16,000

Figure 8. Phytoferritin in alfalfa chloroplast. X 52,400
Figure 9. Starch grains and chloroplasts in one-day-old alfalfa control plant. X 24,000

Figure 10. Lipid content in one-day-old Ranger alfalfa control plant. X 9200

Figure 11. Lipid content in one-day-old Ranger alfalfa treated plant. X 9200

Figure 12. Typical shape of normal chloroplast devoid of starch grain. X 24,000
Figure 13. Three-day-old Ranger alfalfa infected plant. Note the osmiophilic staining of the side of the cell wall, the swollen chloroplasts, the rupture of the outer chloroplast membrane and the swollen thylakoid units (arrow). X 9200

Figure 14. Three-day-old Ranger alfalfa control plant with normal chloroplast, mitochondria, and nucleus. Note the lobing nucleus which is normal in alfalfa hypocotyl. X 13,800

Figure 15. Three-day-old Ranger alfalfa infected by nematode showing swelling and lobing nucleus, rupture of outer nuclear membrane (arrows), and the leakage of nuclear content. X 13,800
Figure 16. Three-day-old Ranger alfalfa control plant with normal cytoplasm. X 9200

Figure 17. Three-day-old Ranger alfalfa infected plant with dense cytoplasm and an abundance of ribosomes and numerous cisternae of smooth endoplasmic reticulum. X 9200

Figure 18. Three-day-old Ranger alfalfa infected tissue. Note the response of host tissue beside the nematode. X 5400
Figure 19. Seven-day-old Ranger alfalfa infected plant showing the rupture of chloroplast. X 9200

Figure 20. Seven-day-old Ranger alfalfa infected plant showing the broken cell wall and the mixing of the cytoplasm which contained numerous vesicles of two cells. X 16,000

Figure 21. Seven-day-old Ranger alfalfa control plant with normal nucleus, mitochondria, cell walls, cytoplasm and chloroplasts. X 16,000

Figure 22. One-day-old Lahontan alfalfa control plant with normal cell content. X 16,000

Figure 23. Lipid content of one-day-old Lahontan alfalfa control plant. X 16,000

Figure 24. Lipid content of one-day-old Lahontan alfalfa infected plant. X 16,000
Figure 25. Three-day-old Lahontan alfalfa infected plant in most cases exhibited normally developed chloroplast. X 16,000

Figure 26. Three-day-old Lahontan alfalfa infected plant with normally developed nucleus. X 16,000

Figure 27. Three-day-old Lahontan alfalfa infected plant with normal cytoplasm. X 9200

Figure 28. Three-day-old Lahontan alfalfa plant with heavily infected cytoplasm. Note the abnormal chloroplast, the sausage-like ER and the vesicles. X 9200

Figure 29. Three-day-old Lahontan alfalfa control plant with normal cytoplasm. X 9200
Figure 30. Seven-day-old Lahontan alfalfa infected plant. X 21,000

Figure 31. Seven-day-old Lahontan alfalfa infected plant. X 21,000

Figure 32. Seven-day-old heavily infected alfalfa plant (Lahontan) showing the abnormal nucleus and chloroplast. X 1400

Figure 33. Seven-day-old heavily infected alfalfa plant (Lahontan) showing the rupture of chloroplast membranes. X 9200
Figure 34. One-day-old alfalfa plants in all treatments contained numerous protein bodies of various sizes and shapes. X 9200

Figure 35. Myelin figure in alfalfa plant. X 16,000

Figure 36. Chloroplasts with peripheral vesicles at 20 C. X 16,000

Figure 37. Seven-day-old Ranger alfalfa inoculated with 100 nematode per plant, showing the plasmalyzed cytoplasm. X 16,000

Figure 38. Seven-day-old Lahontan alfalfa inoculated with 100 nematode per plant, showing the slightly swollen ER. X 16,000

Figure 39. Seven-day-old Lahontan alfalfa inoculated with 100 nematode per plant, showing the osmiophilic cell wall and abnormal cytoplasm. X 16,000
Figure 40. Pectin in the cell wall of one-day-old Ranger alfalfa control plant. X 16,000

Figure 41. Pectin in the cell wall of one-day-old Ranger alfalfa infected plant. X 9200

Figure 42. Pectin in the cell wall of one-day-old Lahontan alfalfa control plant. X 16,000

Figure 43. Pectin in the cell wall of one-day-old Lahontan alfalfa infected plant. X 9200

Figure 44. Pectin in the cell wall of seven-day-old Lahontan alfalfa control plant. X 16,000

Figure 45. Pectin in the cell wall of seven-day-old Lahontan alfalfa infected plant. X 16,000

Figure 46. Pectin in the cell wall of seven-day-old Ranger alfalfa control plant. X 16,000

Figure 47. Pectin in the cell wall of seven-day-old Ranger alfalfa infected plant. X 16,000
Figure 48. One-day-old alfalfa seedling at 15 C. Note the electron dense substances along the tonoplast. X 16,000

Figure 49. Seven-day-old alfalfa seedling at 15 C. Note the electron dense substances in the intercellular space. X 16,000

Figure 50. Seven-day-old infected Ranger alfalfa seedling at 15 C. Note the electron dense spots distributed along the cell wall and the normal nucleus and chloroplast. X 16,000

Figure 51. Seven-day-old Ranger alfalfa seedling at 15 C. Peripheral vesicles in the chloroplast were formed by invagination of the inner membrane of chloroplast (arrow). X 36,000

Figure 52. Seven-day-old Ranger alfalfa control plant at 15 C with normal nucleus, chloroplast, cell wall and cytoplasm. X 16,000

Figure 53. One-day-old Lahontan alfalfa control plant at 15 C with swollen ER in the cytoplasm. X 9200
Figure 54. Three-day-old Lahontan alfalfa control plant at 15 C with normal cell contents. X 16,000

Figure 55. Three-day-old Lahontan alfalfa plant at 15 C showing nematode and host tissue. X 3600

Figure 56. Higher magnification of the host tissue adjacent to the invading nematode in Figure 55. Note the normal chloroplast with phytoferritin, the swollen and broken ER, and the numerous vesicles in the cytoplasm. X 16,000

Figure 57. Three-day-old Lahontan alfalfa infected plant at 15 C with normal chloroplast and lobing nucleus in the cytoplasm. X 9200
Figure 58. Three-day-old Lahontan alfalfa infected plant at 15 C with two normal chloroplasts in adjacent cells. The chloroplast in one cell showed numerous vesicles in the stroma, whereas in the other one swollen ER were evident in the cytoplasm. X 16,000

Figure 59. Three-day-old severely infected Lahontan alfalfa plant at 15 C with broken chloroplast, heavily swollen ER and a nucleus which still contained its nuclear material. X 16,000

Figure 60. Seven-day-old Lahontan alfalfa control plant at 15 C with normal nucleus, cytoplasm, cell wall and chloroplast. Note the slightly swollen ER in the lower left corner. X 16,000

Figure 61. Seven-day-old severely infected Lahontan alfalfa plant at 15 C with swollen chloroplast and ER and nucleus showing less nuclear material. X 16,000

Figure 62. Seven-day-old Lahontan alfalfa control plant at 15 C with long dilated ER in the cytoplasm. X 16,000

Figure 63. Seven-day-old Lahontan alfalfa infected plant at 15 C with active secreting Golgi apparatus and numerous Golgi vesicles. X 16,000
Figure 64. One-day-old Ranger alfalfa control plant at 25 C with normal nucleus, chloroplast and cytoplasm. X 16,000

Figure 65. One-day-old Ranger alfalfa infected plant at 25 C with partially swollen nuclear envelope and wave-like cell wall structure. Note the electron dense spots on the cell wall. X 9200

Figure 66. One-day-old severely infected Ranger alfalfa plant at 25 C with all-round swollen nuclear envelope and electron dense substances. Note the numerous vacuoles in the cytoplasm and the plasmadesmata in the cell wall. X 9200

Figure 67. Glutaraldehyde-OsO₄ combination fixation also showed a swollen nuclear envelope in one-day-old infected Ranger alfalfa plant at 25 C. X 16,000

Figure 68. One-day-old Ranger alfalfa control plant at 25 C. The ER in some cells were heavily swollen. X 9200

Figure 69. One-day-old Ranger alfalfa infected plant at 25 C showing swollen ER. X 16,000
Figure 70. One-day-old alfalfa plant at 25 C with electron dense substances along the vacuoles. X 16,000

Figure 71. One-day-old alfalfa plant at 25 C showing the general pattern of the actively secreting Golgi apparatus in the infected plant. X 24,000

Figure 72. Three-day-old Ranger alfalfa control plant at 25 C with normal cytoplasm and cell wall. X 16,000

Figure 73. Three-day-old severely infected Ranger alfalfa plant at 25 C with round and broken chloroplast and swollen ER. X 16,000

Figure 74. Seven-day-old severely infected Ranger alfalfa plant at 25 C with round and completely disrupted chloroplasts. Note also the swollen mitochondria. X 16,000
Figure 75. Seven-day-old infected Lahontan alfalfa plant at 25 C with round and outer membrane ruptured chloroplast (arrow). X 9200

Figure 76. Seven-day-old heavily infected Lahontan alfalfa plant at 25 C with partially and completely broken chloroplasts. X 16,000

Figure 77. One-day-old Lahontan alfalfa infected plant at 25 C with heavily swollen ER. X 24,000

Figure 78. Vascular region of seven-day-old alfalfa plant at 25 C with normal cell contents. X 3600
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